

Diagnosis of Celiac Disease



Comparative Effectiveness Review

Number 162

Diagnosis of Celiac Disease

Prepared for:

Agency for Healthcare Research and Quality U.S. Department of Health and Human Services 5600 Fishers Lane Rockville, MD 20857 www.ahrq.gov

Contract No. 290-2012-00006-I

Prepared by:

Southern California Evidence-Based Practice Center Santa Monica, CA

Investigators:

Margaret A. Maglione, M.P.P. Adeyemi Okunogbe, M.B.Ch.B. Brett Ewing, M.S. Sean Grant, Ph.D. Sydne J. Newberry, Ph.D. Aneesa Motala, B.A. Roberta Shanman, M.L.S. Nelly Mejia, M.Phil. Aziza Arifkhanova, M.S. Paul Shekelle, M.D., Ph.D. Gregory Harmon, M.D.

AHRQ Publication No. 15(16)-EHC032-EF January 2016

This report is based on research conducted by the Southern California Evidence-based Practice Center (EPC) under contract to the Agency for Healthcare Research and Quality (AHRQ), Rockville, MD (Contract No. 290-2012-00006-I). The findings and conclusions in this document are those of the authors, who are responsible for its contents; the findings and conclusions do not necessarily represent the views of AHRQ. Therefore, no statement in this report should be construed as an official position of AHRQ or of the U.S. Department of Health and Human Services.

None of the investigators have any affiliations or financial involvement that conflicts with the material presented in this report.

The information in this report is intended to help health care decisionmakers—patients and clinicians, health system leaders, and policymakers, among others—make well-informed decisions and thereby improve the quality of health care services. This report is not intended to be a substitute for the application of clinical judgment. Anyone who makes decisions concerning the provision of clinical care should consider this report in the same way as any medical reference and in conjunction with all other pertinent information, i.e., in the context of available resources and circumstances presented by individual patients.

This report is made available to the public under the terms of a licensing agreement between the author and the Agency for Healthcare Research and Quality. This report may be used and reprinted without permission except those copyrighted materials that are clearly noted in the report. Further reproduction of those copyrighted materials is prohibited without the express permission of copyright holders.

AHRQ or U.S. Department of Health and Human Services endorsement of any derivative products that may be developed from this report, such as clinical practice guidelines, other quality enhancement tools, or reimbursement or coverage policies, may not be stated or implied.

This report may periodically be assessed for the currency of conclusions. If an assessment is done, the resulting surveillance report describing the methodology and findings will be found on the Effective Health Care Program Web site at www.effectivehealthcare.ahrq.gov. Search on the title of the report.

Persons using assistive technology may not be able to fully access information in this report. For assistance contact EffectiveHealthCare@ahrq.hhs.gov.

Suggested citation: Maglione MA, Okunogbe A, Ewing B, Grant S, Newberry SJ, Motala A, Shanman R, Mejia N, Arifkhanova A, Shekelle P, Harmon G. Diagnosis of Celiac Disease. Comparative Effectiveness Review No. 162. (Prepared by the Southern California Evidence-based Practice Center under Contract No. 290-2012-00006-I.) AHRQ Publication No. 15(16)-EHC032-EF. Rockville, MD: Agency for Healthcare Research and Quality; January 2016. www.effectivehealthcare.ahrq.gov/reports/final.cfm.

Preface

The Agency for Healthcare Research and Quality (AHRQ), through its Evidence-based Practice Centers (EPCs), sponsors the development of systematic reviews to assist public- and private-sector organizations in their efforts to improve the quality of health care in the United States. These reviews provide comprehensive, science-based information on common, costly medical conditions, and new health care technologies and strategies.

Systematic reviews are the building blocks underlying evidence-based practice; they focus attention on the strength and limits of evidence from research studies about the effectiveness and safety of a clinical intervention. In the context of developing recommendations for practice, systematic reviews can help clarify whether assertions about the value of the intervention are based on strong evidence from clinical studies. For more information about AHRQ EPC systematic reviews, see www.effectivehealthcare.ahrq.gov/reference/purpose.cfm.

AHRQ expects that these systematic reviews will be helpful to health plans, providers, purchasers, government programs, and the health care system as a whole. Transparency and stakeholder input are essential to the Effective Health Care Program. Please visit the Web site (www.effectivehealthcare.ahrq.gov) to see draft research questions and reports or to join an email list to learn about new program products and opportunities for input.

We welcome comments on this systematic review. They may be sent by mail to the Task Order Officer named below at: Agency for Healthcare Research and Quality, 5600 Fishers Lane, Rockville, MD 20857, or by email to epc@ahrq.hhs.gov.

Richard G. Kronick, Ph.D. Director Agency for Healthcare Research and Quality

Stephanie Chang, M.D., M.P.H. Director Evidence-based Practice Center Program Center for Evidence and Practice Improvement Agency for Healthcare Research and Quality Arlene S. Bierman, M.D., M.S. Director, Center for Evidence and Practice Improvement Agency for Healthcare Research and Quality

Karen C. Lee, M.D., M.P.H. Medical Officer, U.S. Preventive Services Task Force Program Task Order Officer Center for Evidence and Practice Improvement Agency for Healthcare Research and Quality

Nahed El-Kassar, M.D., Ph.D. Former Task Order Officer Center for Evidence and Practice Improvement Agency for Healthcare Research and Quality

Acknowledgments

The authors would like to thank Sean Rubin, B.S., and Patricia Smith for their assistance on the project.

Key Informants

In designing the study questions, the EPC consulted several Key Informants who represent the end-users of research. The EPC sought the Key Informant input on the priority areas for research and synthesis. Key Informants are not involved in the analysis of the evidence or the writing of the report. Therefore, in the end, study questions, design, methodological approaches, and/or conclusions do not necessarily represent the views of individual Key Informants.

Key Informants must disclose any financial conflicts of interest greater than \$10,000 and any other relevant business or professional conflicts of interest. Because of their role as end-users, individuals with potential conflicts may be retained. The TOO and the EPC work to balance, manage, or mitigate any conflicts of interest.

The list of Key Informants who provided input to this report follows:

Stefano Guandalini, M.D. University of Chicago Celiac Disease Center Chicago, IL Marilyn Geller, M.S.P.H. Celiac Disease Foundation Woodland Hills, CA

Nancee Jaffee, M.S., R.D. Dietitian, University of California Los Angeles Los Angeles, CA Martin F. Kagnoff, M.D. University of California San Diego Laboratory of Mucosal Immunology San Diego, CA Danna Korn Raising Our Celiac Kids (ROCK) Carlsbad, CA Stephen Levinson, M.D. Gastroenterologist, Community Practice Burbank, CA Joseph Murray, M.D. Gastroenterologist, Mayo Clinic Rochester, MN Mary Schluckebier, M.A. Celiac Sprue Association Omaha, NE

John Whitney, M.D. Wellpoint, Office of Medical Policy Albany, NY

Technical Expert Panel

In designing the study questions and methodology at the outset of this report, the EPC consulted several technical and content experts. Broad expertise and perspectives were sought. Divergent and conflicted opinions are common and perceived as healthy scientific discourse that results in a thoughtful, relevant systematic review. Therefore, in the end, study questions, design, methodologic approaches, and/or conclusions do not necessarily represent the views of individual technical and content experts.

Technical Experts must disclose any financial conflicts of interest greater than \$10,000 and any other relevant business or professional conflicts of interest. Because of their unique clinical or content expertise, individuals with potential conflicts may be retained. The TOO and the EPC work to balance, manage, or mitigate any potential conflicts of interest identified.

The list of Technical Experts who provided input to this report follows:Alessio Fasano, M.D.Director, Center for Celiac Research &TreatmentMassGeneral Hospital for ChildrenBoston, MABoston, MAStefano Guandalini, M.D.University of Chicago Celiac Disease CenterChicago, ILMartin F. Kagnoff, M.D.University of California San DiegoLaboratory of Mucosal ImmunologySan Diego, CAMartin Piezak, M

Daniel A. Leffler, M.D., M.S. Director of Research, The Celiac Disease Center at BIDMC Director of Quality Assurance, Division of Gastroenterology Beth Israel Deaconess Medical Center Boston, MA Joseph Murray, M.D. Gastroenterologist, Mayo Clinic Rochester, MN Michelle Pietzak, M.D. Children's Hospital Los Angeles Los Angeles, CA

Peer Reviewers

Prior to publication of the final evidence report, EPCs sought input from independent Peer Reviewers without financial conflicts of interest. However, the conclusions and synthesis of the scientific literature presented in this report do not necessarily represent the views of individual reviewers.

Peer Reviewers must disclose any financial conflicts of interest greater than \$10,000 and any other relevant business or professional conflicts of interest. Because of their unique clinical or content expertise, individuals with potential nonfinancial conflicts may be retained. The TOO and the EPC work to balance, manage, or mitigate any potential nonfinancial conflicts of interest identified.

The list of Peer Reviewers follows:

Manish J. Gandhi, M.D. Mayo Clinic Rochester, MN Benjamin Lebwohl, M.D., M.S. Celiac Disease Center at Columbia University, New York, NY Melissa Snyder, Ph.D. Mayo Clinic Rochester, MN

Ritu Verma, M.D. Attending Physician/Director of Celiac Center at the Children's Hospital of Philadelphia Philadelphia, PA

Diagnosis of Celiac Disease

Structured Abstract

Objectives. To report the evidence on comparative accuracy and safety of methods used in current clinical practice to diagnose celiac disease, including serological tests, human leukocyte antigen (HLA) typing, and video capsule endoscopy. Diagnostic tests used singly and in combination in various populations were compared against the reference standard of endoscopic duodenal biopsy. In addition, factors affecting biopsy accuracy were reviewed.

Data sources. Electronic searches of PubMed[®], Embase[®], the Cochrane Library, and Web of Science from 1990 through March 2015. Reference lists of included publications were searched for additional relevant studies, and experts were asked to suggest studies.

Review methods. Studies of diagnostic accuracy were included if all participants underwent the index test and endoscopy with duodenal biopsy as the reference standard. Systematic reviews on accuracy and studies on adverse events associated with testing were included. Standard assessment tools were used to evaluate study risk of bias. Where possible, results of accuracy studies were pooled using meta-analysis. When pooling was not possible, findings were described narratively and presented in tables and figures.

Results. A total of 7,254 titles were identified, from which 60 individual studies and 13 prior systematic reviews were included. The majority of studies were conducted in participants with symptoms. New meta-analyses found high-strength evidence to support excellent accuracy of anti-tissue transglutaminase (tTG) immunoglobulin A (IgA) tests (sensitivity = 92.5%; specificity = 97.9%) and excellent specificity of endomysial antibodies (EmA) IgA tests (sensitivity = 79.0%; specificity = 99.0%), as reported in previous systematic reviews. Promising results were reported for deamidated gliadin peptide antibodies (DGP) IgA tests (sensitivity = 87.8%; specificity = 94.1%) in a recent meta-analysis. Evidence for algorithms using multiple tests was insufficient because of diverse results, low number of studies, and heterogeneity of populations. Evidence was also insufficient for accuracy in asymptomatic general population screening and special populations such as children and patients with type 1 diabetes, anemia, and IgA deficiency.

Conclusions. New evidence on accuracy of tests used to diagnose celiac disease supports the excellent sensitivity of tTG IgA tests and excellent specificity of both tTG IgA and EmA IgA tests. Sensitivity of DGP IgA and immunoglobulin G tests is slightly less than for tTG IgA. Additional studies are needed to confirm the accuracy of diagnostic tests in special populations and to validate promising algorithms.

Contents

Executive Summary	ES-1
Introduction	
Background	1
Condition	
Diagnostic Strategies	
Scope and Key Questions	
Scope of the Review	
Key Questions	
Organization of This Report	
Methods	
Topic Refinement and Review Protocol	2
Literature Search Strategy	2
Inclusion and Exclusion Criteria	
Study Selection	4
Data Extraction	4
Quality (Risk of Bias) Assessment of Individual Studies	
Statistical Analyses	6
Strength of the Body of Evidence	7
Applicability	
Peer Review and Public Commentary	
Results	
Results of Literature Searches	10
Key Question 1. Comparative Effectiveness	
Description of Included Studies	
Key Points	26
Detailed Synthesis	
Key Question 2. Duodenal Biopsy Issues	
Key Points	40
Detailed Synthesis	
Key Question 3. Specific Populations	
Key Points	
Detailed Synthesis	
Key Question 4. Adverse Events	50
Key Points	50
Detailed Synthesis	51
Discussion	
Key Findings and Strength of Evidence	
Findings in Relationship to What Is Already Known	
Applicability	
Implications for Clinical and Policy Decisionmaking	
Limitations of the Comparative Effectiveness Review Process	
Limitations of the Comparative Effectiveness Review Process Limitations of the Evidence Base	
Limitations of the Comparative Effectiveness Review Process Limitations of the Evidence Base Volume	

Reporting Quality	
Research Gaps	
Conclusions	
References	66
Abbreviations/Acronyms	70
Tables	
Table A. Summary of findings and strength of evidence	5
Table 1. Literature search methods	2
Table 2. Quality Assessment of Diagnostic Accuracy Studies (QUADAS)-2 questions for	
assessing risk of bias in diagnostic accuracy studies	4
Table 3. AMSTAR (A Measurement Tool to Assess Systematic Reviews) criteria for assessir	ıg
quality of systematic reviews	5
Table 4. McMaster Quality Assessment Scale for Harms (McHarm)	5
Table 5. Strength of evidence definitions	7
Table 6. Domains and their definitions	7
Table 7. Accuracy studies published after systematic reviews: characteristics	13
Table 8. Systematic reviews of tTG tests	25
Table 9. Accuracy of tTG IgA tests	27
Table 10. Systematic reviews of EmA IgA tests	31
Table 11. Accuracy of EmA IgA tests in studies published after NICE and ESPGHAN	
systematic reviews	32
Table 12. Systematic reviews of DGP tests	35
Table 13. Accuracy of algorithms	37
Table 14. Video capsule endoscopy	39
Table 15. Diagnosis by duodenal biopsy: Variation by pathologist and setting characteristics.	42
Table 16. Length of gluten challenge	45
Table 17. Accuracy data for persons with iron deficiency	47
Table 18. Accuracy data for persons with type 1 diabetes	48
Table 19. Accuracy results by age	49
Table 20. Adverse events, video capsule endoscopy used for celiac disease diagnosis	53
Table 21. Quality of adverse events studies.	54
Table 22. Summary of findings and strength of evidence	56

Figures

0	
Figure A. Analytic framework, diagnosis of celiac disease	3
Figure B. Literature flow	4
Figure 1. Analytic framework, diagnosis of celiac disease	4
Figure 2. Literature flow	11
Figure 3. Sensitivity and specificity results for tissue transglutaminase immunoglobulin A test	ts27
Figure 4. Accuracy by threshold level for tissue transglutaminase immunoglobulin A	30
Figure 5. Accuracy of endomysial antibodies immunoglobulin A studies published after NICE	Ξ
and ESPGHAN systematic reviews	32
-	

Appendixes

- Appendix A. Search Strategy
- Appendix B. List of Excluded Studies
- Appendix C. Evidence Table
- Appendix D. Data Abstraction Tools
- Appendix E. AMSTAR Criteria
- Appendix F. Strength of Evidence for Accuracy of Serology Tests

Executive Summary

Background

Condition

Celiac disease (CD) is an immune-mediated disorder triggered in genetically susceptible individuals by ingestion of foods containing gluten, a family of proteins found in wheat, rye, barley, and related grains.¹ The prevalence of CD in the United States has been estimated at approximately 1 percent² but appears to be increasing for reasons that are not clear.³ Risk factors for CD include family history, trisomy 21, Turner syndrome, and Williams syndrome, as well as several autoimmune diseases.

Clinical signs of CD include weight loss, iron deficiency anemia, aphthous ulcers, osteomalacia, dermatitis herpetiformis (a rash due to gluten sensitivity), and gastrointestinal (GI) symptoms, including diarrhea and abdominal bloating. The diagnosis of CD can be challenging because the clinical spectrum of the disease varies, and some individuals present with mild symptoms.⁴

CD causes enteropathy of the small intestine, resulting in poor absorption of nutrients. Malabsorption may result in several of the clinical signs, including iron deficiency anemia, osteomalacia, and weight loss. Young children, in particular, are susceptible to failure to thrive, stunted growth, and delayed puberty.⁵ In women, folate deficiency secondary to CD may lead to poor birth outcomes, including developmental disorders. In the long term, untreated CD increases the risk for non-Hodgkin's lymphoma, certain GI cancers, and all-cause mortality.⁴

The only effective treatment for CD is avoidance of gluten in the diet. Timely diagnosis may be the most important component in the management of CD.

Diagnostic Strategies

A number of diagnostic methods have been developed; the validity and acceptability of some of these methods, particularly newer tests, which include combination tests and algorithms, remain controversial. These methods include various serology tests—anti-gliadin antibodies (AGA), anti-tissue transglutaminase (tTG), endomysial antibodies (EmA), and deamidated gliadin peptide (DGP) antibodies—as well as human leukocyte antigen (HLA) typing, video capsule endoscopy (VCE), and endoscopic duodenal biopsy (often considered the gold standard). Providers may use these tests sequentially in order to increase specificity and prevent false positives, or to increase sensitivity and prevent false negatives. All methods other than HLA typing require the patient to maintain a gluten-containing diet during the diagnostic process.

AGA, immunoglobulin A (IgA) and immunoglobulin G (IgG). Gliadin is one of the two groups of proteins that constitute gluten. AGA determination was used as a diagnostic tool in the 1990s, as it has high sensitivity for CD,⁶ although the test has low specificity. As AGA tests are no longer recommended,^{7,8} they are not addressed in this systematic review.

TTG, IgA. Tissue transglutaminase is an enzyme that causes the crosslinking of certain proteins. Anti-tTG IgA is the single test preferred by the American College of Gastroenterology (ACG) for the detection of CD in those 2 years of age and over⁵ and is included in the algorithms of all recent guidelines. However, as IgA deficiency is more prevalent in CD patients than in the general population, other tests may be ordered as an alternative in those who are IgA deficient.

EmA, IgA. When the intestinal lining is damaged, endomysial antibodies develop. Most patients with active CD and many with dermatitis herpetiformis have the IgA class of anti-EmA antibodies. This test is included in some algorithms of recent guidelines for diagnosis, although it is not as widely used in the United States as in other countries. This test is less useful in IgA-deficient individuals.

DGP antibodies. This is a newer test that may give a positive result in some individuals with CD who are anti-tTG negative, including children under age 2.

HLA typing. Susceptibility to CD is linked to certain HLA class II alleles, especially in the HLA-DQ region. Approximately 95 percent of patients with CD have the HLA-DQ2 heterodimer, while the remaining 5 percent have the HLA-DQ8 heterodimer.⁹ Lack of these heterodimers all but rules out CD and genetic susceptibility for the disorder. These genetic tests are part of the diagnostic algorithms recommended by the European Society for Pediatric Gastroenterology, Hepatology, and Nutrition (ESPGHAN) and the ACG.¹⁰

VCE. For this test, the patient ingests a capsule containing a tiny camera, providing highquality visual evidence of the villous atrophy associated with CD. While not a traditional means of detecting CD, VCE is used in adults who seek to avoid biopsy. During the topic refinement phase of this project, Key Informants suggested that assessment of the evidence for this method be included in this report.

Endoscopic duodenal biopsy. Villous atrophy present on a duodenal biopsy and clinical remission when a gluten-free diet is followed represent the internationally accepted gold standard for CD diagnosis. However, this procedure may be difficult to execute effectively, and some patients and parents of small children are concerned about the possibility of adverse events, including perforations, bleeding, pain, and discomfort.

Scope and Key Questions

Scope of the Review

The purpose of this review is to assess the evidence on the comparative accuracy and possible harms of methods used for the diagnosis of CD, including serological tests, HLA typing, VCE, and endoscopic duodenal biopsy. The review compares the effectiveness of these diagnostic tests singly and in combination in various populations of special interest to the CD community. A protocol for the review was posted online by the Agency for Healthcare Research and Quality (AHRQ) Effective Health Care Program.

Key Questions

Figure A shows an analytic framework to illustrate the populations, interventions, outcomes, and possible adverse effects that guided the literature search and synthesis for this project.

Figure A. Analytic framework, diagnosis of celiac disease



CD = celiac disease; FN = false negative; FP = false positive; IgA = immunoglobulin A; KQ = Key Question; LR+ = positive likelihood ratio; LR- = negative likelihood ratio; SES = socioeconomic status; TN = true negative; TP = true positive.

The Key Questions addressed in this review are as follows:

Key Question 1. What is the comparative effectiveness of the different diagnostic methods (various serological tests, human leukocyte antigen [HLA] typing, video capsule endoscopy, used individually and in combination) compared with endoscopy with biopsy as the reference standard, to diagnose celiac disease (CD) in terms of—

- a. Accuracy: sensitivity, specificity, positive likelihood ratio, negative likelihood ratio, and summary receiver-operating characteristics?
- b. Intermediate outcomes, such as clinical decisionmaking and dietary compliance?
- c. Clinical outcomes and complications related to CD?
- d. Patient-centered outcomes, such as quality of life (QOL) and symptoms?

Key Question 2. Do accuracy/reliability of endoscopy with duodenal biopsy vary by—

- a. Pathologist characteristics (i.e., level of experience or specific training)?
- b. Method (i.e., type or number of specimens)?
- c. Length of time ingesting gluten before diagnostic testing?

Key Question 3. How do accuracy and outcomes differ among specific populations, such as—

a. Symptomatic patients versus nonsymptomatic individuals at risk?

- b. Adults (age 18 and over) versus children and adolescents?
- c. Children under age 24 months versus older children?
- d. Demographics, including race, genetics, geography, and socioeconomic status?
- e. Patients with IgA deficiency?
- f. Patients previously testing negative for CD?

Key Question 4. What are the direct adverse effects (e.g., bleeding from biopsy) or harms (related to false positives, false negatives, indeterminate results) associated with testing for CD?

Methods

Topic Refinement and Review Protocol

Key Informants from professional associations, research centers, payers, and patient organizations were engaged to assist in refining the Key Questions (KQs) and issues to cover in this systematic review. The authors then refined and finalized the KQs after review of public comments collected on the AHRQ Effective Health Care Web site in February 2014. The final protocol was posted on the Web site in June 2014 after input from a Technical Expert Panel representing various areas of expertise in CD.

Literature Search Strategy

An experienced reference librarian designed the search strategies in collaboration with an expert on CD and project staff experienced in systematic review methods. The search strategy included search terms for CD, combined with general terms for diagnosis or terms representing each diagnostic method, plus terms representing all outcomes listed in the PICOTs (populations, interventions, comparators, outcomes, timing, and setting). The full search strategy is presented in Appendix A of the full report.

For KQ 1a, we searched for publications starting from January 1990 but did not abstract studies that were already included in recent high-quality systematic reviews. For KQ 2, on duodenal biopsy, and KQ 3, on specific populations, our search also started at January 1990. For KQ 4, on direct and indirect harms of the diagnostic procedures, our search started at January 2003, as this KQ was covered by an AHRQ-funded systematic review published in 2004.¹¹

PubMed[®], Embase[®], the Cochrane Library, and Web of Science were searched. The AHRQ-funded Scientific Resource Center requested unpublished data from manufacturers of all serological tests. Key Informants, project clinicians, and members of the Technical Expert Panel also suggested studies. Reference lists of included articles were reviewed for identification of additional relevant studies.

Inclusion and Exclusion Criteria

Eligible studies of diagnostic accuracy included controlled trials, prospective and retrospective cohorts, case-control studies, and case series. Studies were included if they met the following criteria:

- Diagnostic method must be currently used in clinical practice, as listed in the PICOTS. Diagnostic methods no longer recommended or still in development were excluded.
- Study was about diagnosis of CD rather than management of existing CD.

- All participants underwent both the "index test" and the reference standard (biopsy).
- The study reported sensitivity, specificity, or data that allowed calculation.
- Study was published in English.
- Study enrolled a consecutive or random sample.
- For representativeness and generalizability, the sample size was 300 or more unless one of the following populations of interest was the focus:
 - Low socioeconomic status
 - Previously negative for CD via serology or biopsy
 - IgA deficient
 - Type 1 diabetes
 - Turner syndrome
 - Trisomy 21/Down syndrome
 - o Iron deficiency anemia
 - Family history
- Accuracy results were stratified by race/ethnicity.

The following were excluded from this systematic review:

- Animal studies
- Individual case reports
- Studies not published in English
- Documents with no original data (commentary, editorial)
- Studies that reported only prevalence

The PICOTS considered in this review are as follows.

Population(s):

For KQs 1, 2, and 4—

All populations tested for CD

For KQ 3—

- Patients with signs and symptoms of CD; for example
 - o Diarrhea
 - Constipation
 - o Dermatitis
 - o Malabsorption (anemia, folate deficiency)
- Asymptomatic individuals at risk of CD because of—
 - Family history
 - Type 1 diabetes
 - Autoimmune disease
 - o Turner syndrome
 - o Trisomy 21
- Children under age 24 months versus older children and adolescents
- Adults (aged 18 and over)
- Ethnic and geographic populations
- Patients with low socioeconomic status
- Patients with IgA deficiency

• Patients previously testing negative for CD

Interventions:

For KQs 1, 3, 4—

- Test for EmA IgA
- Test for tTG IgA
- Test for DGP IgA antibodies
- EmA IgG, tTG IgG, and DGP IgG tests for IgA-deficient individuals
- HLA typing
- VCE
- Combinations of the above

For KQ 2—

• Endoscopy with biopsy

Comparators:

For KQs 1 and 3—

- Endoscopy with duodenal biopsy
- For KQ 2—
- Repeat biopsy

Outcomes:

For KQ 1a, KQ 2, and KQs 3a-f, for accuracy-

- Sensitivity
- Specificity
- Positive predictive value, negative predictive value, false positive, false negative
- Positive and negative likelihood ratios
- For KQ 1b, for clinical decisionmaking—
- Additional testing for CD
- Nutritionist advice on gluten-free diet
- Followup and monitoring by physician

For KQ 1c, for clinical outcomes and complications-

- Nutritional deficits
- Persistence of villous atrophy on biopsy
- Lymphomas
- For KQ 1d, for patient-centered outcomes—
- QOL
- Discomfort
- Bloating
- Abdominal pain
- Depression

For KQ 4, for harms—

- Immediate adverse events from biopsy
- Psychological stress related to false positive results
- Sequelae of false negatives or indeterminate results

Timing:

For KQ 2—

• Length of time ingesting gluten before biopsy

Setting:

For all KQs—

- Outpatient: academic
- Outpatient: community

Study Selection

Each title and abstract identified by the searches was screened independently by two researchers, and the combination of their selections was retrieved for full-text review. Two researchers independently screened each full-text article for inclusion in the project, with a senior researcher resolving discrepancies. A list of excluded studies with reasons for exclusion is presented as Appendix B of the full report.

Data Extraction

The DistillerSR software package was used to manage the search output, screening, and data abstraction. Data collection forms were designed by the project team in DistillerSR, piloted by the reviewers, and further modified; then the final forms were piloted with a random selection of included studies to ensure agreement of interpretation. Articles accepted for inclusion were abstracted in DistillerSR; a statistical analyst abstracted accuracy data in Excel. The project leader reviewed data for all included studies for accuracy and made revisions accordingly. Forms are displayed in Appendix D of the full report.

Quality (Risk-of-Bias) Assessment of Individual Studies

The QUADAS-2¹² instrument (revised Quality Assessment of Diagnostic Accuracy Studies instrument) was used to assess the risk of bias of accuracy studies; the McHarm instrument¹³ was used to assess the quality of studies on adverse events; and the AMSTAR¹⁴ instrument (a measurement tool for the assessment of multiple systematic reviews) was used to assess the quality of prior systematic reviews. These instruments are described in detail in the Methods chapter of the full report. Each study was scored individually by two Evidence-based Practice Center researchers, who met to reconcile any differences; the project leader resolved discrepancies.

Diagnostic Accuracy—Statistical Analyses

Studies that reported sensitivity, specificity, or ROCs, or provided the data to calculate these values, were abstracted for potential inclusion in a synthesis. Sensitivity is also known as the "true positive rate," the ability of a test to correctly classify an individual as having a condition in this case, having CD as confirmed by biopsy. Sensitivity ranges from 0 to 100, with values closer to 100 indicating a greater probability of a test being positive when the disease is present.¹⁵ Specificity, also known as the "true negative rate," is the ability of a test to correctly classify an individual as not having a condition—in this case, when the individual is determined by biopsy not to have CD. Specificity ranges from 0 to 100, with values closer to 100 indicating a greater probability of a test being negative when the disease is not present.¹⁵ A perfect diagnostic test would have both sensitivity and specificity of 100 percent. In general, sensitivity and specificity are considered good if at least 70.0 percent, very good from 80.0 percent to 89.9 percent, and excellent if 90.0 percent or greater.¹⁵ Some studies of the accuracy of diagnostic tests report likelihood ratios (LRs), the probability of a positive finding in patients with a disease divided by the probability of the same finding in patients without the disease. Likelihood ratios can range from 0 to infinity. An LR of 1 indicates no change in the likelihood of disease.¹⁶ As the LR increases from 1, the likelihood of disease increases. LR+ (positive likelihood ratio) is a measure of how the probability of the disease increases in the presence of a positive test finding, while LR- (negative likelihood ratio) is a measure of how the probability of the disease decreases if the test is negative. An LR+ of greater than 10 is considered good, as is an LR- of less than 0.1.¹⁷

Finally, positive predictive value (PPV) is the probability that an individual who tests positive actually has the disease. Similarly, negative predictive value (NPV) is the probability of not having a disease when an individual tests negative. Unlike sensitivity and specificity, predictive values (PPV, NPV) are largely dependent on the prevalence of a disease in a study population. With increased prevalence in a population, PPV increases while NPV decreases.

If three or more studies of the same diagnostic method and comparator reported the number of true positives, false positives, true negatives, and false negatives by arm, their results were pooled in order to estimate overall sensitivity, specificity, LRs, and predictive values. Additional analyses were conducted by stratifying by test type, threshold (titer), and population characteristics of interest. When pooling was not possible, study results were described narratively according to comparisons of interest and presented in tables and figures in the full report.

Strength of the Body of Evidence

The overall strength of evidence for accuracy outcomes was assessed using guidance developed by experts in systematic reviews for the AHRQ Effective Health Care Program.¹⁸ This method classifies the strength of evidence based on the following domains: study limitations (risk of bias), consistency, directness, and precision. The domains are described in the Methods chapter of the full report. In this Executive Summary, we report the strength of evidence for each KQ and subquestion. Appendix F in the full report displays the results for each domain for the evidence on accuracy of serological tests in each population.

Applicability

Applicability assessment was based on the similarity of the populations in terms of characteristics listed in the PICOTs.

Peer Review and Public Commentary

A draft version of this report was reviewed by several CD experts; names and affiliations are listed in the front matter of the report. All Peer Reviewers completed conflict-of-interest disclosure forms; none reported ties to any test manufacturers. A draft version of this report was posted on the AHRQ Effective Health Care Web site in February 2015 for public comment. The authors reviewed the comments and incorporated the feedback into the final version.

Results

Overview

Figure B is a literature flow diagram that displays the number of studies identified through electronic searches and contact with experts. It shows the number of studies accepted at each stage of screening and reasons for excluding the others. Table A presents the key findings from prior systematic reviews, results reported in newly identified studies, summary conclusions by KQ and subquestion, and strength of evidence. The applicability and limitations of the evidence are discussed, followed by overall conclusions.

Results of Literature Searches

As displayed in Figure B, of a total of 7,254 titles from the literature search, 60 individual studies and 13 prior systematic reviews (SRs) were included for evidence synthesis. References for the excluded articles, along with reasons for exclusion, can be found in Appendix B of the full report. Thirty-one articles reporting original data and 11 SRs addressed KQ 1 and KQ 3, 25 articles and 1 SR addressed KQ 2, and 4 articles and 1 SR addressed KQ 4.

Figure B. Literature flow



CD = celiac disease; EPC = Evidence-based Practice Center; KQ = Key Question; SR = systematic review.

Key Findings and Strength of Evidence

The key findings and strength of evidence are summarized in Table A. Additional details on strength-of-evidence ratings are provided as Appendix F of the full report.

Торіс	EPC Conclusions and Strength of Evidence	Prior Systematic Reviews	Additional Findings From EPC
Key Question 1: Accuracy of IgA tTG	High: IgA tTG tests have excellent sensitivity and specificity.	A 2010 meta-analysis that pooled 12 studies found a sensitivity of 93.0% (95% CI, 91.2% to 94.5%) and specificity of 96.5% (95% CI, 95.2% to 97.5%). A 2012 meta-analysis restricted to 5 studies of point-of-care tests in children reported sensitivity and specificity of 96.4% (95% CI, 94.3% to 97.9%) and 97.7% (95% CI, 95.8% to 99.0%), respectively.	Sixteen studies were published after the SRs were pooled. Excluding data for threshold levels higher than used in clinical practice, sensitivity was 92.5% (95% CI, 89.7% to 94.6%) and specificity was 97.9% (95% CI, 96.5% to 98.7%). LR+ was 40.19 and LR- was 0.08. PPV was 89.4%, while NPV was 99.0%.
Key Question 1: Accuracy of IgA EmA	High: IgA EmA tests have lower sensitivity but equal specificity to IgA tTG tests.	A 2009 SR including 23 studies found sensitivity ranging from 68% to 100%, while specificity ranged from 77% to 100%; pooling was not performed. A 2012 SR included 11 studies in children; sensitivity ranged from 82.6% to 100% and pooled specificity was 98.2% (95% CI, 96.7% to 99.1%).	Seven studies were published after the SRs were pooled. Sensitivity was 79.0% (95% CI, 71.0% to 86.0%) and specificity was 99.0% (95% CI, 98.4% to 99.4%) after excluding data points where Marsh Grade I and II villous atrophy was classified as CD (not standard practice). LR+ was 65.98 and LR- was 0.21. PPV was 78.9%; NPV was 99.1%.
Key Question 1: Accuracy of IgA DGP	High: IgA DGP tests are not as accurate as IgA tTG tests.	A 2010 SR pooled 11 studies on accuracy in all ages; sensitivity was 87.8% (95% CI, 85.6% to 89.9%), while specificity was 94.1% (95% CI, 95.2% to 97.5%). LR+ was 13.33, while LR- was 0.12. A 2012 SR reviewed 3 of those studies that included only children: sensitivities ranged from 80.7% to 95.1% (not pooled) and pooled specificity was estimated at 90.7% (95% CI, 87.8% to 93.1%).	One new study reported sensitivity of 97.0% and specificity of 90.7% in symptomatic adults and children at 1 clinic, while another reported both sensitivity and specificity of 96% in a similar population.
Key Question 1: Accuracy of IgG DGP	Moderate: IgG DGP tests are not as sensitive as IgA tTG tests in non–IgA-deficient patients.	A 2013 SR of 7 studies of non–IgA-deficient adults reported sensitivity of 75.4% to 96.7% and specificity of 98.5% to 100%. A 2012 SR of 3 studies in non– IgA-deficient children reported sensitivities of 80.1% to 98.6% and specificities of 86.0% to 96.9%. Authors did not pool data.	One study reported sensitivity of 95.0% and specificity of 99.0% in 200 non–IgA-deficient subjects of all ages.
Key Question 1: Accuracy of HLA- DQ2 or DQ8	High: HLA tests can be used to rule out CD with close to 100% sensitivity.	No SRs of the accuracy of testing for HLA-DQ2 or DQ8 were identified. Based on studies from which sensitivity (but not specificity) could be calculated, the American College of Gastroenterology estimated	Two studies were identified on the accuracy of HLA testing. A large 2013 prospective cohort found that HLA testing had a sensitivity of 100% and specificity of 18.2%.

Table A. Summary of findings and strength of evidence

		the NPV of the HLA-DQ2/DQ8 combination test at over 99%.	A 1999 cohort also reported sensitivity of 100%, while specificity was 33.3%.
Key Question 1: Accuracy of algorithms	Insufficient: Strength of evidence is insufficient to determine comparative accuracy of different algorithms in specific populations.	No SRs of the accuracy of algorithms were identified.	Nine studies of algorithms were identified; all used tTG tests. Adding an EmA test to a tTG test resulted in increased specificity, with either no change or a slight decrease in sensitivity. Adding a DGP test to a tTG test resulted in increased sensitivity but decreased specificity. However, the increase in accuracy compared with individual tests was rarely clinically significant. The sensitivity and specificity results varied widely, populations were diverse, and the evidence base had high heterogeneity.
Key Question 1: Accuracy of VCE	Moderate: VCE has very good sensitivity and excellent specificity.	A previous SR of moderate quality on the accuracy of VCE pooled 6 studies, and estimated sensitivity at 89.0% (95% CI, 82.0% to 94.0%) and specificity at 95.0% (95% CI, 89.0% to 99.0%). LR+ was 12.90 and LR- was 0.16.	No additional studies met our inclusion criteria.
Key Question 1: Intermediate outcomes	Insufficient: Strength of evidence is insufficient regarding how method of diagnosis affects adherence.	A previous SR of low quality (3 studies) reported no statistical difference in adherence levels between patients diagnosed via screening and those diagnosed because they were symptomatic. Association between diagnostic test type and adherence was not addressed.	In 1 study on blood donors in Israel who tested positive for IgA tTG (or IgG tTG if IgA deficient), only 4 of 10 patients with asymptomatic biopsy-proven CD adhered to a gluten-free diet; the other 6 patients did not believe they had CD, and 4 of those were told by physicians that asymptomatic patients did not need to modify their diets.
Key Question 1: Clinical outcomes and complications	Insufficient: Strength of evidence is insufficient regarding how method of diagnosis affects clinical outcomes and complications.	No prior SRs on this topic were identified.	No studies on this topic were identified.
Key Question 1: Patient- centered outcomes such as quality of life	Insufficient: Strength of evidence is insufficient regarding how method of diagnosis affects patient-centered outcomes such as quality of life.	No prior SRs on this topic were identified.	No studies on this topic were identified.

Key Question 2: Biopsy and provider characteristics	Moderate: Physician adherence to biopsy protocol decreases with volume performed per endoscopy suite and increases with number of gastroenterologists per endoscopy suite.	No SRs on this topic were identified.	One very large high-quality national retrospective study found reduced physician adherence to the American Gastroenterological Association's duodenal biopsy protocol (4+ specimens) with higher procedure volume per endoscopy clinic. The OR for each 100 additional procedures was 0.92 (95% CI, 0.88 to 0.97). Adherence increase for each additional gastroenterologist per endoscopy suite was OR 1.08 (95% CI, 1.04 to 1.13).
Key Question 2: Biopsy and pathologist characteristics	Moderate: CD-related histological findings are underdiagnosed in community settings when compared with academic settings.	No SRs on this topic were identified.	Three retrospective studies reported low interobserver agreement between pathologists in community vs. academic settings, with significantly lower accuracy in community settings. Kappa statistics range from 0.16 to 0.53.
Key Question 2: Biopsy specimens— number and location	High: Increasing the number and location of biopsy specimens increases diagnostic accuracy.	No SRs addressed how the number and location of biopsy specimens influence diagnostic findings of biopsy.	Nineteen studies reported that increasing the number and location of biopsy specimens increased the likelihood of diagnosis and diagnostic yield by 25% to 50% in both pediatric and adult populations.
Key Question 2: Biopsy and length of time ingesting gluten	Moderate: A minimum 2-week gluten intake is necessary to induce intestinal changes necessary for diagnosing adults via duodenal biopsy. Low: A 2–3 month diet containing gluten may be necessary to diagnose CD in children via biopsy; strength is lower due to fewer available studies and inconsistent findings.	A previous SR of high quality on clinical response to gluten challenge indicates that 2 weeks of a moderate to high dose (e.g., 15g daily) is sufficient to cause enough intestinal changes to diagnose adults via duodenal biopsy. This same SR reports that for children, 2 to 3 months may be needed.	One small study reported that 3 grams of gluten per day for 2 weeks induces intestinal atrophy sufficient to diagnose CD in 89.5% of adults.
Key Question 3: Symptomatic patients vs. nonsymptomatic individuals at risk	High: EmA and tTG tests have excellent sensitivity and specificity in patients with GI symptoms. Insufficient: How accuracy of serological tests differs between patients with risk factors such as iron deficiency or type 1 diabetes and the	A 2010 SR including only studies of patients with GI symptoms reported pooled sensitivity of 90% (95% CI, 80.0% to 95.0%) and specificity of 99% (95% CI, 98.0% to 100.0%) for IgA EmA tests (8 studies), and pooled sensitivity of 89% (95% CI, 82.0% to 94.0%) and specificity of 98% (95% CI, 95.0% to 99.0%) for IgA tTG tests. No SRs were identified that compared test accuracy in patients with specific symptoms and asymptomatic individuals at risk.	One high-quality study compared the accuracy of the ESPGHAN algorithm (combining tTG IgA and EmA IgA) among subjects with family history, type 1 diabetes, and CD symptoms. Specificity was much higher in those with symptoms. Two small studies provided data that allowed calculation of accuracy in patients with iron deficiency, and 2 provided accuracy data for patients with type 1 diabetes. However, the studies were conducted in the Middle East and Eastern Europe; applicability to the United States is uncertain.

	general symptomatic population could not be determined.		
Key Question 3: Children vs. adults	Low: tTG and DGP tests are less sensitive in adults than children. DGP is more accurate than tTG in children under age 24 months.	No SRs assessing how test accuracy differs by age were identified. Regarding IgG DGP, one SR reported only on studies of adults, while another reported only on studies of children. A 2013 SR of 7 studies of non–IgA-deficient adults reported sensitivity of 75.4% to 96.7% and specificity of 98.5% to 100%. A 2012 SR of 3 studies in non–IgA-deficient children reported sensitivities of 80.1% to 98.6% and specificities of 86.0% to 96.9%.	Two large moderate-quality studies reported that both tTG and DGP tests were less sensitive in adults (range, 29% to 85%) than children (range, 57% to 96%). One study reported sensitivity of 96% and 100% for IgA tTG and IgA DGP, respectively, for children under age 24 months, while specificity was 98% and 31%, respectively. Accuracy was significantly lower for both tests in older children and adolescents.
Key Question 3: Demographics, including race	Insufficient: There was insufficient evidence to estimate the accuracy of diagnostic methods by demographic characteristics.	No SRs on this topic were identified.	No studies reported accuracy by race, ethnicity, or socioeconomic status.
Key Question 3: Patients with IgA deficiency	Insufficient: There was insufficient evidence to estimate the accuracy of diagnostic methods in IgA-deficient patients.	No SRs on this topic were identified.	Two small studies of the accuracy of new combination tests (IgA DGP + IgG DGP combo, IgA tTG + IgG DGP combo) in IgA-deficient patients were published in 2014; results were inconsistent.
Key Question 3: Patients who previously tested negative for CD	Insufficient: There was insufficient evidence to estimate the accuracy of diagnostic methods in patients who previously tested negative for CD.	No SRs on this topic were identified.	A very small study (N = 17) found that patients with biopsy-verified CD who tested negative on IgA tested positive using IgA DGP or IgG DGP.
Key Question 4: Direct adverse events—VCE	High: The rate of capsule retention is less than 5%.	No SRs contained safety data on VCE used specifically for CD diagnosis. An SR of VCE not specific to CD found a capsule retention rate of 1.4% in 150 studies.	In 3 studies specific to CD, the capsule retention rate ranged from 0.9% to 4.6%.
Key Question 4: Direct adverse events— endoscopy with duodenal biopsy	Moderate: Adverse events during upper GI endoscopy are rare.	No SR contained safety data on upper GI endoscopy or duodenal biopsy when used specifically to diagnose CD. A review on upper endoscopy in general found infection very rare and bleeding very rare (1.6 per 1,000) unless a polyp is removed.	No studies specific to diagnosis of CD were identified.
Key Question 4: Indirect adverse events—false negatives or	Insufficient: Strength of evidence is insufficient regarding the impact of misdiagnosis.	No SRs on the impact of misdiagnosis of CD were identified.	In 2 small studies reporting sequelae in children with positive EmA serology but normal biopsy results, 30% to 50% of patients were diagnosed with CD after gluten challenge. These studies were conducted prior

positives	to the availability of other serological tests, so
	applicability is limited.
	A study of 34 children with intestinal villous atrophy
	and simultaneous negative EmA IgA tests found that
	2 infants were confirmed as having CD after 6–10
	years of iterative cycles of gluten challenges and
	gluten-free diet. All 3 studies report high loss to
	followup.

CD = celiac disease; CI = confidence interval; DGP = deamidated gliadin peptide; EmA = endomysial antibodies; EPC = Evidence-based Practice Center; GI = gastrointestinal; ESPGHAN = European Society for Pediatric Gastroenterology, Hepatology, and Nutrition; HLA = human leukocyte antigen; IgA = immunoglobulin A; IgG = immunoglobulin G; LR- = negative likelihood ratio; LR+ = positive likelihood ratio; NPV = negative predictive value; OR = odds ratio; PPV = positive predictive value; SR = systematic review; tTG = anti-tissue transglutaminase; VCE= video capsule endoscopy.

Applicability

Several factors affect the applicability of this review.

To increase generalizability, this report limited inclusion of accuracy studies to those that enrolled consecutive patients or a random sample. Several studies were excluded because enrollment could not be determined given the information available.

Only one study of accuracy in the asymptomatic general population met the criterion that all subjects, regardless of serology results, undergo biopsy. The cost of performing biopsies in all subjects and the low rate of acceptance of biopsy in seronegative asymptomatic individuals make the conduct of such studies challenging. Thus, the evidence on accuracy of diagnosis in the general asymptomatic population with no risk factors for CD is categorized as low strength.

Although this report is limited to diagnostic methods currently used in the United States, study location was not a basis for study exclusion. Many studies were conducted in Europe, the Middle East, and South Asia. Due to differences in genetics and disease prevalence, the applicability of these studies to the U.S. population is uncertain.

No studies stratified accuracy results by racial or ethnic group. Few studies focused on populations of special interest.

Most studies were conducted by gastroenterologists in academic settings. This report found a significant difference in interpretation of biopsy results between academic and nonacademic physicians. The majority of accuracy studies included in this report used Marsh classification to categorize biopsy results. (Marsh III or higher is classified as CD.) In contrast, many community physicians use a simple qualitative assessment of villous atrophy or elevation of intraepithelial lymphocytes to make a diagnosis.

Accuracy of serology assays may vary by both laboratory and manufacturer. For example, Li and colleagues (2009)¹⁹ used 150 samples from subjects of known CD status to compare accuracy of tTG tests at 20 laboratories in the United States and Europe. Sensitivity was less than 75 percent at four laboratories. Rozenberg and colleagues (2012)²⁰ found differences in performance of tTG tests across various manufacturers by using a similar research design.

Finally, VCE is not a first-line diagnostic method: it is indicated for adults who refuse biopsy. A 2012 systematic review of six studies reported very good sensitivity and excellent specificity with VCE. However, there may be differences in patient characteristics between those who refuse and those who accept a biopsy. For example, those with more severe symptoms are hypothesized to be more likely to accept a biopsy.

Implications for Clinical and Policy Decisionmaking

The findings of this review support those of previous SRs on the accuracy of individual diagnostic tests using IgA. All IgA tests for CD have excellent specificity; DGP IgA has slightly lower specificity than tTG IgA and EmA IgA. Testing for tTG IgA has a high PPV for most clinical populations with a modest prevalence of CD. EmA IgA has good sensitivity, DGP IgA has very good sensitivity, and tTG IgA has excellent sensitivity. DGP IgG tests have very good sensitivity and excellent specificity, even in non–IgA-deficient individuals.

Unfortunately, due to a dearth of studies meeting our inclusion criteria, we were unable to determine which tests, if any, are more accurate in patients with specific symptoms or risk factors. Patients with symptoms associated with CD would impact the pretest probability and, as a result, the likelihood of disease based on a positive result. No studies of test accuracy in patients with trisomy 21, Turner syndrome, or Williams syndrome were identified. The few

studies of patients with type 1 diabetes included small samples and were conducted in non-Western countries. Thus, no clinical implications for testing individuals with specific risks can be stated at this time.

New research has found DGP tests to be more accurate than tTG tests in small children; strength of evidence is low but could increase if findings are replicated. Compared with EmA IgA, tTG IgA had greater sensitivity in the one study of the general asymptomatic population identified that met our inclusion criteria that all participants undergo biopsy, regardless of serology results. The quality of this general population study was high, the sample size was large (over 1,000), and it was conducted in a Western country (Sweden) with estimated CD prevalence similar to that in the United States.

This review found insufficient evidence to determine which populations would most benefit from diagnostic algorithms that combine a tTG test with an EmA or DGP test. A combination of positive serological testing with a threshold level at or several times above the upper limit of normal for specific celiac tests may be accurate for diagnosing CD without requiring histopathology specimens. However, the currently available evidence on comparative accuracy of algorithms is inconclusive because of the wide range of results, heterogeneity of populations studied, and lack of clinically significant increases in accuracy compared with individual tests. Future studies aimed at the diagnostic accuracy of multiple-test strategies would strengthen the evidence for this approach.

Finally, regarding biopsy, there is high-strength evidence that multiple specimens should be taken from the duodenal bulb and the distal duodenum for optimal diagnostic yield in both the adult and pediatric population. There is moderate-strength evidence that CD is underdiagnosed by pathologists in community settings compared with academic settings; continued education on diagnostic protocols may be warranted for community physicians.

Research Gaps

Although the accuracy of various serological tests for CD in symptomatic individuals has a high strength of evidence, strength of evidence on the comparative accuracy of algorithms such as those recommended by organizations such as ESPGHAN is insufficient because of the small number of studies, heterogeneity of study populations, and inconsistent results. Further studies should be conducted. Appendix F of the full report contains details on the test combinations, populations, and strength–of-evidence domains for each algorithm studied.

Evidence is insufficient to recommend specific tests for particular at-risk populations. Patient-level factors that have been hypothesized to affect test accuracy include race and ethnicity, but no studies stratified results by these characteristics.

Because of the inherently invasive nature of biopsy, the vast majority of studies of serological test accuracy using biopsy as the reference standard have been conducted in patients presenting for testing due to symptoms. The most common symptoms are GI symptoms (diarrhea, constipation, pain, etc.) as well as signs of malnutrition in children. High accuracy was found in the only general population screening study; however, despite the high scientific quality of this study, the strength of evidence for accuracy in the asymptomatic general population is low because the study has never been replicated. This lack of evidence does not mean the tests are inaccurate in asymptomatic individuals; lack of evidence does not equal evidence of inaccuracy.

No studies were identified that addressed the key issue, "What impact does the method of initial diagnosis have on how a physician follows up with a patient?" Retrospective analyses of existing databases may shed light on this question.

Finally, studies may be needed to investigate the long-term impact of misdiagnosis. False positives and false negatives may be important "harms" because of (a) huge lifestyle changes involved for positive diagnosis and (b) potential harms to health (malabsorption, intestinal damage) from undiagnosed CD.

Conclusions

New evidence on accuracy of tests used to diagnose CD supports the excellent sensitivity of IgA tTG tests and excellent specificity of both IgA tTG and IgA EmA tests reported in prior SRs. High strength of evidence of accuracy, particularly in children, was found for DGP tests in recent SRs. Regarding comparative accuracy, IgA EmA tests have lower sensitivity but similar specificity to IgA tTG tests. IgA DGP and IgG DGP tests are not as sensitive as IgA tTG tests in non–IgA-deficient adults. These conclusions are based primarily on indirect evidence—i.e., pooled results on accuracy of individual tests rather than head-to-head studies comparing accuracy of different tests in the same samples. However, strength of evidence is high given the large numbers of studies, the consistency of results, and the precision of the confidence intervals.

Algorithms combining tTG with either EmA or DGP tests appear to be accurate in both children and adults; however, strength of evidence for comparative accuracy is insufficient given the low number of studies relative to single tests, heterogeneity of populations, and wide range of results. The increase in accuracy over individual tests is not consistently clinically significant. Additional studies of algorithms are needed.

Notably, current ESPGHAN guidelines state that a patient with a tTG result greater than 10 times the normal limit should undergo an EmA test and HLA typing. If the patient tests positive and then responds to a gluten-exclusion diet, a diagnosis of CD can be made without use of biopsy. These guidelines have not been adopted by societies in the United States. Evidence seems to support the accuracy of a multiple-testing strategy without biopsy; however, additional studies are needed to confirm the threshold levels that provide the highest accuracy and population differences, if any.

VCE is a safe and fairly accurate means of diagnosing CD in adults who wish to avoid biopsy; risk of retaining the capsule is approximately 4.6 percent. However, our pooled results reveal that some serological tests have higher sensitivity and specificity. No data are available on how VCE accuracy varies by population characteristics or setting. Endoscopy with biopsy has a very low risk of adverse events; accuracy appears to be greater in academic than community settings.

Importantly, few applicable studies on the sequelae of false positive or false negative diagnoses were identified. Long-term followup of patients, regardless of diagnosis results, should be encouraged.

References

- Rashtak S, Murray JA. Review article: coeliac disease, new approaches to therapy. Aliment Pharmacol Ther. 2012 Apr;35(7):768-81. PMID: 22324389.
- Rubio-Tapia A, Ludvigsson JF, Brantner TL, et al. The prevalence of celiac disease in the United States. Am J Gastroenterol. 2012 Oct;107(10):1538-44; quiz 7, 45. PMID: 22850429.
- Ludvigsson JF, Rubio-Tapia A, van Dyke CT, et al. Increasing incidence of celiac disease in a North American population. Am J Gastroenterol. 2013 May;108(5):818-24. PMID: 23511460.
- See J, Murray JA. Gluten-free diet: the medical and nutrition management of celiac disease. Nutr Clin Pract. 2006 Feb;21(1): 1-15. PMID: 16439765.
- Rubio-Tapia A, Hill ID, Kelly CP, et al. ACG clinical guidelines: diagnosis and management of celiac disease. Am J Gastroenterol. 2013 May;108(5):656-76; quiz 77. PMID: 23609613.
- Bottaro G, Rotolo N, Spina M, et al. [Evaluation of sensitivity and specificity of antigliadin antibodies for the diagnosis of celiac disease in childhood]. Minerva Pediatr. 1995 Dec;47(12):505-10. PMID: 8900559.
- Bai J, Zeballos E, Fried M, et al. WGO-OMGE Practice Guideline: Celiac Disease. World Gastroenterology Organisation; 2007.
- National Institute for Health and Clinical Excellence. Coeliac Disease: Recognition and Assessment of Coeliac Disease. London; National Institute for Health and Clinical Excellence; 2009 May. www.org.uk/guidance.
- Rostom A, Murray JA, Kagnoff MF. American Gastroenterological Association (AGA) Institute technical review on the diagnosis and management of celiac disease. Gastroenterology. 2006 Dec;131(6):1981-2002. PMID: 17087937.

- Husby S, Koletzko S, Korponay-Szabo IR, et al. European Society for Pediatric Gastroenterology, Hepatology, and Nutrition guidelines for the diagnosis of coeliac disease. J Pediatr Gastroenterol Nutr. 2012 Jan;54(1):136-60. PMID: 22197856.
- Rostom A, Dube C, Cranney A, et al. Celiac disease. Evid Rep Technol Assess (Summ). 2004 Jun;(104):1-6. PMID: 15346868.
- 12. Whiting PF, Rutjes AW, Westwood ME, et al. QUADAS-2: a revised tool for the quality assessment of diagnostic accuracy studies. Ann Intern Med. 2011 Oct 18;155(8):529-36. PMID: 22007046.
- Chou R, Aronson N, Atkins D, et al. Chapter 11. Assessing harms when comparing medical interventions. 2009. In: Methods Guide for Effectiveness and Comparative Effectiveness Reviews. AHRQ Publication No. 10(14)-EHC063-EF. Rockville, MD: Agency for Healthcare Research and Quality. January 2014. Chapters available at www.effectivehealthcare.ahrq.gov.
- Shea BJ, Grimshaw JM, Wells GA, et al. Development of AMSTAR: a measurement tool to assess the methodological quality of systematic reviews. BMC Med Res Methodol. 2007;7:10. PMID: 17302989.
- Parikh R, Mathai A, Parikh S, et al. Understanding and using sensitivity, specificity and predictive values. Indian J Ophthalmol. 2008;56(1):45. PMID: 18158403.
- McGee S. Simplifying likelihood ratios. J Gen Intern Med. 2002 Aug;17(8):646-9. PMID: 12213147.
- Deeks JJ, Altman DG. Diagnostic tests 4: likelihood ratios. BMJ. 2004 Jul 17;329(7458):168-9. PMID: 15258077.
- Chang SM, Matchar DB, Smetana GW, et al., eds. Methods Guide for Medical Test Reviews. AHRQ Publication No. 12-EC017. Rockville, MD: Agency for Healthcare Research and Quality; 2012. www.ncbi.nlm.nih.gov/pubmed/22834019.

- Li M, Yu LP, Tiberti C, et al. A report on the International Transglutaminase Autoantibody Workshop for Celiac Disease. Am J Gastroenterol. 2009 Jan;104(1): 154-63. PMID: 19098864.
- 20. Rozenberg O, Lerner A, Pacht A, et al. A novel algorithm for the diagnosis of celiac disease and a comprehensive review of celiac disease diagnostics. Clin Rev Allergy Immunol. 2012 Jun;42(3):331-41. PMID: 21279475.

Introduction

Background

Condition

Celiac disease (CD) is an immune-mediated disorder triggered in genetically-susceptible individuals by ingestion of foods containing gluten, a family of proteins found in wheat, rye, barley, and related grains.¹ The prevalence of CD in the United States has been estimated at approximately one percent,² but appears to be increasing, for reasons that are not clear.³ Risk factors for CD include family history, trisomy 21, Turner syndrome, and Williams syndrome, as well as several autoimmune diseases.

Clinical signs of CD include weight loss, iron deficiency anemia, aphthous ulcers, osteomalacia, dermatitis herpetiformis (a rash due to gluten-sensitivity), and gastrointestinal symptoms, including diarrhea and abdominal bloating. The diagnosis of CD can be challenging because the clinical spectrum of the disease varies, and some individuals present with mild symptoms.⁴

CD causes enteropathy of the small intestine resulting in poor absorption of nutrients. Malabsorption may result in several of the aforementioned clinical signs, including iron deficiency anemia, osteomalacia, and weight loss. Young children, in particular, are susceptible to failure to thrive, stunted growth, and delayed puberty.⁵ In women, folate deficiency secondary to CD may lead to poor birth outcomes, including developmental disorders. In the long-term, untreated CD increases the risk for non-Hodgkin's lymphoma, certain gastrointestinal cancers, and all-cause mortality.⁴

The only effective treatment for CD is avoidance of gluten in the diet. Timely diagnosis may be the most important component in the management of CD.

Diagnostic Strategies

A number of diagnostic methods have been developed; the validity and acceptability of some of these methods, particularly newer tests, which include combination tests and algorithms, remain controversial. Methods include various serology tests, HLA typing, video capsule endoscopy, and endoscopic duodenal biopsy (often considered the gold standard). Serology tests include anti-gliadin antibodies (AGA), IgA & IgG; anti-tissue transglutaminase (tTG), IgA & IgG; Endomysial antibodies (EmA), IgA; and the deamidated gliadin peptide (DGP) Antibodies, IgA & IgG. These tests are often used by providers as a panel in order to increase specificity and prevent false positives or increase sensitivity and prevent false negatives. All methods other than HLA typing require that the patient maintain a gluten containing diet during the diagnostic process. Commonly used diagnostic methods are described below.

Anti-gliadin antibodies (AGA), IgA & IgG. Gliadin is one of the two groups of proteins that constitute gluten. AGA determination was used as a diagnostic tool in the 1990s, as it has high sensitivity for CD.6 However, the test has low specificity, because anti-gliadin IgG is found in both acute and chronic common intestinal childhood diseases. In 2007, the World Gastroenterology Organization recommended against using these tests.7 In 2009, the UK National Institute for Clinical Excellence (NICE) also recommended against using the tests.8 As AGA tests are no longer recommended, they are not addressed in this systematic review.

Anti-tissue transglutaminase (tTG), IgA. Tissue transglutaminase is an enzyme that causes the crosslinking of certain proteins. Anti-tTG, IgA is the single test preferred by the American College of Gastroenterology (ACG) for the detection of celiac disease in those over the age of 2 years.5 These tests are also included in the algorithms of all recent guidelines. It is important to note that IgA deficiency is more prevalent in CD patients than in the general population; therefore, other tests may be ordered as an alternative in those who are IgA deficient.

Endomysial antibodies (EmA), IgA. The thin connective tissue layer that covers individual muscle fibers is called endomysium. When the intestinal lining is damaged, endomysial antibodies (EmA) develop. Most patients with active celiac disease and many with dermatitis herpetiformis have the IgA class of anti-EmA antibodies. Although this test is included in the algorithms of recent guidelines for diagnosis, it is not as widely used in the U.S. as in other countries, and many providers simply order a biopsy if the tTG levels are high. In addition, this test is less useful in individuals with low IgA.

Deamidated gliadin peptide (DGP) Antibodies. Elevated DGP antibodies are often seen in patients with celiac disease on a gluten-containing diet; this newer test may give a positive result in some individuals with CD who are anti-tTG negative, including children younger than 2 years old. Testing both DGP IgG and anti-tTG IgG is recommended by the ACG for those who have low IgA or IgA deficiency.5

Human leukocyte antigen (HLA) typing. Susceptibility to CD is linked to certain human leukocyte antigen (HLA) class II alleles, especially in the HLA-DQ region. HLA molecules are hypothesized to present gluten antigens to T-cells, which in turn induce tissue damage.9 Approximately 95 percent of patients with CD have the HLA-DQ2 heterodimer, while the remaining 5 percent of persons with CD have the HLA-DQ8 heterodimer.10 Since 25 percent to 40 percent of the U.S. population carries either the DQ2 or DQ8 gene, the presence of either is not pathognomonic for CD. However, lack of these heterodimers all but rules out CD and genetic susceptibility for the disorder. Thus, these genetic tests are routinely used to rule out CD and are part of the diagnostic algorithms recommended by the European Society for Pediatric Gastroenterology, Hepatology, and Nutrition (ESPGHAN) and the ACG.11

Video capsule endoscopy. In this procedure, a capsule containing a tiny camera is ingested by the patient, providing high quality visual evidence of CD. While not a traditional means of detecting CD, it is used in adults who seek to avoid biopsy. During Topic Refinement, Key Informants (KI) requested assessment of the evidence for this method in this report.

Endoscopic duodenal biopsy. Villous atrophy present on a duodenal biopsy and clinical remission when a gluten-free diet is followed represent the internationally accepted gold standard for CD diagnosis. The Modified Marsh criteria are utilized by most pathologists in evaluating histology findings from duodenal biopsy specimens for celiac disease diagnosis and progression of treatment during follow up.12 The criteria are graded from 0-3 with grade 3 further subdivided to 3a, 3b, and 3c.13, 14 Patients with Marsh grade 0 have normal histologic findings and are very unlikely to have celiac disease. In Marsh grade 1 and 2, biopsy specimens demonstrate raised Intraepithelial lymphocytes (IELs>30 per enterocytes) alone and raised IELs with crypt hyperplasia respectively. These histologic outcomes may be found in celiac patients on treatment or in patients with dermatitis herpetiformis. However, grade 1 or 2 lesions alone in the absence of clinical or serology evidence are nonspecific and are suggestive, but not confirmatory, of celiac disease.15 Patients with Grade 3 lesions have raised IELs with crypt hyperplasia and a measure of villous atrophy. Grade 3a, 3b, and 3c, in addition to raised IELs with crypt hyperplasia, also have findings of partial villous atrophy, subtotal villous atrophy, and

total villous atrophy respectively.14 Marsh grade 3 lesion is the classic celiac lesion and is characteristic, but not diagnostic, of celiac disease.16 Of note, some community physicians use a simple qualitative assessment of villous atrophy or elevation of intraepithelial lymphocytes to make a diagnosis rather than relying on Marsh criteria.

Obtaining properly oriented tissue samples can be difficult, patchy mucosal lesions can be missed, and limiting the portion of gut examined may risk missing the diagnosis of CD-related complications such as lymphoma and ulcerative jejunoileitis. Some patients and parents are concerned about the risk of adverse events such as perforations and bleeding. Patients may feel pain and discomfort, which is especially problematic for small children.

Combinations of the above. Many providers use a serology panel or sequential approach in order to prevent false positives that are associated with tests that don't work well under varying circumstances. The current systematic review compares the effectiveness of diagnostic tests, singularly and in combination.

Scope and Key Questions

Scope of the Review

Several systematic reviews and guidelines on diagnosis of CD have been published in the past decade, often with contradictory findings and recommendations. At least five recent guidelines for the diagnosis of CD have been published by recognized research/academic/medical bodies such as the North American Society for Pediatric Gastroenterology, Hepatology, and Nutrition (NASPGHAN)¹⁷ and ESPGHAN.¹¹

These clinical practice guidelines are complex and recommend different approaches to diagnosis. For example, some guidelines propose different sequences of tests for diagnosing population groups such as children versus adults, and symptomatic versus asymptomatic patients at increased risk (e.g. ESPGHAN). In addition, some guidelines (ACG and *World Gastroenterology Organization* [WGO]) uphold endoscopic biopsy as the gold standard for confirming diagnosis,^{5, 18} whereas other guidelines (ESPGHAN) explore the use of other tests to serve as substitutes for biopsy. The diagnosis of celiac disease is further complicated by lack of provider knowledge and variability in laboratory cut-off levels to indicate "positive" results. It is also unknown whether the same diagnostic criteria apply to different racial, ethnic, or other demographic subgroups, or if they may be incorrectly diagnosed or underdiagnosed. In addition, false positives and false negatives may have significant consequences: Positive diagnosis requires huge lifestyle changes, and undiagnosed CD can result in potential health harm (nutrient malabsorption, osteoporosis, and lymphoma).

This report compares the accuracy of the diagnostic methods listed above in children, adults, and sub-populations of interest to clinicians and patient groups. Diagnostic methods that are no longer included in guidelines or still in development (not approved in the U.S.) are beyond the scope of this project. Accuracy of serological tests and VCE are based on biopsy results as the reference standard. We also assess how biopsy results may vary by provider characteristics, technique, and the length of time the patient has consumed gluten. Finally, we report adverse events associated with invasive diagnostic methods (biopsy, VCE) and sequelae of false or indeterminate results of diagnosis. We provide below an analytic framework to illustrate the populations, interventions, outcomes, and adverse effects that guided the literature search and synthesis for this project (Figure 1).

Figure 1. Analytic framework, diagnosis of celiac disease



CD = celiac disease; FN = false negative; FP = false positive; IgA = immunoglobulin A; KQ = Key Question; LR+ = positive likelihood ratio; LR- = negative likelihood ratio; SES = socioeconomic status; TN = true negative; TP = true positive.

Key Questions

Four Key Questions guided this systematic review, as follows.

Key Question 1. What is the comparative effectiveness of the different diagnostic methods (various serological tests, HLA typing, video capsule endoscopy, used individually and in combination) compared with endoscopy with biopsy as the reference standard to diagnose celiac disease (CD) in terms of—

- a. Accuracy: sensitivity, specificity, positive likelihood ratio (LR+), negative likelihood ratio (LR-), summary receiver operating characteristics (ROCs)?
- b. Intermediate outcomes, such as clinical decisionmaking and dietary compliance?
- c. Clinical outcomes and complications related to CD?
- d. Patient-centered outcomes, such as quality of life (QOL) and symptoms?

Key Question 2. Does accuracy/reliability of endoscopy with duodenal biopsy vary by—

- a. Pathologist characteristics (i.e., level of experience or specific training)?
- b. Method (i.e., type or number of specimens)?
- c. Length of time ingesting gluten before diagnostic testing?

Key Question **3.** How do accuracy, (sensitivity, specificity, LR+, LR-, summary ROCs) and outcomes differ among specific populations (subgroups of Key Question 1), such as—

- a. Symptomatic patients versus nonsymptomatic individuals at risk?
- b. Adults (age 18 and over) versus children and adolescents?
- c. Children under age 24 months versus older children?
- d. Demographics, including race, genetics, geography, and socioeconomic status?
- e. Patients with IgA deficiency?
- f. Patients previously testing negative for CD?

Key Question 4. What are the direct adverse effects (i.e., bleeding from biopsy) or harms (related to false positives, false negatives, indeterminate results) associated with testing for CD?

In addition, we identify the following PICOTS (Populations, Interventions, Comparators, Outcomes, and Timing) for the Key Questions:

Population(s):

- For KQ 1, 2, and 4:
 - All populations tested for CD
- For KQ 3:
 - Patients with signs and symptoms of celiac disease, for example:
 - Diarrhea
 - Constipation
 - Dermatitis
 - Malabsorption (anemia, folate deficiency)
 - o Asymptomatic individuals at risk of celiac disease
 - Family history
 - Type 1 diabetes
 - Auto-immune disease
 - Turner's syndrome
 - Trisomy 21
 - o Children, under age 24 months vs older children & adolescents
 - o Adults (aged 18+)
 - Ethnic and geographic populations
 - o Low socioeconomic status (SES)
 - Patients with IgA deficiency
 - o Patients previously testing negative for CD

Interventions:

- For KQ 1, 3, 4:
 - Endomysial antibodies (EmA) IgA test
 - Anti-tissue transglutaminase (tTG) IgA test
 - o Deamidated gliadin poeptide (DGP) IgA antibodies
 - o EmA IgG, tTG IgG, and DGP IgG tests for IgA deficient individuals
 - HLA typing
 - Video capsule endoscopy
 - Combinations of the above
- For KQ 2:
 - Endoscopy with biopsy

Comparators:

- For KQ 1, 3:
 - Endoscopy with duodenal biopsy
- For KQ 2:
 - o Repeat biopsy

Outcomes:

- For KQ 1a, KQ2 and 3a-f, for Accuracy
 - o Sensitivity
 - o Specificity
 - o PPV, NPV, FP, FN
 - o Positive and negative likelihood ratios
- For KQ 1b, for Clinical decisionmaking
 - o Additional testing for CD
 - Nutritionist advice on gluten-free diet
 - Follow up and monitoring by MD
- For KQ 1c, for Clinical outcomes and complications
 - o Nutritional deficits
 - o Persistence of villous atrophy on biopsy
 - o Lymphomas
- For KQ 1d, for Patient-centered outcomes
 - Quality of life
 - Discomfort
 - o Bloating
 - o Abdominal pain
 - o Depression
- For KQ 4, for Harms
 - o Immediate AEs from biopsy
 - Psychological stress related to false positive results
 - o Sequelae of false negatives or indeterminate results

Timing:

- For KQ 2
 - Length of time ingesting gluten before biopsy

Setting:

- For all KQs
 - Outpatient: Academic
 - Outpatient: Community

Organization of This Report

The remainder of this report presents the methods used to conduct the literature searches, data abstraction, and analyses; the results of the literature searches and analyses; the conclusions; and a discussion of the limitations as well as suggestions for future research.
Methods

The methods for this comparative effectiveness review (CER) follow the methods suggested in the Agency for Healthcare Research and Quality (AHRQ) "Methods Guide for Effectiveness and Comparative Effectiveness Reviews" (available at

http://www.effectivehealthcare.ahrq.gov/methodsguide.cfm). The main sections in this chapter reflect the elements of the protocol established for the CER.

Topic Refinement and Review Protocol

KIs representing a variety of end-user perspectives were recruited to assist in refining the KQs and topics to address in this systematic review. Conference calls were held with community and academic-affiliated gastroenterologists, celiac disease researchers, a celiac disease centric nutritionist, national patient organizations, and a payer representative.

AHRQ posted the KQs on the Effective Health Care Web site for public comment in February, 2014. The EPC refined and finalized the KQs after review of the public comments, taking into consideration the prior input from KIs.

A study protocol was drafted and a technical expert panel (TEP) was recruited to provide high-level content and methodological expertise throughout the development of the review.

The final protocol for the project was posted on the AHRQ Effective Health Care Web site on June 11, 2014.

Literature Search Strategy

The literature search methods are summarized in Table 1 below. The full search strategy is presented as Appendix A.

Publication dates	For KQ1a, several recent high quality SRs exist. We searched for publication from January, 1990, but did not abstract studies that were already included in those SRs.			
	For KQ3, on specific populations, our search starts at January, 1990, the year the current EmA test began undergoing validation in the U.S. The other serological tests for CD were developed after that date (for example, the current tTG became available in 2000) so would be identified using a 1990 start date.			
	For KQ2, on duodenal biopsy, our search also starts at January, 1990. ESPGHAN published revised criteria for diagnosis of CD that year, ¹⁹ which reduced the suggested number of duodenal biopsies from three to one due to the advent of serological tests.			
	For KQ4, on direct and indirect harms of the diagnostic procedures, our search starts at January, 2003, as this Key Question was covered by an AHRQ-funded systematic review published in 2004. ²⁰			
Search terms	The search strategies were designed by a highly experienced reference librarian in collaboration with an expert on celiac disease and project staff experienced in SR methods. In brief, the strategy included search terms for celiac disease, combined with general terms for diagnosis or terms representing each diagnostic method, plus terms representing all outcomes listed in the PICOTs. The full search strategy is presented as Appendix A.			
	draft underwent peer review and public comment.			
Electronic databases	PubMed, Embase, Cochrane, Web of Science			
Scientific Information	Unpublished data were requested by an AHRQ-funded contractor from			

Table 1. Literature search methods

Packets (SIPs)	manufacturers of all serological tests.
Suggestions from experts	During the Topic Refinement period, KIs and project clinicians provided suggestions for studies to review. Members of this project's TEP also suggested studies. During review of the draft report, peer reviewers and the public had the opportunity to suggest additional studies.
Reference Mining	The reference lists of included articles were reviewed for identification of additional relevant studies.

Inclusion and Exclusion Criteria

The topic refinement phase involved a preliminary environmental scan. Several recent high quality systematic reviews (SR) on the accuracy of serological tests and VCE for diagnosis of celiac disease were identified. Electronic searches were conducted to identify additional primary studies that were not included in these SRs. Studies of diagnostic accuracy used designs such as controlled trials, prospective and retrospective cohorts, case-control studies, and case series. Studies were included if they met the following criteria:

- Diagnostic method is recommended in a current guideline (see list in Introduction.) For example, visualization via endoscopy (without biopsy) and AGA tests were excluded. Methods in development (genotyping other than HLA, ultrasound, mucosal immunohistochemistry, etc.) were also excluded.
- The tests were used to diagnose celiac disease, rather than for management of existing CD. For example, studies where serology was used to monitor adherence to diet or biopsy was used to monitor improvement of intestinal atrophy were excluded.
- All participants underwent both the "index test" and the reference standard (biopsy). If only subjects testing positive via serology underwent biopsy, the study was excluded.
- The study reports sensitivity, specificity, or data that allow their calculation.
- Study must be published in English
- Study must enroll consecutive or random sample
- Sample size is 300 or greater, unless one of the following is the focus
 - Low SES
 - Previously negative for celiac disease via serology or biopsy
 - o IgA deficient
 - o Type 1 diabetes
 - Turner's syndrome
 - o Trisomy 21 / Downs
 - Iron deficiency anemia
 - Family history
 - o Accuracy results are stratified by race / ethnicity

The following were excluded from this systematic review.

- Animal studies
- Individual case reports
- Studies not published in English
- Documents with no original data (commentary, editorial)
- Studies that measured only prevalence

Study Selection

Each title and abstract identified by the searches was screened by two researchers independently and the combination of their selections was retrieved for full-text review. Two researchers independently screened each full text article for inclusion in the project, with a senior researcher resolving discrepancies. A list of excluded studies and their reasons for exclusion is presented as Appendix B.

Data Extraction

The DistillerSR software package was used to manage the search output, screening, and data abstraction. The database can be used to calculate inter-rater reliability statistics of agreement and agreement adjusted for chance (kappa statistic) before resolution of disagreements. Assessment of inter-rater reliability can be used to guard against selection bias in choosing the articles for further review. Forms are displayed in Appendix D.

Data collection forms were designed by the project team in Distiller SR, piloted by the reviewers, further modified, and then the final forms piloted with a random selection of included studies to ensure agreement of interpretation. Study-level data abstracted included sample size; subjects' demographic characteristics, symptoms, and risk factors; study design; type of diagnostic test including cut-off level; and any other potential confounders. A statistician abstracted all accuracy data (sensitivity, specificity, and data needed to calculate). At the project's end, all abstracted data were uploaded to the federally-fund Systematic Review Data Repository.

Quality (Risk of Bias) Assessment of Individual Studies

Bias might be introduced at many points during the conduct of a study, affecting validity, reliability, and applicability of the results. Risk of bias of accuracy studies was assessed using the QUADAS-2 instrument; domains and items are described in Table 2 below.²¹ The AMSTAR instrument,²² described in Table 3, was used to assess the quality of prior systematic reviews. Finally, the McHarm instrument,²³ presented in Table 4, was used to assess the quality of studies on adverse events. Each study was scored individually by two EPC researchers who met to reconcile any differences; discrepancies were resolved by an experienced methodologist.

The Evidence Tables (Appendix C) display each QUADAS-2 item score for each accuracy study included in this review. Appendix E displays the AMSTAR scores. The McHarm scores are presented in the body of the report as there were few studies on adverse events. The strengths and weaknesses of the studies are discussed throughout the report and are reflected in the final Strength of Evidence ratings and Discussion.

Table 2. Quality Assessment of Diagnostic Accuracy Studies (QUADAS) -2 questions for
assessing risk of bias in diagnostic accuracy studies
Domain 1: Patient Selection

Was a consecutive or random sample of patients enrolled? (Yes/No/Unclear)
Was a case-control design avoided? (Yes/No/Unclear)
Did the study avoid inappropriate exclusions? (Yes/No/Unclear)
Could the selection of patients have introduced bias? Risk: Low/High/Unclear
Domain 2: Index Test(s) (complete for each index test used)
Were the index test results interpreted without knowledge of the reference standard? (Yes/No/Unclear)

If a threshold was used, was it pre-specified? (Yes/No/Unclear)

Could the conduct or interpretation of the index test have introduced bias? Risk: Low/High/Unclear

Domain 3: Reference Standard

Is the reference standard likely to correctly classify the target condition? (Yes/No/Unclear)

Were the reference standard results interpreted without knowledge of the results of the index test? (Yes/No/Unclear)

Could the reference standard, its conduct, or its interpretation have introduced bias?

Risk: Low/High/Unclear

Domain 4: Flow and Timing

Was there an appropriate interval between index test(s) and reference standard? (Yes/No/Unclear)

Did all patients receive a reference standard? (Yes/No/Unclear)

Did all patients receive the same reference standard? (Yes/No/Unclear)

Were all patients included in the analysis? (Yes/No/Unclear)

Could the patient flow have introduced bias? Risk: Low/High/Unclear

Table 3. AMSTAR (A Measurement Tool to Assess Systematic Reviews) criteria for assessing quality of systematic reviews

AMSTAR criteria:

- 01. Was an *a priori* study design provided?
- 02. Was there duplicate study selection and data extraction?
- 03. Was a comprehensive literature search performed?
- 04. Was the status of publication (gray literature) used as an inclusion criterion?
- 05. Was a list of studies (included/excluded) provided?
- 06. Were the characteristics of the included studies provided?
- 07. Was the scientific quality of the included studies assessed and documented?
- 08. Was the scientific quality of the included studies used appropriately in formulating conclusions?
- 09. Were the methods used to combine the findings of studies appropriate?
- 10. Was the likelihood of publication bias assessed?
- 11. Was the conflict of interest stated?

Table 4. McMaster Quality Assessment Scale for Harms (McHarm)

Item	Rating
1. Were the harms PRE-DEFINED using standardized or precise definitions?	~ Yes ~No ~Unsure
2. Were SERIOUS events precisely defined?	~ Yes ~No Not Applicable ~Unsure
3. Were SEVERE events precisely defined?	~ Yes ~No Not applicable ~Unsure
4. Were the number of DEATHS in each study group specified OR were the reason(s) for not specifying them given?	~ Yes ~No ~Unsure
5. Was the mode of harms collection specified as ACTIVE?	~ Yes ~No ~Unsure
6. Was the mode of harms collection specified as PASSIVE?	~ Yes ~No ~Unsure

7. Did the study specify WHO collected the harms?	~ Yes	~No
		~Unsure
8. Did the study specify the TRAINING or BACKGROUND of who ascertained the harms?	~ Yes	~No
		~Unsure
9. Did the study specify the TIMING and FREQUENCY of collection of the harms?	~ Yes	~No
		~Unsure
10. Did the author(s) use STANDARD scale(s) or checklist(s) for harms collection?	~ Yes	~No
		~Unsure
11. Did the authors specify if the harms reported encompass ALL the events collected or a	~ Yes	~No
selected SAMPLE?		~Unsure
12. Was the NUMBER of participants that withdrew or were lost to follow-up specified for each	~ Yes	~No
study group?		~Unsure
13. Was the TOTAL NUMBER of participants affected by harms specified for each study arm?	~ Yes	~No
		~Unsure
14. Did the author(s) specify the NUMBER for each TYPE of harmful event for each study	~ Yes	~No
group?		~Unsure
15. Did the author(s) specify the type of analyses undertaken for harms data?	~ Yes	~No
		~Unsure

Statistical Analyses

Studies that reported sensitivity, specificity, or receiver-operator characteristics, or provided the data to perform such calculations were abstracted for potential inclusion in a synthesis. Sensitivity is the ability of a test to correctly classify an individual as 'diseased' or in this case, having celiac disease, as confirmed by the "gold" standard (biopsy). Sensitivity ranges from 0 to 100 with values closer to 100 indicating a greater probability of a test being positive when the disease is present.²⁴ Sensitivity = (true positive) / (true positive + false negative). Specificity is the ability of a test to correctly classify an individual as 'disease-free' or in this case, as having symptoms or serologic results that indicate not having celiac disease when the individual doesn't have celiac disease, as determined by the gold standard (biopsy). Specificity ranges from 0 to 100 with values closer to 100 indicating a greater probability of a test being negative when the disease is not present.²⁴ Specificity = (true negative) / (true negative + false positive). A perfect diagnostic test would have both sensitivity and specificity of 100 percent. In general, sensitivity and specificity are considered good if at least 70.0 percent, very good from 80.0 percent to 89.9 percent, and excellent if 90.0 percent or greater.²⁴

Some studies of the accuracy of diagnostic tests report Likelihood Ratios (LR): probability of a positive finding in patients with a disease divided by the probability of the same finding in patients without the disease. Likelihood ratios can range from 0 to infinity. Diagnostic findings with LRs close to 0 indicate a decrease in the likelihood of disease. An LR of 1 indicates no change in the likelihood of disease.²⁵ As the LR increases from 1, the likelihood of disease increases in the presence of a positive test finding: LR+ = sensitivity / (1-specificity). Negative LRs are a measure of how the probability of the test is negative: LR- = (1-sensitivity) / specificity. An LR+ of more than 10 is considered good, as is an LR- of less than 0.1.²⁶

Finally, positive predictive value (PPV) is the probability that an individual who tests positive actually has the disease. Similarly, negative predictive value is the probability of not having a disease when an individual tests negative. Unlike sensitivity and specificity, predictive

values (PPV, NPV) are largely dependent on the prevalence of a disease in a study population. With increased prevalence in a population, PPV increases while NPV decreases.

If three or more studies of the same diagnostic method and comparator reported sensitivity and specificity, we considered pooling their result. In such cases, studies were weighted by sample size. Sensitivity and specificity were pooled using the 'mada' package in R which runs a bivariate diagnostic random-effects meta-analysis. The random effects model estimates a pooled sensitivity and false positive rate (used to calculate specificity) along with the associated confidence limits. This approach accounts for the interrelated sensitivity and specificity estimates.^{27, 28} The bivariate modeling approach that was used accounts for between study variability by allowing for the nonindependence of sensitivity and specificity across the studies.²⁹

Sensitivity analyses were conducted by stratifying by test type, cut-off level (titer), and population characteristics of interest. When pooling was not possible, study results were described narratively, according to comparisons of interest and study design, and presented either in tables or on figures.

Strength of the Body of Evidence

The overall strength of evidence for accuracy outcomes was assessed using guidance developed by experts in systematic reviews for the AHRQ Effective Health Care Program. This method classifies (grades) the evidence based on the following domains: study limitations (risk of bias), consistency, directness, and precision. The grades and their definitions are presented below in Table 5.

High	We are very confident that the estimate of effect lies close to the true effect for this outcome.
	The body of evidence has few or no deficiencies. We believe that the findings are stable, i.e.,
	another study would not change the conclusions
Moderate	We are moderately confident that the estimate of effect lies close to the true effect for this
	outcome. The body of evidence has some deficiencies. We believe that the findings are likely
	to be stable, but some doubt remains.
Low	We have limited confidence that the estimate of effect lies close to the true effect for this
	outcome. The body of evidence has major or numerous deficiencies (or both). We believe
	that additional evidence is needed before concluding either that the findings are stable or that
	the estimate of effect is close to the true effect.
Insufficient	We have no evidence, we are unable to estimate an effect, or we have no confidence in the
	estimate of effect for this outcome. No evidence is available or the body of evidence has
	unacceptable deficiencies, precluding reaching a conclusion.

Table 5. Strength of evidence definitions

Table 6 below, taken from the AHRQ Methods Guide for Diagnostic Tests,³⁰ briefly describes the methods used to rate each domain. The rating system was originally designed to assess the body of evidence on health care interventions rather than diagnostic tests; thus, assessing these domains presents unique challenges. For example, in assessing the precision of estimates of test performance, it may be difficult to judge whether a particular confidence interval has any practical clinical implications. In addition, there may be no direct evidence to link a specific test with clinical outcomes.

Table 6. Domains a	nd their definitions
--------------------	----------------------

Domain	Definition and Elements	Application
Study Limitations /	Risk of bias is the degree to which the	Use one of three levels of aggregate risk of bias:
Risk of Bias	included studies for a given outcome or	Low risk of bias
	comparison have a high likelihood of	Medium risk of bias
	adequate protection against bias (i.e.,	High risk of bias

Domain	Definition and Elements	Application
	good internal validity), assessed through main elements: Study design (e.g., RCTs or observational studies) Aggregate quality of the studies under consideration from the rating of quality (good/fair/poor) done for individual studies	Well designed and executed studies of new tests compared against an adequate criterion standard are rated as "Low risk of bias."
Consistency	Consistency is the degree to which reported study results (e.g., sensitivity, specificity, likelihood ratios) from included studies are similar. Consistency can be assessed through two main elements: The range of study results is narrow. Variability in study results is explained by differences in study design, patient population or test variability.	Use one of three levels of consistency: Consistent (i.e., no inconsistency) Inconsistent Unknown or not applicable (e.g., single study) Single-study evidence bases should be considered as "consistency unknown (single study)."
Directness	Directness relates to whether the evidence links the interventions directly to outcomes. For a comparison of two diagnostic tests, directness implies head- to-head comparisons against a common criterion standard. Directness may be contingent on the outcomes of interest.	Score dichotomously as one of two levels of directness: Direct Indirect When assessing the directness of the overarching question, if there are no studies linking the test to a clinical outcome, then evidence that only provides diagnostic accuracy outcomes would be considered indirect. If indirect, specify which of the two types of indirectness account for the rating (or both, if this is the case); namely, use of intermediate/surrogate outcomes rather than health outcomes, and use of indirect comparisons, If the decision is made to grade the strength of evidence of an intermediate outcome such as diagnostic accuracy, then the reviewer does not need to automatically "downgrade" this outcome for being indirect.
Precision	Precision is the degree of certainty surrounding an effect estimate with respect to a given outcome (i.e., for each outcome separately). If a meta-analysis was performed, the degree of certainty will be the confidence interval around the summary measure(s) of test performance (e.g., sensitivity, or true positive).	Score dichotomously as one of two levels of precision: Precise Imprecise A precise estimate is an estimate that would allow a clinically useful conclusion. An imprecise estimate is one for which the confidence interval is wide enough to include several distinct conclusions.

Applicability

Applicability assessment was based on the similarity of the populations in terms of characteristics listed in the PICOTs. These include age, gender, ethnicity, geographic location, SES, co-morbidities such as Type 1 diabetes, and symptoms such as iron deficiency. For example, a test might have high sensitivity and specificity in adults but not in small children, due to biological changes during the life course. These issues are addressed by KQ3.

Peer Review and Public Commentary

A draft version of this report was reviewed by several celiac disease experts. Reviewer names and affiliations are listed in the Acknowledgements section of the front matter. All peer

reviewers completed conflict of interest disclosures; none reported ties to any test manufacturer. The draft report was also posted on the AHRQ Effective Health Care Web site for public comment in February, 2015. Feedback from these sources was incorporated into the current version.

Results

Results of Literature Searches

As seen in Figure 2, the literature search identified a total of 7,254 titles as possibly related to the aims of this project. Of these, 68 were recommended by Key Informants, Technical Expert Panel (TEP) members, and a local content expert, 7,178 were identified by the electronic database searches, and an additional 8 were found through review of references of included studies. The titles of all 7,254 articles were screened, and 1,905 were rejected. Abstracts for the remaining 5,349 articles were reviewed, and an additional 3,649 articles were excluded, primarily because they were not about celiac disease (837) or were about celiac disease management rather than diagnosis (1,443). Other reasons for exclusion at abstract were the following: diagnostic methods out of our scope (endoscopic view without biopsy, experimental methods, etc.), focus on test processing (ELISA, PCR, etc.), serology with no biopsy comparison, simple prevalence report, no human subjects, no original data (commentary, editorial), data duplicated in other article, or individual case report. An additional 95 studies were rejected after abstract review, as they were already included in the identified relevant systematic reviews; this project was funded as a small systematic review (SR), which incorporated the aggregate results of prior SRs.

In total, 1,700 studies went on to the next phase of review. Of those, 345 did not have an abstract; upon review of a random 20-percent sample, we found none relevant and thus felt confident to exclude all. (Details are available upon request.) Four hundred and seventy were background articles, such as nonsystematic reviews, histories of diagnostic tests, or detailed descriptions of the biological processes involved. Excluding those studies left 1,230 research studies to retrieve for full-text screening. Thirteen prior systematic reviews were accepted. Based on full-text screening, 1,155 articles were excluded for the following reasons: not in English (1), not human (1), not about celiac disease (12), not about diagnostic tests (150), no original data (20), individual case reports (19), prevalence only (30), diagnostic methods beyond the scope of study (155), test processing issue (20), included in prior systematic reviews on topic (6), did not assess the accuracy or effectiveness of diagnostic tests (150), index test not compared to reference standard (biopsy) (62), not all subjects underwent both index test and reference standard (410), accuracy studies without consecutive or random sample (43), and accuracy studies where the sample size was less than 300 and not a special population (83). References for these excluded articles along with reasons for exclusion can be found in Appendix B.

Therefore, 60 individual studies and 13 SRs were included for evidence synthesis. Eleven SRs and 27 studies answered KQ1 and 3, one SR and 25 additional studies answered KQ2, and one SR and four studies answered KQ4.

Figure 2. Literature flow



CD = celiac disease; KQ = Key Question; SR = systematic review(s).

Key Question 1. Comparative Effectiveness

Description of Included Studies

We identified eleven systematic reviews (SRs) on the accuracy of the diagnostic methods within the scope of this review. Thirty-one studies published after the SRs reported sensitivity and specificity of these tests (or data that allowed their calculation) and met our inclusion criteria (consecutive or random sample, all subjects received index test and biopsy, sample size 300 or more, or special population of interest). The study characteristics are displayed in Table 7. With the exception of one study of over 12,000 subjects, studies ranged in size from 17 to 1,071subjects. Notably, only two studies were conducted in the U.S. Four studies were conducted in the UK, five in the Middle East, one in India, and the rest in Western Europe. Ethnicity and race of participants were rarely described. All but one study included individuals presenting for testing due to symptoms, risk factors, or family history. Only one study using general population screening met our inclusion criterion that sero-negative subjects undergo biopsy. Of the accepted studies, very few included asymptomatic individuals; only one compared accuracy results in symptomatic individuals with those of asymptomatic individuals. Many of the newer studies focused on algorithms using more than one serologic test.

The far right column in Table 7 summarizes the risk of bias of the new studies with regard to their ability to determine test accuracy. Bias in patient selection was rated high in almost half of the studies; eight studies were rated so because they used a case-control design. No studies had high risk of bias due to conduct of the index test, although almost half of the studies were missing some information on this issue. No studies had high risk of bias due to the conduct of the reference test (biopsy), although eight studies were missing information to make a determination. Finally, patient flow may have introduced a high risk of bias in four studies.

In sum, the risk or bias and applicability of these studies varied widely. These issues will be discussed throughout the report and summarized in the Discussion section.

Author, Year, Country	Study Design, Sample Size	Percent Female, Percent of Each Ethnicity, Population	QUADAS
Barada et al., 2014 ³¹	Prospective cohort	Percentage female: 55.2%	QUADAS Domain 1
			Biased patient selection: Low
Lebanon	Sample Size: 998	Middle eastern 100%	
			QUADAS Domain 2
		Population: Adults aged 18-70 (mean 43 years) with GI	Bias due to testing: Low
		symptoms of CD. 2.6% had family history. 20% with anemia	
			QUADAS Domain 3
			Bias due to reference test: Low
			QUADAS Domain 4
			Could patient flow have introduced
			bias: Not Applicable
Basso et al., 2011 ³²	Retrospective cohort	Percentage female: 62.4%	QUADAS Domain 1
	•	, , , , , , , , , , , , , , , , , , ,	Biased patient selection: Low
Italy	Sample Size: 703	Population: 100 U/ml. Children and adolescents with CD,	•
		latent CD, or controls. Mean age 8.	QUADAS Domain 2
			Bias due to testing: Unclear
			QUADAS Domain 3
			Bias due to reference test. Unclear
			QUADAS Domain 4
			Could patient flow have introduced
			bias: Not Applicable
Bienvenu et al., 2014 ³³	Retrospective cohort	Percentage female: 40%	QUADAS Domain 1
			Biased patient selection: Low
France	Sample Size:	Population:	
	45	IgA deficient children tested with CD-LFIA (detects both	QUADAS Domain 2
		human IgA and IgG anti DGP)	Bias due to testing: Low
			QUADAS Domain 3
			Bias due to reference test: Low
			QUADAS Domain 4
			Could patient flow have introduced
			bias: High

Table 7. Accuracy studies published after systematic reviews: characteristics

Author, Year, Country	Study Design, Sample Size	Percent Female, Percent of Each Ethnicity, Population	QUADAS
Cekin et al., 2012 ³⁴	Prospective cohort	Percentage female: 70.2%	QUADAS Domain 1
			Biased patient selection: Low
Turkey	Sample Size: 84	Population: Patients with Iron Deficiency Anemia of obscure	
		origin aged between 16 and 80 years were evaluated for	QUADAS Domain 2
		anti EmA IgA.	Bias due to testing: Low
			OLIADAS Domain 3
			Bias due to reference test: Unclear
			QUADAS Domain 4
			Could patient flow have introduced
			bias: Unclear
Dahlbom et al., 2010 ³⁵	Case control	Percentage female: Not reported	QUADAS Domain 1
			Biased patient selection: High
Sweden	Sample size: 301	Population: 52 children with severe malabsorption (group	
		I)median age of 1.6 yrs, 59 children with mild	QUADAS Domain 2
		symptoms(group II)median age of 8.1 yrs, 59 adults(group	Bias due to testing: Unclear
		III) median age of 39.5 yrs and 131 disease controls (adult	
		and children-median age of 10.8yrs).	QUADAS Domain 3
			Bias due to reference test: Unclear
			QUADAS Domain 4
			Could patient flow have introduced
			bias: Not Applicable
Dahle et al., 2010 ³⁶	Retrospective cohort	Percentage female: 54.5%	QUADAS Domain 1
			Biased patient selection: Low
Sweden	Sample size: 176	Population: Patients with symptoms who underwent	
		endoscopy and biopsy without previous serological testing	QUADAS Domain 2
		for anti-tTG or EmA	Bias due to testing: Unclear
			OLIADAS Domoin 2
			QUADAS Domain S Bias due te reference test: Low
			Dias due lo reference lest. Low
			QUADAS Domain 4
			Could patient flow have introduced
			bias: Unclear

Author, Year, Country	Study Design, Sample Size	Percent Female, Percent of Each Ethnicity, Population	QUADAS
DeGaetani et al., 2013 ³⁷	Retrospective cohort	Percentage female: 51.4%	QUADAS Domain 1
			Biased patient selection: High
US	Sample size: 72	Population: Adult patients (mean age 59yr) with villous	
		atrophy on biopsy and negative celiac serologies.	QUADAS Domain 2
			Bias due to testing: Unclear
			QUADAS Domain 3
			Bias due to reference test: Unclear
			OLIADAS Domoin 4
			QUADAS Domain 4
			bias: Unclear
Dutta et al., 2010 ³⁸	Retrospective cohort	Percentage female: 40.2%	QUADAS Domain 1
		l'oleonage lenale. leiz/	Biased patient selection: Low
India	Sample size: 92	Population: Symptomatic patients (age: 32.8 ± 17.4 years).	
			QUADAS Domain 2
			Bias due to testing: Low
			C C
			QUADAS Domain 3
			Bias due to reference test: Low
			QUADAS Domain 4
			Could patient flow have introduced
20			bias: Not Applicable
Emami et al., 2012 ³⁹	Retrospective cohort	Percentage female: 67.7%	QUADAS Domain 1
			Biased patient selection: Low
Iran	Sample size: 130	Middle eastern 100%	
			QUADAS Domain 2
		Population: Adult patients (mean age of 35.5+/-13.7) with	Bias due to testing: Unclear
		Iron deficiency anemia	OLIADAS Demain 2
			QUADAS Domain 3 Bios due te reference test: Lineleer
			Dias due lo reference lest. Officieal
			OLIADAS Domain 4
			Could patient flow have introduced
			bias: Unclear

Author, Year, Country	Study Design, Sample Size	Percent Female, Percent of Each Ethnicity, Population	QUADAS
Harrison et al., 2013 ⁴⁰	Retrospective cohort	Percentage female: Not reported	QUADAS Domain 1
			Biased patient selection: Low
UK	Sample size: 12289	Population: Patients tested for celiac disease using IgA tTG.	
		4 were IgA deficient(had error reading for IgA tTG, but	QUADAS Domain 2
		elevated levels of IgG tTG	Bias due to testing: Low
			OLIADAS Domain 3
			Bias due to reference test: I ow
			Dias due lo reference lest. Low
			QUADAS Domain 4
			Could patient flow have introduced
			bias: Not Applicable
Kaukinen et al., 1999 ⁴¹	Case control	Percentage female: 66.1%	QUADAS Domain 1
			Biased patient selection: High
Finland	Sample size: 26	Population: Patients with endocrine disorders with median	
		age of 37.7 years	QUADAS Domain 2
			Bias due to testing: Low
			OULADAS Domain 2
			QUADAS Domain 3 Biog due to reference test: Low
			bias due lo reference lest. Low
			QUADAS Domain 4
			Could patient flow have introduced
			bias: High
Mansour et al., 2011 ⁴²	Prospective cohort	Percentage female: 40.3%	QUADAS Domain 1
			Biased patient selection: Low
Iraq	Sample size: 62	Population: Children and adults with diabetes 1. Mean age	
		23.4 years	QUADAS Domain 2
			Bias due to testing: Low
			OULADAS Domoin 2
			QUADAS Domain 5 Bias due to reference test: Low
			Dias due lo reference lest. Low
			QUADAS Domain 4
			Could patient flow have introduced
			bias: Unclear

Author, Year, Country	Study Design, Sample Size	Percent Female, Percent of Each Ethnicity, Population	QUADAS
Mozo et al., 2012 ⁴³	Case control	Percentage female: 59.5%	QUADAS Domain 1
			Biased patient selection: High
Spain	Sample size: 200	Population: Children and adults diagnosed with CD or with	
		various digestive pathologies. CD children mean age 2,	QUADAS Domain 2
		control children mean age 2.8. CD adults mean age 39.1,	Bias due to testing: Low
		and control adults mean age 43.0.	
			QUADAS Domain 3
			Bias due to reference test: Unclear
			QUADAS Domain 4
			Could patient flow have introduced
			bias: Unclear
Nevoral et al., 201344	Retrospective cohort	Percentage female: Not reported	QUADAS Domain 1
	Sample size: 345	Population: Children and adolescents. 32 had first degree	Biased patient selection: Low
Czech Republic		relatives with CD, 60 had Type 1 diabetes, and 187 were	
		symptomatic	QUADAS Domain 2
			Bias due to testing: Low
			QUADAS Domain 3
			Bias due to reference test: Low
			QUADAS Domain 4
			Could patient flow have introduced
			bias: Not Applicable
Olen et al., 2012 ⁴⁵	Retrospective cohort	Percentage female: 57.4%	QUADAS Domain 1
			Biased patient selection: High
Sweden	Sample size:	Population: Children and adolescents under 18 years of	
	537	age. 71 children were under 2 years old. 16 were IgA	QUADAS Domain 2
		deficient.	Bias due to testing: Low
			QUADAS Domain 3
			Bias due to reference test: Low
			QUADAS Domain 4
			Could patient flow have introduced
			bias: Not Applicable

Author, Year, Country	Study Design, Sample Size	Percent Female, Percent of Each Ethnicity, Population	QUADAS
Sakly et al., 2012 ⁴⁶	Case control	Percentage female: Not reported	QUADAS Domain 1
			Biased patient selection: High
Tunisia	Sample size: 297	Population: Adult and pediatric patients with CD as well as	
		controls tested for IgA DGP	QUADAS Domain 2
			Bias due to testing: Unclear
			QUADAS Domain 3
			Bias due to reference test: Unclear
			QUADAS Domain 4
			Could patient flow have introduced
			bias: Unclear
Srinivas et al., 201447	Retrospective cohort	Percentage female: 66%	QUADAS Domain 1
			Biased patient selection: Low
UK	Sample Size:	Population:	
	752	CD vs. without CD in each of these groups: with clinical	QUADAS Domain 2
		features of CD, serology profile, and duodenal macroscopic	Bias due to testing: Unclear
		appearance	
			QUADAS Domain 3
			Bias due to reference test: Unclear
			QUADAS Domain 4
			Could patient flow have introduced
			bias: High
Srinivas et al., 201348	Retrospective cohort	Percentage female: 66%	QUADAS Domain 1
			Biased patient selection: Low
UK	Sample size: 752	Population: Symptomatic patients classified into 4 clinical	
		groups: high risk of CD, low risk of CD, nutrient deficiencies	QUADAS Domain 2
		and screening for diabetes.	Bias due to testing: Low
			OUADAS Demain 2
			Rias due to reference test: Low
			Dias due lo reference lest. LOW
			QUADAS Domain 4
			Could patient flow have introduced
			bias: Not Applicable

Author, Year, Country	Study Design, Sample Size	Percent Female, Percent of Each Ethnicity, Population	QUADAS
Sugai et al., 2010 ⁴⁹	Prospective cohort	Percentage female: Not reported	QUADAS Domain 1
			Biased patient selection: Low
Country Not Reported	Sample size: 17	Population: IgA tTG negative adults with villous atrophy	
			QUADAS Domain 2
			Bias due to testing: Low
			QUADAS Domain 3
			Bias due to reference test: Low
			OLIADAS Domoin 4
			QUADAS Domain 4
			bias: High
Swallow et al 2013 ⁵⁰	Retrospective cohort	Percentage female: Not reported	OLIADAS Domain 1
	Renospective conort	r ereentage female. Not reported	Biased patient selection: High
UK	Sample size: 756	Population: Adults	Blassa pallont selection. Thigh
	Campie 6126. 166		QUADAS Domain 2
			Bias due to testing: Low
			5
			QUADAS Domain 3
			Bias due to reference test: Low
			QUADAS Domain 4
			Could patient flow have introduced
			bias: Low
Van Meensel et al., 2004 ³¹	Retrospective cohort	Percentage female: 59.4%	QUADAS Domain 1
5.1.1			Biased patient selection: High
Belgium	Sample size:175	Population: Children and adult patients with biopsy-	
		confirmed CD, with GFD, 5 were IgA deficient.	QUADAS Domain 2
			Bias due to testing: Unclear
			OLIADAS Domain 3
			Bias due to reference test: Low
			QUADAS Domain 4
			Could patient flow have introduced
			bias: Unclear

Author, Year, Country	Study Design, Sample Size	Percent Female, Percent of Each Ethnicity, Population	QUADAS
Vermeersch et al., 2010 ⁵²	Case control	Percentage female: 59.9%	QUADAS Domain 1
			Biased patient selection: High
Belgium	Sample size: 827	Population: 86 consecutive CD patients (recruited between	
		August 1st 2000 and November 31st 2008) and 741	QUADAS Domain 2
		consecutive disease control patients (recruited between May	Bias due to testing: Unclear
		1st 2004 and October 31st 2007). The study population	
		consisted of 599 adults and 228 children with a mean age of	QUADAS Domain 3
		29 years	Bias due to reference test: Low
			OLIADAS Domain 4
			Could patient flow have introduced
			bias: Unclear
Vermeersch et al., 2010 ⁵³	Case control	Percentage female: 64.1%	QUADAS Domain 1
			Biased patient selection: High
Belgium	Sample size: 588	Population: 43 CD (mean age of 43.7) and 545 non-CD	
		(mean age of 39.8)	QUADAS Domain 2
			Bias due to testing: Unclear
			QUADAS Domain 3
			Bias due to reference test: Low
			QUADAS Domain 4
			Could patient flow have introduced
			bias: Unclear
Vermeersch et al., 2012 ⁵⁴	Case control	Percentage female: 59.2%	QUADAS Domain 1
			Biased patient selection: High
Belgium	Sample size: 649	Population: Children and adults. Average age of celiac	
		patients is 30.0 and that of controls is 26.4.	QUADAS Domain 2
			Bias due to testing: Unclear
			QUADAS Domain 3 Riss due to reference test: Low
			Bias due to reference test. Low
			QUADAS Domain 4
			Could patient flow have introduced
			bias: Unclear

Author, Year, Country	Study Design, Sample Size	Percent Female, Percent of Each Ethnicity, Population	QUADAS
Wakim-Fleming et al.,	Prospective cohort	Percentage female: 46.1%	QUADAS Domain 1
2014 ⁵⁵			Biased patient selection: Low
		White: 82.8%	
US		Black: 13.3%	QUADAS Domain 2
		Asian: 0.49%	Bias due to testing: Low
		Latino: 2.9%	
		American Indian: 0.49%	QUADAS Domain 3
	Operation Officer	Demoletien	Bias due to reference test: Low
	Sample Size:	Population:	OUADAS Domain 4
	204	Patients with biopsy proven cirmosis	QUADAS Domain 4
			bias: Low
			bias. Low
Walker 2010 56	Prospective cohort	Percentage female: 51 2%	OLIADAS Domain 1
			Biased patient selection: Low
Sweden			Diacoa pationi colocitoni 200
			QUADAS Domain 2
	Sample size:	Population:	Bias due to testing: Low
	1,001	Random sample of general population, adults	
			QUADAS Domain 3
			Bias due to reference test: Low
			QUADAS Domain 4
			Could patient flow have introduced
			bias: Low
W K to be 201457			
Wolf et al., 2014	Case control	Percentage female: 55%	QUADAS Domain 1
Multiple Europeen			Blased patient selection: High
	Sampla siza:	Population:	OUADAS Domain 2
countries	1071	Selective I_{α} deficiency (sI_{\alpha} \Delta D) in 27 patients	Bias due to testing: Low
	1071	Selective igA deliciency (sigAD) in 27 patients	Dias due to testing. Low
			QUADAS Domain 3
			Bias due to reference test: Low
			QUADAS Domain 4
			Could patient flow have introduced
			bias: Low

Author, Year, Country	Study Design, Sample Size	Percent Female, Percent of Each Ethnicity, Population	QUADAS
Zanini et al., 201258	Retrospective cohort	Percentage female: 76%	QUADAS Domain 1
			Biased patient selection: High
Italy	Sample size: 945	Population:	
-		Adult patients, aged 16 to 82 years, with symptoms,	QUADAS Domain 2
		familiarity or presence of associated diseases. Mean age 36.5	Bias due to testing: Unclear
			QUADAS Domain 3
			Bias due to reference test: Unclear
			QUADAS Domain 4 Could patient flow have introduced bias: Not Applicable

Au/ml = absorbance units per milliliter; CD = celiac disease; CI = confidence interval; DGP = deamidated gliadin peptide (DGP); HLA = human leukocyte antigen; IgA = immunoglobulin A; IgG = immunoglobulin G; L = liter; NR = not reported; QUADAS = Quality Assessment of Diagnostic Studies; tTG = anti-tissue transglutaminase; U = units; U/mL = units per milliliter.

Key Points

IgA tTG. A 2010 meta-analysis pooled 12 studies and found a sensitivity of 93.0 % (95% CI: 91.2%, 94.5%) and specificity of 96.5% (95% CI: 95.2%, 97.5%). A 2012 meta-analysis restricted to point of care tests in children reported sensitivity and specificity of 96.4% (95% CI: 94.3%, 97.9%) and 97.7% (95% CI: 95.8%, 99.0%) respectively when five studies were pooled. A 2013 systematic review did not pool data and echoed these results. We sixteen studies published after the prior SR search dates reported data needed for pooling; sensitivity was 92.6% (95% CI: 90.2%, 94.5%) and specificity 97.6% (95% CI: 96.3%, 98.5%). LR+ was 40.19% (95% CI: 25.29, 62.22) and LR- was 0.08 (95% CI: 0.06, 0.10). Significant heterogeneity was detected, as studies used a wide range of thresholds. Lower threshold levels increased sensitivity while higher threshold levels increased specificity. Our sensitivity analysis excluding data for threshold levels not used in clinical practice found sensitivity of 92.5% (95% CI: 89.7%, 94.6%) and specificity of 97.9% (95% CI: 96.5%, 98.7%). Positive predictive value (PPV) was 89.4% (95% CI: 88.3%, 90.5%) and negative predictive value (NPV) was 99.0% (95% CI: 98.8%, 99.1%).

IgA EmA. These tests have lower sensitivity than—and similar specificity to—IgA tTG tests, as confirmed by three SRs and several subsequent studies. LR+ was 65.98 (95% CI:29.64, 126.33) and LR- was 0.21 (95% CI: 0.14, 0.30) in our pooled analysis of seven studies published after the SRs, after dropping data points where Marsh 1 and 2 level atrophy was classified as CD (not standard practice). Pooled sensitivity was 79.0% (95% CI: 71.0%, 86.0%%); specificity was 99.0% (95% CI: 98.4%, 99.4%). PPV was 78.9% (95% CI: 71.0%, 85.5%) and NPV was 99.1% (95% CI: 98.6%, 99.5%). Significant heterogeneity was detected, likely due to variation in patient populations.

IgA DGP. A 2010 SR pooled eleven studies; sensitivity was estimated at 87.8% (95% CI: 85.6%, 89.9%) and specificity at 94.1% (95% CI: 95.2%, 97.5%) for all age groups combined. LR+ was 13.33 (95% CI: 9.64, 18.42) and LR- was 0.12 (95% CI: 0.08, 0.18). A 2012 SR reviewed the three studies that included only children: Sensitivities ranged from 80.7% to 95.1% (not pooled) and pooled specificity was estimated at 90.7% (95% CI: 87.8%, 93.1%). One new study reported sensitivity of 97.0% and specificity of 90.7% in symptomatic adults and children at one clinic, while another reported both sensitivity and specificity of 96% in a similar population.

IgG DGP. A 2013 SR of seven studies of non-IgA-deficient adults reported sensitivities of 75.4% to 96.7% and specificities of 98.5% to 100%. A 2012 SR of three studies in non-IgA-deficient children reported sensitivities of 80.1% to 98.6% and specificities of 86.0% to 96.9%. Authors of these reviews did not pool data. A new study reported sensitivity of 95.0% and specificity of 99.0% in 200 participants of all ages. In children under age seven, both sensitivity and specificity were 100.0%

<u>HLA-DQ2 and HLA-DQ8</u>. No SRs of the accuracy of testing for HLA-DQ2 or DQ8 were identified. These tests are used to rule out CD; the ACG estimates the negative predictive value of the combination at over 99%.

<u>Algorithms</u>. Nine studies of algorithms were identified; all used tTG tests. Adding an EmA test to a tTG test resulted in increased specificity, with either no change or a slight decrease in sensitivity. Adding a DGP test to a tTG test resulted in increased sensitivity but decreased specificity. However, more studies are needed to confirm the findings due to the wide range of values reported and populations studied.

<u>Video capsule endoscopy</u>. A previous SR of moderate quality pooled six studies and reported sensitivity of 89% (95% CI: 82.0%, 94.0%) and specificity of 95% (95% CI: 89.0%, 99.0%). LR+ was 12.90 (95% CI: 2.89, 57.58) and LR- was 0.16 (95% CI: 0.10, 0.25).

Adherence to gluten free diet. A previous SR of low quality indicated that 42% to 91% of adult CD patients in research studies strictly adhere to gluten-free diets; estimates varied primarily due to the definition and measurement of "strict." The SR included three studies that reported no statistical difference in adherence levels between patients whose celiac disease was detected via screening and those whose celiac disease was "symptom- detected." Association between specific diagnostic method and adherence was not addressed. We found only one study on this topic. Of ten blood donors in Israel who tested positive by tTG, only four adhered to gluten free diet; the other six patients did not believe they had CD, and four of those were told by physicians that asymptomatic patients did not need to modify their diets.

<u>Clinical outcomes, complications, or patient-centered outcomes.</u> We identified no studies of how these outcomes differ by initial diagnostic method.

Detailed Synthesis

Key Question 1a. Accuracy of Diagnostic Methods and Algorithms

Anti-tissue transglutaminase (tTG) tests. Four recent SRs assessed the accuracy of IgA tTG tests in symptomatic adults and children. They are discussed below; corresponding data are presented in Table 8. In 2010, Lewis⁵⁹ published a meta-analysis that pooled 12 studies and found a sensitivity of 93.0% (95% CI: 91.2%, 94.5%) and specificity of 96.5% (95% CI: 95.2%, 97.5%). LR+ was 25.62 (95% CI: 15.64, 41.99) and LR- was 0.07 (95% CI: 0.05, 0.12). Significant heterogeneity was detected. The previous year, the NICE clinical guidelines⁸ reported that based on 19 studies, sensitivities ranged from 38% to 100% and specificities ranged from 25% to 100%; pooling was not performed.

In another systematic review, Giersiepen, 2012⁶⁰ compared the accuracy of IgA tTG tests by assay method in children. Fifteen studies using ELISA reported sensitivities from 73.9% to 100% and specificities from 77.89% to 100%. Data were not pooled due to heterogeneity. Three studies using receptor binding assay (RBA) reported sensitivities ranging from 89.0% to 100%; pooled specificity was 95.6% (95% CI: 91.3%, 98.5%). Pooled sensitivity and specificity were 96.4% (95% CI: 94.3%, 97.9%) and 97.7% (95% CI: 95.8%, 99.0%) respectively in five point-of-care tests. Finally, Collatz-Schyum and colleagues (2013)⁶¹ reported in their systematic review that sensitivities ranged from 76.0% to 96.8% and specificities from 90.9% to 98.0% in eight studies of adults.

Collatz-Schyum⁶¹ and NICE each reported accuracy results from two studies of IgG tTG tests. The populations were heterogeneous, with some including IgA deficient subjects. Evidence is insufficient to estimate accuracy of this test in the non-IgA-deficient population.

Three of the SRs were of moderate quality. The NICE review did not present a table of characteristics of the included studies, Giersipen did not report whether dual selection or abstraction was used, and Lewis did not report an *a priori* design or protocol. Schyum's review was poor quality; the quality of the included studies was not assessed, and it was unclear whether studies were screened and abstracted in duplicate. A full assessment of the quality of each SR using the AMSTAR criteria is presented as Appendix E.

Table 8. Systematic reviews of tTG tests

Reference	Serolog ic Test	# of Studies	# Participants	Baseline Prevalence	Threshold for Positive	Method for Pooling	Sensitivity	Specificity	Additional Information
Lewis, Scott, 2010 ⁵⁹	IgA tTG	13 (5 adult, 2 children, 6 child/adult; 1 study did not report information to calculate sensitivity)	NR	NR	The Spearman correlation coefficient (calculated between the log odds of sensitivity and 1- specificity) gave no indication of a threshold effect (0.34, p=0.28).	DerSimonian Laird method in a random effects model	93.0% (95% Cl: 91.2-94.5)	96.5% (95% Cl: 95.2-97.5)	LR+ = 25.62 (95% CI: 15.64, 41.99) LR- = 0.07 (95% CI: 0.05, 0.12)
NICE Clinical Guidelines, 2009 ⁸	IgA tTG	19 (6 child, 9 adult, 4 child/adult)	4,799	NR	NR	None	Range 38 to 100% (adults 71 to 100%) (children 89 to 100%)	Range 25 to 100% (adults 65 to 100%) (children 25 to 100%)	One study that compared children <=2 years old vs. >2 years old found IgA tTG and IgA EmA to be similarly accurate.
Giersiepen et al., 2012 ⁶⁰	IgA tTG (ELISA only)	15 child	1,694 CD; 1,138 non- CD	59.82%	cut-off given by manufacturer	None	73.9 to 100% (not pooled due to heterogeneity)	77.8 to 100% (not pooled due to heterogeneity)	
Giersiepen et al., 2012 ⁶⁰	IgA tTG(RB As only)	3 child	255 CD; 146 non-CD	63.59%	cut-off given by manufacturer	MetaDiSc Software; weighted based on sample size	89.0 to 100% (not pooled due to heterogeneity)	95.9% (95% Cl: 91.3-98.5)	
Giersiepen et al., 2012 ⁶⁰	IgA TG2 POC (point of care)	5 child	470 CD; 399 non-CD	54.09%	cut-off given by manufacturer	MetaDiSc Software; weighted based on sample size	96.4% (95% Cl: 94.3-97.9)	97.7% (95% Cl: 95.8-99.0)	
NICE Clinical Guidelines, 2009 ⁸	lgG tTG	2 (1 adult, 1 child/adult)	365	NR	NR	None	23 to 85%	89 to 98%	

Reference	Serolog ic Test	# of Studies	# Participants	Baseline Prevalence	Threshold for Positive	Method for Pooling	Sensitivity	Specificity	Additional Information
Schyum, 2013 ⁶¹	lgA tTG	8 adult	3,871	39.1% to 44.9% in cohorts	20 U/ml	None	76.0% to 96.8%	90.9% to 98.0%	
Schyum, 2013 ⁶¹	lgG tTG	2 adult	Unclear	NR	NR	None	41.4% to 84.2%	98.8%	

tTG = anti-tissue transglutaminase; IgA = immunoglobulin A; ELISA = enzyme-linked immunosorbent assay; IgG = immunoglobulin G; NICE = National Institute for Clinical Excellence; NR = not reported; CD = celiac disease; CI = confidence interval RBA= radio blinding assays.

We identified 19 additional original studies reporting accuracy of tTG IgA tests. Study characteristics and risk of bias are displayed in Table 7 at the beginning of this chapter. Three included only IgA-deficient individuals^{43, 45, 52} and will be discussed in the results for Key Question 3 on populations of special interest. The pooled sensitivity and specificity results for the sixteen other studies^{47, 55, 57 31, 40 32, 39, 48, 50, 54, 58 35, 36, 53 42, 51} are displayed in Figure 3 and Table 9 below. (Data from the 16 new studies could not be pooled with the results of the prior published meta-analyses because the prior reviews did not report the number of true and false positives and negatives by arm for each study.) Pooled sensitivity was 92.6% (95% CI: 90.2%, 94.5%) and pooled specificity was 97.6% (95% CI: 96.3%, 98.5%). The LR+ was 40.19 (95% CI: 25.29, 62.22) and the LR- was 0.08 (95% CI: 0.06, 0.10). We did not calculate a summary ROC because the data reported high accuracy at a wide variety of threshold levels. The sensitivity and specificity obtained in our pooled analysis are not statistically different from those of the recent Lewis meta-analysis and confirm the high accuracy rate of tTG tests in non-IgA deficient individuals. The I-squared value in our analysis was 96.9%, indicating evidence of heterogeneity, likely due to the wide range of threshold levels used in the studies.



Figure 3. Sensitivity and specificity results for tissue transglutaminase immunoglobulin A tests

Table 9. Accur	acy of	tTG l	gA tests
----------------	--------	-------	----------

		True Positive	False Negative	False Positive	True Negative	Sensitivity	Specificity
tTG lgA	Threshold						
Van Meensel, 2004 ⁵¹	2.64 kilounits/L	101	4	1	69	0.96	0.99
Dahlbom,	3 U m/L	170	0	1	130	1.00	0.99

		True Positive	False Negative	False Positive	True Negative	Sensitivity	Specificity
tTG IgA	Threshold		•		•		
2010 ³⁵							
Van Meensel, 2004 ⁵¹	3.13 kilounits/L	101	4	1	69	0.96	0.99
Van Meensel, 2004 ⁵¹	3.69 kilounits/L	101	4	0	70	0.96	1.00
Van Meensel, 2004 ⁵¹	4 kilounits/L	98	7	1	69	0.93	0.99
	4.43 kilounits/L	104	1	1	69	0.99	0.99
	5 kilounits/L	98	7	1	69	0.93	0.99
Dahle, 2010 ³⁶	5 U/mL	NR	NR	NR	NR	0.76	0.95
Van Meensel, 2004 ⁵¹	7 kilounits/L	96	9	0	70	0.91	1.00
	7 kilounits/L	102	3	0	70	0.97	1.00
Vermeersch, 2010 ⁵³	7 U/mL	NR	NR	NR	NR	0.95	0.93
Vermeersch, 2012 ⁵⁴	7 U/mL	NR	NR	NR	NR	0.81	0.99
Zanini, 2012 ⁵⁸	7 U/mL	NR	NR	NR	NR	0.95	0.76
Van Meensel, 2004 ⁵¹	7.16 kilounits/L	102	3	0	70	0.97	1.00
	7.98 kilounits	101	4	0	70	0.96	1.00
Zanini, 2012 ⁵⁸	8 U/mL	NR	NR	NR	NR	0.88	0.92
Van Meensel, 2004 ⁵¹	9.73 kilounits/L	99	6	0	70	0.94	1.00
	10 kilounits/L	99	6	0	70	0.94	1.00
Emami, 2012 ³⁹	10 AU/mL	5	8	4	113	0.38	0.97
Srinivas, 201348	10 IU/mL	NR	NR	NR	NR	0.84	0.96
Wolf, 2014 ³⁷	10 ULN	310	10 (32 grey zone)	2 (17 grey zone)	673	0.88	0.97
Srinivas, 2014 47	10 IU/mL	73	15	29	635	0.83	0.96
Van Meensel, 2004 ⁵¹	15 kilounits	99	6	0	70	0.94	1.00
Vermeersch, 2010 ⁵³	15 U/mL	NR	NR	NR	NR	0.86	0.95
Mansour, 2011 ⁴²	15 U/mL	5	2	4	51	0.71	0.93
Zanini, 201258	16 U/mL	NR	NR	NR	NR	0.89	0.88
Basso, 2011 ³²	17.5 U/mL	NR	NR	NR	NR	0.95	0.97
Van Meensel, 2004 ⁵¹	19.05 kilounits/L	98	7	0	70	0.93	1.00
	20 kilounits/L	102	3	3	67	0.97	0.96
	20 kilounits/L	98	7	0	70	0.93	1.00
Vermeersch, 2012 ⁵⁴	20 U/mL	NR	NR	NR	NR	0.84	0.96
Basso, 2011 ³²	20 U/mL	NR	NR	NR	NR	0.94	0.97
Wakim-Fleming, 2014 55	20 IU	5	0	7	192	1.00	0.96

_		True Positive	False Negative	False Positive	True Negative	Sensitivity	Specificity
tTG lgA	Threshold						
Wolf, 2014 57	20 U/mL	342	10	19	673	0.97	0.97
Van Meensel, 2004 ⁵¹	20.47 kilounits	102	3	0	70	0.97	1.00
Zanini, 2012 ⁵⁸	21 U/mL	NR	NR	NR	NR	0.38	0.97
Basso, 2011 ³²	24 U/mL	NR	NR	NR	NR	0.96	0.81
Zanini, 2012 ⁵⁸	24 U/mL	NR	NR	NR	NR	0.59	0.99
	35 U/mL	NR	NR	NR	NR	0.10	1.00
Van Meensel, 2004 ⁵¹	40 kilounits/L	101	4	3	67	0.96	0.96
Zanini, 2012 ⁵⁸	40 U/mL	NR	NR	NR	NR	0.43	1.00
	48 U/mL	NR	NR	NR	NR	0.70	0.59
Van Meensel, 2004 ⁵¹	50 kilounits/L	98	7	5	65	0.93	0.93
	56.9 kilounits/L	96	9	1	69	0.91	0.99
Basso, 2011 ³²	75.6 U/mL	NR	NR	NR	NR	0.91	0.97
Zanini, 2012 ⁵⁸	80 U/mL	NR	NR	NR	NR	0.59	0.43
Basso, 2011 ³²	100 U/mL	NR	NR	NR	NR	0.76	1.00
	909.3 U/mL	NR	NR	NR	NR	0.63	1.00
Barada, 2014 ³¹	NR	NR	NR	NR	NR	0.72	0.98
Harrison, 2013 $_{40}$	NR	66	10	11	12,202	0.87	1.00
Swallow, 2013 ⁵⁰	NR	5	2	72	654	0.75	0.90
	NR	26	4	72	654	0.87	0.90
	NR	20	3	77	656	0.88	0.90

Pooled Results* Sensitivity: 0.93 (0.90, 0.95) Specificity: 0.98 (0.96, 0.99) *pooled results include only studies with all information (true and false negatives and positives) reported

tTG = anti-tissue transglutaminase; IgA = immunoglobulin A; U = unit; mL = milliliter; grey zone = indeterminate results; NR = not reported.

Different laboratories or test manufacturers may define "positive" celiac disease diagnosis using different threshold levels of IgA tTG. Due to the wide range of thresholds used in the research studies and the heterogeneity identified in our meta-analysis, we conducted a sensitivity analysis by omitting accuracy results for thresholds not used in clinical practice. This pooled analysis of tests that used threshold levels of less than 40 U/mL resulted in sensitivity of 92.5% (95% CI: 89.7%, 94.6%) and specificity of 97.9% (95% CI: 96.5%, 98.7%). This result does not differ significantly from the results when all thresholds were pooled. Positive predictive value (PPV) was 89.4% (95% CI: 88.3%, 90.5%) and negative predictive value was 99.0% (95% CI: 98.8%, 99.1%).

Figure 4 displays sensitivity and specificity by threshold or "cut-off" where reported. Data were insufficient to pool by threshold level, but diagnostic thresholds of 31 to 40 U/ml reported low sensitivity, and thresholds of 40 U/ml or higher reported very low sensitivity. The higher thresholds are not currently recommended; they were reported by studies that aimed to define a cut-off value of tTG antibody with 100% specificity^{51, 62} or high positive likelihood ratio for duodenal atrophy in patients with suspected celiac disease⁵⁸ One study concluded that a cutoff

level five-fold higher than the upper limit of normal is 100% specific for duodenal atrophy and using this cut-off could prevent the need for biopsy in one-third of patients.⁵⁸



Figure 4. Accuracy by threshold level for tissue transglutaminase immunoglobulin A

Endomysial antibodies (EmA) tests. Three recent SRs of IgA EmA tests were identified. A review conducted for the 2009 NICE clinical guidelines⁸ included 23 studies: 10 of children, nine of adults, and four of both adults and children. Sensitivity ranged from 68% to 100%, while specificity ranged from 77% to 100%. Pooling was not performed. Giersiepen⁶⁰ included 11 studies of IgA EmA accuracy in children. Sensitivity ranged from 82.6% to 100%. Pooled specificity was 98.2% (95% CI: 96.7%, 99.1%). Finally, Schyum, 2013⁶¹ described four studies of adults that were included in the previous reviews; pooling was not conducted. As mentioned above, quality of the NICE and Giersiepen SRs was rated as moderate. Data are presented in Table 10.

Table 10. Systematic reviews of EmA IgA tests

Reference	Serologic Test	# of Studies	# Participants	Baseline Prevalence	Threshold for Positive	Method for Pooling	Sensitivity	Specificity	Additional Information
Giersiepen et al., 2012 ⁶⁰	IgA EmA	11 child	1,034 CD; 558 non-CD	64.95%	cut-off given by manufacturer	Weighted based on sample size	82.6 to 100% (not pooled due to heterogeneity)	98.2% (95% CI: 96.7-99.1)	
NICE Clinical Guidelines, 2009 ⁸	IgA EmA	23 (10 child, 9 adult, 4 child/adult)	5,529	NR	NR	None	Range 68 to 100% (adults 68 to 100%) (children 46 to 100%)	Range 89 to 100% (adults 94 to 100%) (children 77 to 100%)	Reported results on 1 study that compared children <=2 years old vs. >2 years old and found IgA tTG and IgA EMA to have similar accuracy.
Schyum, 2013 ⁶¹	IgA EmA	4 adult	2,537	Unclear	NR	None	61.0% to 93.7%	98.0% to 100%	

EmA = endomysial antibodies; IgA = immunoglobulin A; CD = celiac disease; CI = confidence interval NICE= National Institute for Clinical Excellence; NR = not reported.

Seven additional studies that reported sensitivity and specificity of IgA EmA tests^{31, 34, 36, 42, 47, 50, 55} were identified. Accuracy results are displayed in Figure 5 and Table 11. Swallow (2013)⁵⁰ reported results for three different biopsy reference standards (definitions of celiac disease): a) Marsh 1-2 villous atrophy, b) Marsh 1-3 villous atrophy, and c) Marsh 3villous atrophy. Sensitivities were 42.9%, 73.3%, and 82.6%, respectively.

Two studies^{31, 36} reported inadequate data for inclusion in pooling. Our pooling of the seven available data points resulted in sensitivity of 76.6% (95% CI: 68.7%, 82.9%) and specificity of 99.0% (95% CI: 98.4%, 99.4%). After excluding the two data points where, contrary to standard practice, Marsh 1 and 2 level villous atrophy were classified as CD , pooled sensitivity was 79.0% (95% CI: 71.0%, 86.0%) and pooled specificity was 99.0% (95% CI: 98.4%, 99.4%). LR+ was 65.98 (95% CI: 29.64, 126.33) and LR- was 0.21 (95% CI: 0.14, 0.30). Positive predictive value (PPV) was 78.9% (95% CI: 71.0%, 85.5%) and negative predictive value was 99.1% (95% CI: 98.6%, 99.5%). I-squared value was 81.8%, indicating evidence of heterogeneity. These accuracy results support findings of prior systematic reviews.,

Figure 5. Accuracy of endomysial antibodies immunoglobulin A studies published after NICE and ESPGHAN systematic reviews



Table 11. Accuracy of EmA IgA tests in studies published after NICE and ESPGHAN systematic reviews

	Threshold	True Positive	False Negative	False Positive	True Negative	Sensitivity	Specificity
Mansour, 2011 ⁴²	20 U/mL	5	2	2	53	0.71	0.96

	Threshold	True Positive	False Negative	False Positive	True Negative	Sensitivity	Specificity
Barada, 2014 ³¹	NR	NR	NR	NR	NR	0.72	1.00
Swallow, 2013 ⁵⁰	NR, Marsh 1- 3classified as celiac NR, Marsh 2 &	3	4	4	722	0.43	1.00
	3 classified as celiac NR, March 3 classified as	22	8	4	722	0.73	1.00
Makim Flaming	Cellac Corum dilution	19	4	1	120	0.63	0.99
2014 ⁵⁵	≥ 1/10	5	0	0	199	1.00	1.00
Srinivas, 2014 47	NR	70	18	5	659	0.80	0.99
Cekin, 2012 ³⁴	NR	6	0	1	77	1.00	0.99
Dahle, 2010 ³⁶	Serum dilution	NR	NR	NR	NR	0.61	1.00
Pooled Results*	Sensitivity:	0.77 (0.69,0.8	33)	Specificity: ().99 (0.98,0.99))	

*Pooled results include only studies with all information (false and true negatives and positives) reported.

EmA = endomysial antibodies; IgA = immunoglobulin A; NICE = National Institute for Clinical Excellence; ESPGHAN = European Society for Paediatric Gastroenterology Hepatology and Nutrition; U = unit; mL = milliliter; NR = not reported.

Deamidated gliadin peptide (DGP) antibodies test -IgA. Four recent SRs of DGP tests were identified; details are presented in Table 12. The 2009 NICE guideline report⁸ identified only two studies of IgA DGP. Both studies were included in a 2010 review by Lewis and colleagues of 12 studies;⁵⁹ we were able to pool eleven studies and sensitivity was estimated at 87.8% (95% CI: 85.6%, 89.9%) while pooled specificity was 94.1% (95% CI: 92.5%, 95.5%). LR+ was 13.33 (95% CI: 9.64, 18.42) and LR- was 0.12 (95% CI: 0.08, 0.18). Significant heterogeneity was detected. Giersiepen⁶⁰ reviewed three studies of IgA DGP accuracy in children that were included in the Lewis SR. Sensitivity ranged from 80.7% to 95.1% (not pooled) and specificity was estimated at 90.7% (95% CI: 87.8%, 93.1%). Schyum ⁶¹ identified seven studies in adults; sensitivity ranged from 69.0% to 98.4% while specificity ranged from 90.3% to 98.0%. We identified three studies not included in prior SRs^{46, 49, 43} that reported sensitivity and specificity of IgA DGP tests in non-IgA-deficient individuals. Sugai⁴⁹ assessed accuracy in 17 IgA tTG-negative patients with villous atrophy; results will be discussed under Key Question 3, which considers populations of special interest (previously seronegative subjects). Sakly⁴⁶ reported sensitivity of 97.0% and specificity of 90.7% in 297 symptomatic adults and children at one clinic. Mozo (2012)⁴³ conducted a case-control study in Spain that included 100 newly diagnosed adults and children and 100 age-matched controls. (Six patients were IgA deficient.) Both sensitivity and specificity of IgA DGP were 96.0% and AUC was 98.8%. Deamidated gliadin peptide (DGP) antibodies test -IgG. Two recent SRs reported accuracy of IgG DGP tests in non-IgA deficient subjects. Details are displayed in Table 12. Giersiepen, 2012 ⁶⁰ reported sensitivity ranging from 80.1% to 98.6% and specificity from 86.0% to 96.9% in three studies of children. In a SR of seven studies of adults compiled by Schyum, sensitivity ranged from 75.4% to 96.7%,⁶¹ while specificity ranged from 98.5% to 100%.

We identified one study of IgG DGP tests published after these systematic reviews. In the case control study noted above, Mozo (2012)⁴³ reported sensitivity and specificity of IgG DGP test

were 95.0% and 99.0% respectively, with AUC of 99.5%. In children age seven or younger, both sensitivity and specificity of IgG DGP were 100.0%.

Table 12. Systematic reviews of DGP tests

Reference	Serologic Test	# of Studies	# Participants	Baseline Prevalence	Threshold for Positive	Method for Pooling	Sensitivity	Specificity	Additional Information
NICE Clinical Guidelines, 2009 ⁸	DGP-based assays	2 (1 children, 1 adult)	317	NR	NR	none	Range 90.8 to 100% (adults 96.7 to 100%) (children 90.8 to 100%)	Range 93.8 to 100% (adults 93.8 to 100%) (children 94.7 to 98.2%)	Studies described narratively
Giersiepen et al., 2012	IgA DGP	3 children	422 CD; 346 non-CD	54.95%	cut-off given by manufacturer	MetaDiSc Software; weighted based on sample size	80.7 to 95.1% (not pooled due to heterogeneity)	90.7% (95% CI: 87.8-93.1)	
Lewis, Scott 2010 ⁵⁹	IgA DGP	12 (1 study did not report information to calculate sensitivity; 4 adult, 2 children, 6 child/adult)	NR	NR	The Spearman correlation (calculated between the log odds of sensitivity and 1- specificity) gave no indication of a threshold effect (-0.23, p=0.50).	Meta-DiSc and STATA; DerSimonian Laird method in a random effects model	87.8% (95% Cl: 85.6-89.9)	94.1% (95% CI: 92.5-95.5)	LR+ = 13.33 (95% CI: 9.64, 18.42) LR- = 0.12 (%% CI: 0.08, 0.18)
Schyum, 2013 ⁶¹	IgA DGP	7 adult	2,555	Unclear	NR	None	69.0% to 98.4%	90.3% to 98.0%	
Schyum, 2013 ⁶¹	IgG DGP	7 adult	2,322	Unclear	NR	None	75.4% to 96.7%	98.5% to 100%	
Giersiepen, 2012 ⁶⁰	IgG DGP	3 children	422 CD, 346 non-CD	54.95%	NR	None	80.1% to 98.6%	86.0% to 96.9%	

IgA= immunoglobulin A; DGP= deamidated gliadin peptide; NICE= National Institute for Clinical Excellence; NR = not reported; CD = celiac disease; CI: confidence interval.

Human leukocyte antigen (HLA) DQ2 and DQ8. These tests are used to rule out CD, as the ACG⁵ estimates the negative predictive value of the combination at over 99%. It is estimated that 95% of CD patients are positive for HLA-DQ2, and the remainder are positive for HLA-DQ8. Patients who test negative for HLA-DQ2 and HLA-DQ8 simultaneously will rarely undergo further testing for CD. One SR⁶¹ discussed studies of HLA typing but did not report sensitivity or specificity.

<u>Algorithms.</u> We identified nine studies that used multiple serological tests simultaneously in diagnostic algorithms. Each algorithm combined a tTG screen with an additional serological test. Accuracy results are displayed in Table 13 below. Data could not be pooled due to study heterogeneity. Algorithms that combined a tTG test and EmA test had sensitivity ranging from 57% to 93% and specificity from 64% to 99%. One study⁵⁰ reported low sensitivity when the definition of celiac disease included patients with Marsh 1-2 level atrophy. In the same study, when only patients with atrophy level Marsh 3 or higher were considered to have celiac disease, sensitivity was 87% and specificity was 97%.

Algorithms combining tTG and DGP screens reported sensitivity ranging from 65% to 97% and specificity from 80% to 100%. Low sensitivity (65%) was reported when a threshold of 145 U/ml was used for tTG test. However, this is not common clinical practice: The authors used this high threshold level to achieve specificity of 100%. A high threshold was also used by Sugai⁴⁹ for the same purpose. Of note, two new combined tTG IgA + DGP IgG tests reported high sensitivity and specificity.⁵⁴

While this report underwent peer review, we identified a SR that included three studies of combination assays.⁶¹ Two were already included in this report; the third did not meet our inclusion criteria.

In general, the sensitivity and specificity of algorithms were not significantly higher than that of the individual tests used alone, and it is unclear whether the small increases in accuracy are clinically meaningful.

Table 13. Accuracy of algorithms

	Test Type	Notes	True Positive	False Negative	False Positive	True Negative	Sensitivity	Specificity
Barada, 2014 ³¹	tTG IgA, DGP IgA	Symptomatic adults in Lebanon	13	5	25	955	0.722	0.974
	tTG lgA	Symptomatic adults in Lebanon	13	5	16	965	0.722	0.984
	EmA IgA	Symptomatic adults in Lebanon	13	5	3	978	0.722	0.995
Basso, 2011 ³²	tTG IgA, DGP IgA	Children, threshold 145 U/ml	NR	NR	NR	NR	0.653	1.000
	tTG IgA	Children, threshold 100 U/ml	NR	NR	NR	NR	0.757	1.000
	tTG IgA, DGP IgA	Children, threshold 20 U/ml	NR	NR	NR	NR	0.967	0.898
	tTG lgA	Children, threshold 20 U/ml	NR	NR	NR	NR	0.942	0.973
	tTG IgA, DGP IgA	Children, Threshold 32 U/ml	NR	NR	NR	NR	0.945	0.957
	tTG IgA	Children, Threshold 17.5 U/ml	NR	NR	NR	NR	0.945	0.971
Dahle, 2010 ³⁶	tTG IgG or IgA combined w/ DGP IgG or IgA	Symptomatic adults, Threshold 20 Au/mL	NR	NR	NR	NR	0.91	0.80
	tTG IgG or IgA combined w/ DGP IgG or IgA	Symptomatic adults, Threshold 35 Au/ml	NR	NR	NR	NR	0.85	0.98
	tTG lgA	Symptomatic adults, Threshold 5 U/mL	NR	NR	NR	NR	0.76	0.95
	DGP IgG or IgA	Symptomatic adults, Threshold 20 Au/mL	NR	NR	NR	NR	0.87	0.96
	EmA IgA	Symptomatic adults	NR	NR	NR	NR	0.61	1.00
Nevoral, 201344	tTG IgA, EmA IgA	Marsh 2 or 3	NR	NR	NR	NR	0.76	0.85
	tTG IgA, EmA IgA	First degree relatives	NR	NR	NR	NR	0.81	0.70
	tTG IgA, EmA IgA	Asymptomatic Marsh 2 or 3	NR	NR	NR	NR	0.83	0.67
	tTG IgA, EmA IgA	Type 1 diabetes	NR	NR	NR	NR	0.93	0.64
Srinivas, 2013 ⁴⁸	tTG IgA, EmA IgA	Symptomatic or Type 1 Diabetes	NR	NR	NR	NR	0.83	0.99
	tTG IgA	Symptomatic or Type 1 Diabetes	NR	NR	NR	NR	0.84	0.96
	EmA IgA	Symptomatic or Type 1 Diabetes	NR	NR	NR	NR	0.83	0.99
	Test Type	Notos	True Positive	False Negative	False Positive	True Negative	Sonsitivity	Specificity
--------------------------------	---	---	------------------	------------------------	------------------------	------------------	-------------	-------------
Sugai, 2010 ⁴⁹	tTG IgA, DGP IgA or IgG	tTG positive and negative patients with enteropathy	6	8 8	0	3	0.43	1.00
Swallow, 2013 ⁵⁰	tTG IgA, EmA IgA - NICE two step strategy	Adults, Marsh 1-2	4	3	20	706	0.57	0.97
	tTG IgA, EmA IgA - NICE two step strategy	Adults, Marsh 1-3 considered celiac	24	6	20	706	0.80	0.97
	tTG IgA, EmA IgA - NICE two step strategy	Adults, Marsh 3 considered celiac	20	3	23	710	0.87	0.97
	tTG IgA, 2008-2009 data	Adults, Marsh 3 considered celiac	NR	NR	NR	NR	0.875	0.895
	EmA IgA, 2008-2009 data	Adults, Marsh 3 considered celiac	NR	NR	NR	NR	0.826	0.991
Vermeersch, 2012 ⁵⁴	tTG IgA, DGP IgG combined screen	Adults, Brand 1	NR	NR	NR	NR	0.897	0.933
	tTG IgA, DGP IgG combined screen	Adults, Brand 2	NR	NR	NR	NR	0.888	0.956
	tTG IgA	Adults, Brand 1	NR	NR	NR	NR	0.841	0.959
	tTG IgA	Adults, Brand 2	NR	NR	NR	NR	0.813	0.985
	DGP IgG	Adults, Brand 1	NR	NR	NR	NR	0.850	0.993
	DGP lgG	Adults, Brand 2	NR	NR	NR	NR	0.869	0.956
Wolf, 2014 57	tTG IgA, DGP IgG	Children without IgA deficiency	314	8 (30 grey zone)	2 (31 grey zone)	659	0.892	0.952
	tTG IgA, DGP IgG	Children with IgA deficiency	7	6 (11 grey zone	0	3	0.292	1.000

tTG = anti-tissue transglutaminase; IgA= immunoglobulin A; DGP = deamidated gliadin peptide antibodies; IgG= immunoglobulin G; EmA = endomysial antibodies; NICE = National Institute for Clinical Excellence; CI = confidence interval; grey zone = indeterminate results; NR = not reported.

<u>Video capsule endoscopy.</u> We identified two systematic reviews (SR) on the accuracy of video capsule endoscopy (VCE). A 2012 SR⁶³ by Rokkas and colleagues included six studies, three of which represented all studies included in the prior SR⁶⁴ by El-Matary on the topic. The Rokkas SR pooled five studies with a total of 166 subjects; sensitivity of 89% (95% CI: 82.0%, 94.0%) and specificity of 95% (95% CI: 89.0%, 99.0%) were reported. LR+ was 12.90 (95% CI: 2.89, 57.58), and LR- was 0.16 (95% CI: 0.10, 0.25).

Data are presented in Table 14 below. The Area under the Curve (AUC) of the weighted symmetric summary ROC curve was 0.95. The quality of this SR was rated moderate, as the publication made no mention of assessment of the quality of the included studies. In addition, patient ages and symptomatology were not described. No additional studies of the accuracy of VCE that met our inclusion criteria were identified.

Reference	# of Studies	# of Participants	Baseline Prevalence	Method for Pooling	Sensitivity	Specificity
El-Matary, 2009 ⁶⁴	3	107	58.88%	Simple pooling method with CIs computed by using the modified-Wald approach.	83% (95% Cl: 71- 90%)	95% (95% Cl: 88- 99.6%)
Rokkas & Niv, 2012 ⁶³	6	166	NR	Mantel Haenszel method	89% (95% Cl: 82- 94%)	95% (95% CI: 89-98%)

 Table 14. Video capsule endoscopy

CI = confidence interval; NR = not reported.

Key Question 1b. Intermediate Outcomes Such as Clinical Decisionmaking and Dietary Compliance

We identified a 2009 SR on factors associated with adherence to gluten-free diet in adult celiac disease patients.⁶⁵ The authors identified 38 studies published between January 1980, and November 2007. Studies were included regardless of whether adherence was a primary or secondary focus. The quality of the review was rated low according to AMSTAR criteria (see appendix). The rate of "strict" adherence ranged from 42% to 91% in the included studies and varied by definition of "strict" and whether adherence was measured by self-report or estimated via biological markers. Most factors investigated were socio-demographic; none of the 38 studies compared adherence by type of diagnostic test. Three included studies reported no statistical difference in adherence levels between patients whose celiac disease was detected via screening and those whose celiac disease was "symptom- detected." These three studies were CD patients.

We identified one additional relevant study, whose results conflict with those reported in the 2009 SR. This study, conducted in Israel, sheds light on clinician and patient decisionmaking after asymptomatic individuals screened positive. In 2007, researchers conducted a study on prevalence of CD by screening 1,571 healthy adult blood donors using the IgA-tTG (or IgG-tTG for IgA deficient.⁶⁶ The fifty-nine patients who tested positive were given their results and

counseled; 51 participated in a telephone interview about three years later. Only 30 had undergone biopsy; of these 30, ten were diagnosed with CD. Four of the ten strictly adhered to a gluten-free diet; the other six did not believe they had celiac disease, because they considered themselves asymptomatic. Four of these six reported having been told by their physicians (two gastroenterologists, two primary care physicians) to ignore the test results because there was no need for asymptomatic patients to modify their diet. Of the 29 who did not undergo biopsy, twelve consulted a physician, and nine of these twelve were advised against biopsy due to lack of symptoms. While this study provides interesting information, risk of bias is high (the sample size is small and the information on clinical and patient decisionmaking is self-reported rather than assessed via medical record) and applicability to the current U.S. situation is not certain.

We identified no other studies of how clinical decisions differ by method of patient diagnosis. The official guidance on clinical management of celiac disease does not differ after the initial diagnosis is finalized.⁵

Key Question 1c. Clinical Outcomes and Complications Related to CD

No studies reporting how clinical outcomes or complications differ by diagnostic method were identified. Differences due to false negative results are discussed in the results section for Key Question 4.

Key Question 1d. Patient-Centered Outcomes Such as QOL and Symptoms

No studies reporting how patient-centered outcomes differ by diagnostic method were identified. Differences due to false negative results are discussed in the results section for Key Question 4.

Key Question 2. Duodenal Biopsy Issues

Key Points

One very large retrospective national study found that in the U.S., adherence to American Gastroenterological Association duodenal biopsy protocol (4+ specimens) was worse at endoscopy suites with a higher volume of endoscopies with duodenal biopsy, while adherence was better at endoscopy suites with a higher number of gastroenterologists.

Three retrospective studies evaluating inter-observer variability in histological diagnosis of CD between different pathologists and clinical settings indicate that CD-related histological findings are underdiagnosed in community-based hospital- and practice settings when compared to academic settings.

Two previous SRs and several additional primary studies indicate that the number and location of biopsy specimens influence diagnostic findings of biopsy, and they recommend taking multiple specimens from different sites of the duodenum.

A 2013 SR of high quality on clinical response to gluten challenge indicates that a 3-month gluten challenge with a moderate-to-high dose (e.g., 15g daily) should be sufficient to diagnose the majority of CD patients; however, based on more recent data, the ACG recommends three grams daily for two weeks and six additional weeks if tolerable for adults.

Detailed Synthesis

Key Question 2a. Characteristics of Pathologists and Health Care Providers

One of the Key Questions for this systematic review is whether the likelihood of positive diagnosis with endoscopy with duodenal biopsy varies depending on the characteristic of pathologists or other medical staff (e.g. level of training or experience). Four studies addressing this issue were identified; results are displayed in Table 15. In their study of the influence of provider characteristics (e.g. procedure volume [defined by the number of endoscopies with duodenal biopsy performed] and the number of physicians in each endoscopy suite) on adherence to the American Gastroenterological Association's protocol (submissions of four or more specimens during duodenal biopsy), Lebwohl and colleagues (2013) found that adherence reduced with increasing procedure volume, but increased with increasing number of gastroenterologists working in an endoscopy suite.⁶⁷

Three studies investigated the inter-observer variability in the histological diagnosis of CD between different pathologists and clinical settings. In a study from the Netherlands in which the histological diagnoses of suspected pediatric CD patients were reviewed,⁶⁸ Mubarak et al. (2011) found a moderate inter-observer agreement (k=0.486) in the Marsh classification of the villous atrophy of biopsy specimens between the referring/originating pathologist and the study pathologist as well as a high inter-observer agreement (k=0.850) for CD diagnosis. A similar study by Arguelles-Grande et al. from a US center in 2012 found the inter-observer agreement in pathologists' diagnosis—based on biopsy interpretation—between the pathologist in the study hospital and the pathologist in the referring or originating hospital to be moderate (k=0.529, p<0.0001), leading to a 20% increase in CD diagnosis.⁶⁹ The same study examined interobserver agreement by type of pathology practice setting and found it ranged between 'verv good' when the study hospital was compared with other university hospitals (k=0.888) and 'moderate' when the study hospital was compared with community hospitals or commercial pathology laboratories (k=0.465 and k=0.419, respectively) (Arguelles-Grande et al. 2012). A study in Argentina that evaluated the accuracy of the histologic diagnosis of CD performed in a community clinical setting compared with that of an experienced academic center found a divergence of 46.3% in diagnosis and a poor agreement (k=0.16) between settings (Sanchez et al. 2007).⁷⁰ These studies support that CD-related histological findings are underdiagnosed in community-based hospital and practice settings.

Study (Author, Year)	Study Objective	Methodology/Data Dource	Sample Size/ Pop.	Results
Picarelli et al., 2014	Verified the correct and uniform application of Marsh–Oberhuber criteria, observing their reliability and diagnostic accuracy in CD diagnosis by testing the repeatability of histological evaluation of the same histological samples carried out by 5 different operators (histologists).	Evaluation of histological findings of duodenal biopsies by five different histologists not aware of patients' clinical data. (the most experienced histologist in CD, was used as standard for comparison)	66 active CD patients and 48 controls with no CD.	The strength of agreement was good/very good for Marsh–Oberhuber classification (Kappa statistic: 0.54- 0.78) as well as CD diagnosis (Kappa statistic: 0.78).
Lebwohl et al.,2013 ⁶⁷	Studied the influence of provider characteristics (procedure volume, defined by the number of endoscopies with duodenal biopsy performed, the number of physicians in each endoscopy suite, and the regional physician density) on adherence to standard of care (four or more specimens during duodenal biopsy)	National pathology database/ Multivariate analysis.	92,580 adults with potential CD	Reduced adherence was observed with higher procedure volume [odds ratio (OR) for each additional 100 procedures, 0.92; 95% CI, 0.88–0.97; P = 0.002]. An increased adherence was reported for gastroenterologists working at suites with higher numbers of gastroenterologists (OR for each additional gastroenterologist, 1.08; 95% CI, 1.04–1.13; P < 0.001).
Arguelles- Grande et al., 2012 ⁶⁹	Evaluated the agreement in biopsy interpretation (characteristic histological alterations of small bowel mucosa) between different pathology practice types (university hospitals, community hospitals, commercial pathology laboratories)	Retrospective review of biopsy slides of patients from referring centers.	102 suspected adult/ pediatric CD patients	Inter-observer agreement in the diagnosis of CD between study pathologist and the referring pathologist was moderate (k=0.529, p<0.0001). In addition, agreement ranged between 'very good' with other university hospitals (k=0.888) and 'moderate' with community hospitals or commercial laboratories (k=0.465 and k=0.419, respectively).
Mubarak et al.,2011 ⁶⁸	Determination of the inter-observer variability in the histological diagnosis of CD.	Retrospective review of histology slides of biopsy specimens by a single experienced pathologist	297 consecutive pediatric patients with suspected CD.	The inter-observer variability for the Marsh classification was found to be moderate with a Kappa value of 0.486 while the Kappa value for CD diagnosis was high (0.850) A total of 160 (53.9%) patients were original diagnosed with CD while 172 (57.9%) patients were diagnosed according to the second pathologist.
Sanchez et al., 2007 ⁷⁰	Evaluated the accuracy of the diagnosis of CD performed in the community clinical setting compared with that of an academic experienced center.	Retrospective review of original biopsy slides and reports (from community clinical setting).	70 consecutive adult CD patients	25 patients (46.3%) had a divergent diagnosis between the two practice settings (23 patients originally identified as CD and 2 diagnosed as non CD) (Kappa statistic: 0.16 -denoting poor agreement).

Table 15. Diagnosis by duodenal biopsy: Variation by pathologist and setting characteristics

CD = celiac disease; CI = confidence interval.

Key Question 2b. Specimens: Number and Location

Two nonsystematic reviews provided background information on biopsy strategy, i.e. specimen location and number. After reference-mining these two reviews and conducting electronic searches, we identified 19 studies relevant to this key sub-question. These studies are described narratively due to heterogeneity. Sensitivity and specificity were rarely discussed, as there was no "reference" test per se; the studies generally compared the number of positive diagnoses by strategy.

Several studies reported that the number of biopsy specimens influenced the likelihood of positive diagnosis, suggesting that multiple biopsy specimens should be taken from the duodenum in order to optimize diagnostic yield in both pediatric⁷¹⁻⁷⁴ and adult populations.⁷⁵⁻⁷⁷ One study in an adult population showed that the probability of a new diagnosis was increased when at least 4 specimens were submitted compared to when fewer than 4 specimens were taken (1.8% vs 0.7%; p < .0001).⁷⁵ Another retrospective study found that obtaining two duodenal biopsy specimens led to a confirmed diagnosis in 90% of cases, whereas obtaining at least four duodenal specimens led to confirmed diagnosis in all cases.⁷⁶ Another study suggests a minimum of three biopsies incorporating a duodenal bulb biopsy as essential to detect villous atrophy, with a five-biopsy regime useful for detecting the most severe lesion.⁷⁷

Most of the studies recommend obtaining biopsies from multiple sites in the duodenum, as the site/location from which duodenal specimens are taken could affect the likelihood of CD diagnosis. The evidence suggests that duodenal specimens should include the duodenal bulb and the distal duodenum for optimal diagnostic yield in adult/general ^{77-79 80} and pediatric populations. ^{71-74, 81, 82} In a suspected pediatric CD population, a study suggested taking biopsies from both the bulb and the second part of the duodenum, because mucosal changes may occur in only one site. ⁸³

Key Question 2c. Length of Gluten Challenge

Table 16 presents data on the length of time a patient needs to remain on a gluten-containing diet for accurate diagnosis. A SR (Bruins, 2013)^{84¹} addressed clinical response to gluten challenge among adult or pediatric patients with suspected or diagnosed CD previously consuming a gluten-free diet. This review was rated as high quality based on AMSTAR criteria (see appendix). The main focus of the review is serology; here we present results regarding intestinal histology. According to the authors, 51 to100 percent of children developed moderate to severe mucosal histological abnormalities within 2 to 3 months of gluten challenge, with data from two trials indicating that 59 to 78 percent of children may experience an increase in villous atrophy within 3 months of gluten challenge. Evidence from a small body of trials found no more than 50 percent of adult patients to be positive for EmA-IgA, tTG-IgA, or DGP-IgA/IgG antibodies within 6 weeks to 3 months of gluten challenge. Mucosal tTG-IgA deposits can appear in the majority of adult patients within 2 weeks of gluten challenge; however, two weeks or more of high-dose gluten challenge may be needed to detect small intestinal mucosal morphology changes for most patients. The ACG recommends an additional six weeks for patients who can tolerate gluten exposure. Bruins and colleagues concluded that a 3-month gluten challenge with a moderate-to-high dose (such as 15g daily, in accord with ESPGHAN guidelines) should be sufficient to diagnose the majority of CD patients, with combination testing of antibodies and mucosal histology potentially accelerating diagnoses. More recently, Leffler, 2013⁸⁵ conducted small gluten challenge study of adults with biopsy-proven celiac

disease. Patients were assigned a dose of either 3 or 7.5 grams of gluten daily. At 14 days, 89.5% of patients had sufficient villous atrophy to diagnose celiac disease via duodenal biopsy.

Table 16. Length of gluten challenge	, ⁵	84
--------------------------------------	----------------	----

Diagnostic Method	Population	Results			
EmA-IgA Antibodies	Children	Majority show positive levels within 3 months.			
	Adults	Few to none develop positive levels within 2 months.			
tTG-IgA Antibodies Children Majority show positive levels within 12 weeks.		Majority show positive levels within 12 weeks.			
	Adults	At most half have positive levels within 3 months.			
DGP-IgA/IgG Antibodies	Adults	At most half have positive levels within 4 weeks.			
HLA-DQ2 or -DQ8	Adults	Not affected by gluten intake			
	Children	Not affected by gluten intake			
Histology	Children	Most developed moderate-to-severe abnormalities within 2 to 3 months.			
	Adults	At least 2 weeks needed to detect changes for most patients; 6 additional weeks recommended for those who are able to continue without severe discomfort			

EmA = endomysial antibodies; IgA = immunoglobulin A; tTG = anti-tissue transglutaminase; IgG = immunoglobulin G.

Key Question 3. Specific Populations

Key Points

- A 2010 SR limited to studies of patients with GI symptoms reported pooled sensitivity of 90% (95% CI: 80.0%, 95.0%) and specificity of 99% (95% CI: 98.0%, 100.0%) for IgA EmA tests (8 studies) and pooled sensitivity of 89% (95% CI: 82.0%, 94.0%) and specificity of 98% (95% CI: 95.0%, 99.0%) for IgA tTG tests. These results are similar to those presented for Key Question 1, which included patients with various symptoms of CD.
- Only one study of general population screening met the inclusion criteria. Sensitivities were 100% and 85.7% for tTG IgA and EmA IgA, respectively. Specificities were 97.4% and 99.0%, respectively, in this high quality study.
- Two low quality studies provided data that allowed calculation of accuracy of serology in patients with iron deficiency. Studies were conducted in the Middle East; applicability to the U.S. is uncertain.
- Two high quality studies reported accuracy in children with Type 1 diabetes. These studies were conducted in Iraq and the Czech Republic with small samples. Applicability to the U.S. is uncertain.
- No studies provided test accuracy data on patients with other auto-immune diseases, Turner's syndrome, or Trisomy 21.
- One high quality study compared the accuracy of the ESPGHAN algorithm (combining tTG IgA and EmA IgA) among subjects with family history, Type 1 diabetes, and CD symptoms. Specificity was much higher in those presenting with symptoms.
- Two large moderate quality studies found both tTG and DGP tests less sensitive in adults than in children.

No studies reported accuracy by race, ethnicity, or SES.

Two small studies of the accuracy of new combination tests in IgA-deficient patients were published in 2014; results were inconsistent.

Detailed Synthesis

Key Question 3a. Symptomatic Patients Versus Nonsymptomatic Individuals at Risk

Two SRs reported on diagnostic test accuracy in individuals with symptoms of CD. In 2009, Ford and colleagues published an SR on yield of diagnostic tests in patients with Irritable Bowel Syndrome (IBS).⁸⁶ The goal was to estimate prevalence of celiac disease in unselected adults with IBS; test accuracy was not a primary outcome. The authors included 14 studies; 54 percent of the 4,204 individuals met diagnostic criteria for IBS. Although sensitivity and specificity were not reported, the authors computed odds ratios for positive diagnostic method. Pooled odds ratios for celiac disease in subjects with IBS versus controls were 3.40, (95% CI [1.62, 7.13]) for IgA EmA; 2.94, 95% CI (1.36, 6.35) for IgA tTG; and 4.34, (95% CI [1.78, 10.60]) for biopsy. Prevalence was estimated at 4.0 percent, 1.65 percent, and 4.34 percent by IgA EmA, IgA tTG, and biopsy, respectively.

In 2010, van der Windt and colleagues published an SR on performance of diagnostic tests in patients with abdominal symptoms typical of celiac disease.⁸⁷ Studies where 50 percent or more of participants reported gastrointestinal (GI) symptoms were included. Eight such studies that used IgA EmA were pooled, resulting in estimates of 90.0% (95% CI: 80.0%, 95.0%) for sensitivity and 99.0% (95% CI: 98.0%, 100%) for specificity. Pooled results of seven studies on IgA tTG estimated sensitivity at 89.0% (95% CI: 82.0%, 94.0%) and specificity at 98.0% (95% CI: 95.0%, 99.0%).

The quality of both SRs was moderate according to AMSTAR criteria. The Ford (2009) SR did not report quality assessment of included studies. Neither the Ford ⁸⁶ nor the van der Windt⁸⁷ SRs included lists of excluded studies. (Full AMSTAR criteria for all SRs are presented in Appendix E.)

Only one study of general population screening met the inclusion criteria.⁵⁶ This study presented data for 1,000 randomly selected Swedes. Sensitivity was 100% and 85.7% for tTG IgA and EmA IgA, respectively. Specificity was 97.4% and 99.0%, respectively.

Only one study compared accuracy in symptomatic versus asymptomatic individuals at risk for celiac disease. Nevoral, 2013⁸⁸ studied the accuracy of the new ESPGHAN guideline (combining tTG IgA and EmA IgA tests) in 32 first degree relatives, 60 patients with Type 1 diabetes, and 187 subjects presenting with symptoms of CD in the Czech Republic. Specificity was lower in the asymptomatic subjects at risk: 0.70 for those with family history and 0.64 for those with Type 1 diabetes, compared to 0.85 for subjects presenting with symptoms. Sensitivity was 0.81 for those with family history, 0.93 for those with diabetes, and 0.76 for symptomatic subjects. This is the only study that provided accuracy results specifically for subjects with a family history.

Two individual studies provided test accuracy data for patients with iron deficiency, a common symptom of CD. Data are presented in Table 17. One study conducted in Iran reported sensitivity of 0.38 and specificity of 0.97 for IgA tTG.³⁹ The other study,³⁴ conducted in Turkey, reported sensitivity of 1.00 and specificity of 0.99 for IgA EmA and sensitivity of 0.33 and specificity of 0.96 for IgG EmA. The primary goal of the first study was to assess accuracy, while the goal of the latter study was prevalence estimation. In both publications, it was unclear whether interpretation of the index test and reference test (biopsy) were blinded; the interval between these tests was also unclear.

	Test Type	True Positive	False Negative	False Positive	True Negative	Sensitivity	Specificity
Emami, 2012 ³⁹	tTG IgA	5	8	4	113	0.38	0.97
Cekin, 2012 ³⁴	EmA IgA	6	0	1	77	1.00	0.99
	EmA IgG	2	4	3	75	0.33	0.96

Table 17. Accuracy data for persons with iron deficiency

tTG = anti-tissue transglutaminase; IgA = immunoglobulin A; EmA = endomysial antibodies; IgG = immunoglobulin G.

Two studies provided accuracy data in patients with Type 1 diabetes. Data are displayed in Table 18 below. Mansour, 2011⁴² reported on use of IgA tTG, IgG tTG, and IgA EmA tests in 62 adults and children with Type 1 diabetes in Iraq. The study's primary goal was to assess prevalence of asymptomatic CD in persons with Type 1 diabetes. IgA tests had higher accuracy than the IgG test, as displayed below. As noted above, Nevoral (2013)⁴⁴ followed the recent ESPGHAN guidelines (tTG and EmA test) with 60 children and adolescents with Type 1

diabetes in the Czech Republic and reported sensitivity of 0.93 and specificity of 0.64 for the two tests combined. Both studies were rated high quality according to QUADAS 2 criteria.

	Test Type	True Positive	False Negative	False Positive	True Negative	Sensitivity	Specificity
Mansour,2011 ⁴²	tTG IgA	5	2	4	51	0.71	0.93
	tTG IgG	4	3	4	51	0.57	0.93
	EmA IgA	5	2	2	53	0.71	0.96
Nevoral, 2013 ⁴⁴	tTG IgA, EmA IgA					0.93	0.64

Table 18. Accuracy data for persons with type 1 diabetes

tTG = anti-tissue transglutaminase; IgA = immunoglobulin A; IgG = immunoglobulin G; EmA = endomysial antibodies.

We found no studies that provided accuracy data on patients with other auto-immune diseases, Turner's syndrome, or Trisomy 21. Although cirrhosis of the liver is not a risk factor for CD, one study of prevalence in patients with biopsy proven cirrhosis allowed calculation of sensitivity and specificity. Wakin-Fleming, 2014 ⁵⁵ conducted a high quality prospective cohort study that allowed comparison of IgA EmA (serum dilution >=1/10) to IgA tTG (> 20 U) in 204 cirrhosis patients. Based on biopsy results, five patients were diagnosed with celiac disease. IgA EmA had both sensitivity and specificity of 100%, while sensitivity was 100% and specificity was 96% for IgA tTG. This study had low risk of bias despite not being designed as an accuracy study.

Key Question 3b. Adults (Age 18 and Over) Versus Children and Adolescents

Studies that address this comparison are discussed in section c, below.

Key Question 3c. Children Under Age 24 Months Versus Older Children

Most studies reported mean age or age range of subjects, whereas a few simply described their patients as "adults" or "children." Accuracy data could not be pooled by age group due to heterogeneity of populations, varying definitions of adult, and missing age data in some studies.

Three studies compared the accuracy of tests among age groups. These studies are important to highlight because in each study the populations are fairly homogenous, and each patient received the same diagnostic tests in terms of assay and threshold level. Data from these three studies are displayed in Table 19 below. As described earlier, Mozo $(2012)^{43}$ compared the accuracy of IgA and IgG DGP tests in patients aged six months to 74 years. In children age seven or younger, the IgG tests had the same accuracy as IgA tests. In patients over age seven, the IgG tests had higher specificity and positive predictive value but lower sensitivity and negative predictive value. Olen $(2012)^{45}$ compared accuracy of tTG and DGP tests in 537 children and adolescents. In children with normal IgA who were younger than two years old (N = 71), sensitivity and specificity were 96.0% and 98.0%, respectively. Sensitivity of a combined IgA/IgG DGP test was 100%, and specificity was 31% in children younger than two. As presented below, these tests were less accurate in the entire sample, which ranged in age from one to eighteen years. (Accuracy results were not presented separately for patients over 24 months old.) Finally, Vermeersch (2010)⁵² compared accuracy in adults (over age 16) versus

children as part of a study comparing IgG DGP tests with IgA tTG screens. Using a case-control design, they compared accuracy of three IgG DGP tests, three IgA and two IgG anti-tTG assays, and one IgA DGP screen in 827 patients. Results are presented below; all tests reported lower sensitivity in adults than in children.

	Age Category	True Positive	False Negative	False Positive	True Negative	Sensitivity	Specificity
tTG lgA							
Mozo, 2012 ⁴³	<=7 yrs old	40	3	2	37	0.93	0.95
	>7 yrs old	49	8	4	57	0.86	0.93
Olen, 2012 ⁴⁵	<=18 yrs old	259	17	36	218	0.94	0.86
	<2 yrs old	25	1	1	40	0.96	0.98
Vermeersch, 2010 ⁵²	<16 yrs old	27	1	11	189	0.96	0.95
	<16 yrs old	27	1	3	197	0.96	0.99
	<16 yrs old	27	1	22	178	0.96	0.89
	>=16 yrs old	49	9	27	514	0.85	0.95
	>=16 yrs old	45	13	9	532	0.78	0.98
	>=16 yrs old	46	12	39	502	0.79	0.93
tTG lgG							
Vermeersch, 2010 ⁵²	>=16 yrs old	31	27	4	537	0.53	0.99
	>=16 yrs old	17	41	9	532	0.29	0.98
	<16 yrs old	21	7	10	190	0.75	0.95
	<16 yrs old	16	12	2	198	0.57	0.99
DGP lgA							
Mozo, 2012 ⁴³	<=7 yrs old	41	2	1	38	0.95	0.97
	>7 yrs old	55	2	3	58	0.97	0.95
Olen, 2012 ⁴⁵	<=18 yrs old	172	16	164	56	0.91	0.26
	<2 yrs old	24	0	31	14	1.00	0.31
Vermeersch, 2010 ⁵²	>=16 yrs old	40	18	3	538	0.69	0.99
	<16 yrs old	26	2	3	197	0.93	0.99
DGP lgG							
Mozo, 2012 ⁴³	<=7 yrs old	43	0	0	39	1.00	1.00
<u></u>	>7 yrs old	52	5	1	60	0.91	0.98
Vermeersch, 2010 ⁵²	>=16 yrs old	40	18	3	538	0.69	0.99
	>=16 yrs old	46	12	2	539	0.79	1.00
	>=16 yrs old	47	11	11	530	0.81	0.98
	>=16 yrs old	42	16	12	529	0.72	0.98
	<16 yrs old	26	2	3	197	0.93	0.99
	<16 yrs old	26	2	3	197	0.93	0.99
	<16 yrs old	26	2	6	194	0.93	0.97
	<16 yrs old	27	1	9	191	0.96	0.96

Table 19. Accuracy results by age

tTG=anti-tissue transglutaminase; IgA = immunoglobulin A; IgG = immunoglobulin G; DGP = deamidated gliadin peptide (DGP) antibodies.

Key Question 3d. Demographics, Including Race, Genetics, Geography, SES

No studies reported accuracy by race, ethnicity, or SES. Many studies were conducted outside the U.S., most often in Europe or the Middle East. However, even though we identified studies conducted in one country only, and even if that country has a homogeneous ethnic population, results may have little applicability to persons of the same racial or ethnic group living in the U.S.

Key Question 3e. Patients With IgA Deficiency

Two studies of accuracy in IgA-deficient patients met our inclusion criteria. Beinvenu (2014) ³³ studied a multi-analytic lateral-flow immunochromatographic assay (CD-LFIA) based on both IgA DGP and IgG DGP in 45 IgA-deficient children presenting with symptoms of CD or having risk factors. The study was retrospective —the new test was used on stored blood samples of patients who previously underwent biopsy. Researchers were blinded to those results. The authors reported a sensitivity of 100% and specificity of 89.2%. Wolf (2014) ⁵⁷ conducted a case-control study of adding an IgG DGP test to an IgA tTG test; 27 of their 1,071 subjects were IgA deficient. From the data presented in their article, we calculated 100% specificity in the children with IgA deficiency. However, sensitivity in this group was only 29%.

In addition, Dutta (2010)³⁸ reported on 92 consecutive patients who presented with symptoms of possible celiac disease at a clinic in Vellore, India. Eighteen patients (19.5%) were diagnosed with CD; 14 had positive serology. Sensitivity and specificity of IgG tTG tests were 77.8% and 89.1%, respectively. It was unclear why IgG tests were the only serology tests conducted and whether patients were IgA deficient. The study had no major flaws.

Key Question 3f. Patients Who Previously Tested Negative for CD

One study of test accuracy in patients with previously negative serology was identified. As part of a larger study, Sugai (2010)⁴⁹ retrospectively conducted IgA and IgG DGP tests on 17 IgA tTG-negative serum samples from patients with indications of celiac disease (either intestinal enteropathy or dermatitis herpetiformis). In the IgA tTG-negative patients, detection of "gluten sensitivity" increased 31.6% when an IgA tTG/ IgA DGP dual screen was applied and 26.3% when a dual IgA and IgG DGP screen was used. The sensitivity and specificity of the tTG/DGP dual screen were 35.7% and 100% in this group, whereas those rates were 42.9% and 100% for dual DGP screen. However, the results may be biased: Five of the 22 originally identified IgA-deficient patients refused biopsy, leaving 17 for this small retrospective analysis.

Key Question 4. Adverse Effects

Key Points

- A systematic review of 150 studies on VCE not specific for CD found a capsule retention rate of 1.4%; in three studies specific to CD, the rate ranged from 0.9% to 4.6%.
- No studies on safety of upper GI endoscopy and / or duodenal biopsy for diagnosis of CD were identified. According to the American Society for Gastrointestinal Endoscopy

(ASGE), infection from upper GI endoscopy is very rare, as is bleeding unless a polyp is removed during the procedure.

- Two studies reporting sequelae in patients with positive EmA serology but normal biopsy indicated that 30 percent to 50 percent of these patients are diagnosed with CD after a gluten-free diet or gluten challenge.
- One study of 34 children with intestinal villous atrophy and simultaneous negative EmA-IgA tests found 2 infants to have confirmed CD after 6-10 years of iterative cycles of gluten challenges and gluten-free diet.

Detailed Synthesis

Direct Adverse Events Associated With Invasive Methods of Diagnosing Celiac Disease

"Invasive " methods used to diagnose CD include upper GI endoscopy with duodenal biopsy, often referred to as the "gold standard," as well as VCE for patients who wish to avoid biopsy. A systematic review on the general safety of these procedures was beyond the scope of this small project; however, due to the dearth of data specific to their use in CD diagnosis, we review evidence on the safety of their use to provide some context. Applicability to patients with suspected CD is uncertain.

The main adverse event from VCE is capsule retention, which could cause acute small bowel obstruction or the need for surgical removal.⁸⁹ According to International Center for Clinical Excellence (ICCE) consensus, capsule retention may occur in patients with refractory celiac disease who may have strictures. Three studies reporting retention rates with VCE used for CD diagnosis were identified; rates ranged from 0.9% to 4.6%.⁹⁰⁻⁹² Study characteristics are displayed in Table 20. Gomez and colleagues (2013), in a comparison of patients aged 80 years or older with those under age 80, found a similar frequency in occurrence of capsule retention and concluded that capsule endoscopy can be performed safely in the elderly population.

A 2010 systematic review of VCE by Liao and colleagues in populations suspected of various small intestine pathologies found a pooled retention rate of 1.4% in 104 prospective studies and 46 retrospective studies published between 2000 and 2008.⁹³ The rates of capsule retention appear to depend on VCE indication; pooled retention rates of 1.2 percent, 2.6 percent, and 2.1 percent for obscure gastrointestinal bleeding, Crohn's disease, and neoplastic lesions indications were estimated from 47, 23, and 12 studies respectively. There were no studies in this systematic review that investigated capsule retention when VCE was used specifically to evaluate patients with suspected CD. This systematic review was rated as low quality, as a listing or bibliography of the included articles was not provided.

Only two studies on biopsy to diagnose CD were identified; they estimated the amount of absorbed radiation due to X-ray fluoroscopy-guided small intestine biopsies in a pediatric population.^{94, 95} These studies were excluded, as fluoroscopy has been replaced by video systems. No studies on the safety of upper GI endoscopy when used specifically for diagnosis of CD were identified. According to ASGE,⁹⁶ infection from upper GI endoscopy is very rare, as is bleeding, unless a polyp is removed during the procedure.

We employed the McHarm scale to evaluate the quality of the VCE adverse event studies. Data are displayed in Table 21. All used precise definitions to define harms (generally, capsule retention was defined as a situation in which a capsule endoscope remains in the digestive tract for a minimum of two weeks, which may necessitate surgical intervention in order to retrieve the capsule endoscope.) No studies explicitly mentioned severe or serious adverse events. Adverse events were actively collected or ascertained in all studies. All studies reported the number and full spectrum of adverse events that occurred; the total number of study participants affected by harms; and the type of analyses conducted for adverse events data.

Table 20. Adverse events, video capsule endoscopy used for cellac disease diad
--

Study (Author, Year)	Study Design (Claims Data Analysis, RCT, Survey, Etc.)	Diagnostic Procedures Assessed (Endoscopy, Serology Test, Etc.)	Sample Size	Population	Event	Results
Vere et al.,2012 ⁹⁰	Retrospective study	Video Capsule endoscopy	43	Adults	Capsule retention (Slower intestinal transit time)	Capsule retention in two patients (4.6%)
Gomez,2013 ⁹¹	Retrospective matched cohort study (>80yr old patients vs. <80 yr old patients)	Video capsule endoscopy	780	Adults	Capsule retention	Capsule retention occurred at a similar frequency in patients age >80yrs (1.0%) compared to those <80 yrs (0.9%).
Atay et al.,2009 ⁹²	Retrospective chart review	Video capsule endoscopy	207	Children	Capsule retention	Capsule retention occurred in 3 of 207 procedures (1.4%)

RCT = randomized controlled trial.

	McHarm Items	Vere et al. 2012 ⁹⁰	Gomez 2013 ⁹¹	Atay et al. 2009 ⁹²
1	Were the harms PRE-DEFINED using standardized or precise definitions?	Yes	Yes	Yes
2	Were SERIOUS events precisely defined, if mentioned?	NA	NA	NA
3	Were SEVERE events precisely defined, if mentioned?	NA	NA	NA
4	Were the number of DEATHS in each study group specified OR were the reason(s) for not specifying them given?	No	No	Yes
5	Was the mode of harms collection specified as ACTIVE?	Yes	Yes	Yes
6	Was the mode of harms collection specified as PASSIVE?	No	No	No
7	Did the study specify WHO collected the harms?	No	No	Yes
8	Did the study specify the TRAINING or BACKGROUND of who ascertained the harms?	No	No	Yes
9	Did the study specify the TIMING and FREQUENCY of collection of the harms?	No	Yes	Yes
10	Did the author(s) use STANDARD scale(s) or checklist(s) for harms collection?	NA	NA	NA
11	Did the authors specify if the harms reported encompass ALL the events collected or a selected SAMPLE?	Yes	Yes	Yes
12	Was the NUMBER of participants that withdrew or were lost to follow-up specified for each study group?	NA	NA	NA
13	Was the TOTAL NUMBER of participants affected by harms specified for each study arm?	Yes	Yes	Yes
14	Did the author(s) specify the NUMBER for each TYPE of harmful event for each study group?	Yes	Yes	Yes
15	Did the author(s) specify the type of analyses undertaken for harms data?	Yes	Yes	Yes

Table 21. Quality of adverse events studies

NA = not applicable.

Sequelae of False Positives, False Negatives, and Indeterminate Results

Three studies reported sequelae in patients whose serology and biopsy results were discordant. All were conducted in Europe and involved EmA test results. Kurppa (2012)⁹⁷ evaluated 405 consecutive EmA positive children and adults at a university hospital who were referred by physicians for suspicion of CD or who participated in population-based research studies. Of these, 40 patients had low and 17 had high EmA and tTG serum antibody values without simultaneous villous atrophy at baseline. Eventually 12 (30 percent) in the low-titer group and 8 (47 percent) in the high-titer group were diagnosed with CD based on villous atrophy on a gluten-challenge, while 17 (43 percent) and 8 (47 percent) were diagnosed with CD based on villous atrophy on a gluten-challenge, while 17 (units provided the disappearance of early mucosal changes during a gluten-free diet. Unfortunately, length of follow-up was not report, so no data are provided on the length of gluten challenge or gluten-free diet.

A 2014 study by the same authors⁹⁸ randomized 40 EmA-positive yet otherwise asymptomatic adults to either a gluten-free diet or gluten-challenge.⁹⁸ These 40 patients were identified from 3,031 consecutive individuals screened for CD at a university hospital in Finland. Of these 40 participants, two in the gluten-free diet group and two in the gluten-challenge group had positive EmA but no villous atrophy. Of these 4 participants, one in each group were EmAnegative and had increased villous atrophy ratios after one-year follow-up; however, the glutenfree diet participant demonstrated improved gastrointestinal symptoms and self-perceived general health, whereas the gluten-challenge participant worsened in both of these areas.

The final study, by Kwiecien (2005), documented sequelae of discordant results via a retrospective analysis of data from 1985-2000 on 34 children with subtotal or total intestinal villous atrophy with simultaneously negative IgA EmA tests.⁹⁹ This group included all children with discordant results from over 1,300 consecutive diagnoses. Of the 34 children, 15 completed diagnostic follow-up with three biopsies, and two of these had confirmed CD with repeated positive IgA EmA tests. One 13 month-old with subtotal villous atrophy had a second biopsy after three years of a gluten-free diet, which revealed normal intestinal mucosa. Then, after a 12month gluten challenge, positive IgA EmA occurred simultaneously with mucosal relapse and clinical symptoms of malabsorption syndrome, leading to a CD diagnosis at 5 years, 9 months. The other child, with total villous atrophy at 11 months, had a gluten-free diet for 2.5 years, then underwent a gluten-challenge for 2 years; at this point, subtotal villous atrophy was discovered, yet EmA-IgA tests were negative. Another two-year gluten-free diet led to normalization of intestinal mucosa, while a 3-year gluten-challenge led to positive IgA EmA and subtotal villous atrophy, and thus a CD diagnosis at 10 years, 3 months. Although these examples shed light on clinical pathways, this study was conducted prior to the availability of other serological tests. Thus, the results may have little applicability to current clinical practice.

Discussion

Key Findings and Strength of Evidence

The key findings and strength of evidence are summarized in Table 22. Additional details on strength of evidence ratings are provided as Appendix F.

Торіс	EPC Conclusions and Strength of Evidence	Prior Systematic Reviews	Additional Findings From EPC
Key Question 1: Accuracy of IgA tTG	High: IgA tTG tests have excellent sensitivity and specificity.	A 2010 meta-analysis that pooled 12 studies found a sensitivity of 93.0% (95% Cl, 91.2% to 94.5%) and specificity of 96.5% (95% Cl, 95.2% to 97.5%). A 2012 meta-analysis restricted to 5 studies of point-of-care tests in children reported sensitivity and specificity of 96.4% (95% Cl, 94.3% to 97.9%) and 97.7% (95% Cl, 95.8% to 99.0%), respectively.	Sixteen studies were published after the SRs were pooled. Excluding data for threshold levels higher than used in clinical practice, sensitivity was 92.5% (95% CI, 89.7% to 94.6%) and specificity was 97.9% (95% CI, 96.5% to 98.7%). LR+ was 40.19 and LR- was 0.08. PPV was 89.4%, while NPV was 99.0%.
Key Question 1: Accuracy of IgA EmA	High: IgA EmA tests have lower sensitivity but equal specificity to IgA tTG tests.	A 2009 SR including 23 studies found sensitivity ranging from 68% to 100%, while specificity ranged from 77% to 100%; pooling was not performed. A 2012 SR included 11 studies in children; sensitivity ranged from 82.6% to 100% and pooled specificity was 98.2% (95% CI, 96.7% to 99.1%).	Seven studies were published after the SRs were pooled. Sensitivity was 79.0% (95% CI, 71.0% to 86.0%) and specificity was 99.0% (95% CI, 98.4% to 99.4%) after excluding data points where Marsh Grade I and II villous atrophy was classified as CD (not standard practice). LR+ was 65.98 and LR- was 0.21. PPV was 78.9%; NPV was 99.1%.
Key Question 1: Accuracy of IgA DGP	High: IgA DGP tests are not as accurate as IgA tTG tests.	A 2010 SR pooled 11 studies on accuracy in all ages; sensitivity was 87.8% (95% CI, 85.6% to 89.9%), while specificity was 94.1% (95% CI, 95.2% to 97.5%). LR+ was 13.33, while LR- was 0.12. A 2012 SR reviewed 3 of those studies that included only children: sensitivities ranged from 80.7% to 95.1% (not pooled) and pooled specificity was estimated at 90.7% (95% CI, 87.8% to 93.1%).	One new study reported sensitivity of 97.0% and specificity of 90.7% in symptomatic adults and children at 1 clinic, while another reported both sensitivity and specificity of 96% in a similar population.
Key Question 1: Accuracy of IgG DGP	Moderate: IgG DGP tests are not as sensitive as IgA tTG tests in non–IgA-deficient patients.	A 2013 SR of 7 studies of non–IgA-deficient adults reported sensitivity of 75.4% to 96.7% and specificity of 98.5% to 100%. A 2012 SR of 3 studies in non– IgA-deficient children reported sensitivities of 80.1% to 98.6% and specificities of 86.0% to 96.9%. Authors did not pool data.	One study reported sensitivity of 95.0% and specificity of 99.0% in 200 non–IgA-deficient subjects of all ages.
Key Question 1: Accuracy of HLA-	High: HLA tests can be used to	No SRs of the accuracy of testing for HLA-DQ2 or DQ8 were identified. Based on studies from which	Two studies were identified on the accuracy of HLA testing. A large 2013 prospective cohort found that
DQ2 or DQ8	rule out CD with close to	sensitivity (but not specificity) could be calculated,	HLA testing had a sensitivity of 100% and specificity

Table 22. Summary of findings and strength of evidence

Торіс	EPC Conclusions and Strength of Evidence	Prior Systematic Reviews	Additional Findings From EPC
	100% sensitivity.	the American College of Gastroenterology estimated the NPV of the HLA-DQ2/DQ8 combination test at over 99%.	of 18.2%. A 1999 cohort also reported sensitivity of 100%, while specificity was 33.3%.
Key Question 1: Accuracy of algorithms	Insufficient: Strength of evidence is insufficient to determine comparative accuracy of different algorithms in specific populations.	No SRs of the accuracy of algorithms were identified.	Nine studies of algorithms were identified; all used tTG tests. Adding an EmA test to a tTG test resulted in increased specificity, with either no change or a slight decrease in sensitivity. Adding a DGP test to a tTG test resulted in increased sensitivity but decreased specificity. However, the increase in accuracy compared with individual tests was rarely clinically significant. The sensitivity and specificity results varied widely, populations were diverse, and the evidence base had high heterogeneity.
Key Question 1: Accuracy of VCE	Moderate: VCE has very good sensitivity and excellent specificity.	A previous SR of moderate quality on the accuracy of VCE pooled 6 studies, and estimated sensitivity at 89.0% (95% CI, 82.0% to 94.0%) and specificity at 95.0% (95% CI, 89.0% to 99.0%). LR+ was 12.90 and LR- was 0.16.	No additional studies met our inclusion criteria.
Key Question 1: Intermediate outcomes	Insufficient: Strength of evidence is insufficient regarding how method of diagnosis affects adherence.	A previous SR of low quality (3 studies) reported no statistical difference in adherence levels between patients diagnosed via screening and those diagnosed because they were symptomatic. Association between diagnostic test type and adherence was not addressed.	In 1 study on blood donors in Israel who tested positive for IgA tTG (or IgG tTG if IgA deficient), only 4 of 10 patients with asymptomatic biopsy-proven CD adhered to a gluten-free diet; the other 6 patients did not believe they had CD, and 4 of those were told by physicians that asymptomatic patients did not need to modify their diets.
Key Question 1: Clinical outcomes and complications	Insufficient: Strength of evidence is insufficient regarding how method of diagnosis affects clinical outcomes and complications.	No prior SRs on this topic were identified.	No studies on this topic were identified.

Торіс	EPC Conclusions and Strength of Evidence	Prior Systematic Reviews	Additional Findings From EPC
Key Question 1: Patient- centered outcomes such as quality of life	Insufficient: Strength of evidence is insufficient regarding how method of diagnosis affects patient-centered outcomes such as quality of life.	No prior SRs on this topic were identified.	No studies on this topic were identified.
Key Question 2: Biopsy and provider characteristics	Moderate: Physician adherence to biopsy protocol decreases with volume performed per endoscopy suite and increases with number of gastroenterologists per endoscopy suite.	No SRs on this topic were identified.	One very large high-quality national retrospective study found reduced physician adherence to the American Gastroenterological Association's duodenal biopsy protocol (4+ specimens) with higher procedure volume per endoscopy clinic. The OR for each 100 additional procedures was 0.92 (95% CI, 0.88 to 0.97). Adherence increase for each additional gastroenterologist per endoscopy suite was OR 1.08 (95% CI, 1.04 to 1.13).
Key Question 2: Biopsy and pathologist characteristics	Moderate: CD-related histological findings are underdiagnosed in community settings when compared with academic settings.	No SRs on this topic were identified.	Three retrospective studies reported low interobserver agreement between pathologists in community vs. academic settings, with significantly lower accuracy in community settings. Kappa statistics range from 0.16 to 0.53.
Key Question 2: Biopsy specimens— number and location	High: Increasing the number and location of biopsy specimens increases diagnostic accuracy.	No SRs addressed how the number and location of biopsy specimens influence diagnostic findings of biopsy.	Nineteen studies reported that increasing the number and location of biopsy specimens increased the likelihood of diagnosis and diagnostic yield by 25% to 50% in both pediatric and adult populations.
Key Question 2: Biopsy and length of time ingesting gluten	Moderate: A minimum 2-week gluten intake is necessary to induce intestinal changes necessary for diagnosing adults via duodenal biopsy. Low: A 2–3 month diet containing gluten may be necessary to diagnose CD in children via biopsy; strength is lower due to fewer available studies and inconsistent findings.	A previous SR of high quality on clinical response to gluten challenge indicates that 2 weeks of a moderate to high dose (e.g., 15g daily) is sufficient to cause enough intestinal changes to diagnose adults via duodenal biopsy. This same SR reports that for children, 2 to 3 months may be needed.	One small study reported that 3 grams of gluten per day for 2 weeks induces intestinal atrophy sufficient to diagnose CD in 89.5% of adults.

Торіс	EPC Conclusions and Strength of Evidence	Prior Systematic Reviews	Additional Findings From EPC
Key Question 3: Symptomatic patients vs. nonsymptomatic individuals at risk	High: EmA and tTG tests have excellent sensitivity and specificity in patients with GI symptoms. Insufficient: How accuracy of serological tests differs between patients with risk factors such as iron deficiency or type 1 diabetes and the general symptomatic population could not be determined.	A 2010 SR including only studies of patients with GI symptoms reported pooled sensitivity of 90% (95% CI, 80.0% to 95.0%) and specificity of 99% (95% CI, 98.0% to 100.0%) for IgA EmA tests (8 studies), and pooled sensitivity of 89% (95% CI, 82.0% to 94.0%) and specificity of 98% (95% CI, 95.0% to 99.0%) for IgA tTG tests. No SRs were identified that compared test accuracy in patients with specific symptoms and asymptomatic individuals at risk.	One high-quality study compared the accuracy of the ESPGHAN algorithm (combining tTG IgA and EmA IgA) among subjects with family history, type 1 diabetes, and CD symptoms. Specificity was much higher in those with symptoms. Two small studies provided data that allowed calculation of accuracy in patients with iron deficiency, and 2 provided accuracy data for patients with type 1 diabetes. However, the studies were conducted in the Middle East and Eastern Europe; applicability to the United States is uncertain.
Key Question 3: Children vs. adults	Low: tTG and DGP tests are less sensitive in adults than children. DGP is more accurate than tTG in children under age 24 months.	No SRs assessing how test accuracy differs by age were identified. Regarding IgG DGP, one SR reported only on studies of adults, while another reported only on studies of children. A 2013 SR of 7 studies of non–IgA-deficient adults reported sensitivity of 75.4% to 96.7% and specificity of 98.5% to 100%. A 2012 SR of 3 studies in non–IgA-deficient children reported sensitivities of 80.1% to 98.6% and specificities of 86.0% to 96.9%.	Two large moderate-quality studies reported that both tTG and DGP tests were less sensitive in adults (range, 29% to 85%) than children (range, 57% to 96%). One study reported sensitivity of 96% and 100% for IgA tTG and IgA DGP, respectively, for children under age 24 months, while specificity was 98% and 31%, respectively. Accuracy was significantly lower for both tests in older children and adolescents.
Key Question 3: Demographics, including race	Insufficient: There was insufficient evidence to estimate the accuracy of diagnostic methods by demographic characteristics.	No SRs on this topic were identified.	No studies reported accuracy by race, ethnicity, or socioeconomic status.
Key Question 3: Patients with IgA deficiency	Insufficient: There was insufficient evidence to estimate the accuracy of diagnostic methods in IgA-deficient patients.	No SRs on this topic were identified.	Two small studies of the accuracy of new combination tests (IgA DGP + IgG DGP combo, IgA tTG + IgG DGP combo) in IgA-deficient patients were published in 2014; results were inconsistent.
Key Question 3: Patients who previously tested negative for CD	Insufficient: There was insufficient evidence to estimate the accuracy of diagnostic methods in patients who previously tested negative for CD.	No SRs on this topic were identified.	A very small study (N = 17) found that patients with biopsy-verified CD who tested negative on IgA tested positive using IgA DGP or IgG DGP.

Торіс	EPC Conclusions and Strength of Evidence	Prior Systematic Reviews	Additional Findings From EPC
Key Question 4: Direct adverse	High: The rate of capsule retention	No SRs contained safety data on VCE used specifically for CD diagnosis. An SR of VCE not	In 3 studies specific to CD, the capsule retention rate ranged from 0.9% to 4.6%.
events—VCE	is less than 5%.	specific to CD found a capsule retention rate of 1.4% in 150 studies.	
Key Question 4: Direct adverse events— endoscopy with duodenal biopsy	Moderate: Adverse events during upper GI endoscopy are rare.	No SR contained safety data on upper GI endoscopy or duodenal biopsy when used specifically to diagnose CD. A review on upper endoscopy in general found infection very rare and bleeding very rare (1.6 per 1,000) unless a polyp is removed.	No studies specific to diagnosis of CD were identified.
Key Question 4: Indirect adverse events—false negatives or positives	Insufficient: Strength of evidence is insufficient regarding the impact of misdiagnosis.	No SRs on the impact of misdiagnosis of CD were identified.	In 2 small studies reporting sequelae in children with positive EmA serology but normal biopsy results, 30% to 50% of patients were diagnosed with CD after gluten challenge. These studies were conducted prior to the availability of other serological tests, so applicability is limited. A study of 34 children with intestinal villous atrophy and simultaneous negative EmA IgA tests found that 2 infants were confirmed as having CD after 6–10 years of iterative cycles of gluten challenges and gluten-free diet. All 3 studies report high loss to followup.

CD = celiac disease; CI = confidence interval; DGP = deamidated gliadin peptide; EmA = endomysial antibodies; EPC = Evidence-based Practice Center; GI = gastrointestinal;ESPGHAN = European Society for Pediatric Gastroenterology, Hepatology, and Nutrition; HLA = human leukocyte antigen; IgA = immunoglobulin A; IgG = immunoglobulin G;LR- = negative likelihood ratio; LR+ = positive likelihood ratio; NPV = negative predictive value; OR = odds ratio; PPV = positive predictive value; SR = systematic review; tTG= anti-tissue transglutaminase; VCE= video capsule endoscopy.

Findings in Relationship to What Is Already Known

Table 22 displays findings from prior SRs along with the findings from the newly identified studies that met our inclusion criteria. We identified enough studies of the accuracy of tTG IgA tests and EmA IgA tests to conduct new meta-analyses. Our findings confirm the excellent specificity of both tests, the excellent sensitivity of tTG IgA and the good specificity of EmA IgA reported in prior SRs. A prior SR reported promising accuracy results for DGP tests; we found only one new study.

Several studies on whether adding an EmA or DGP test to a tTG test increases accuracy have recently been published. Results are insufficient to determine whether such increases are clinically meaningful.

No SRs have been conducted on the association between setting (academic vs community) and provider performance in CD diagnosis. We identified three retrospective studies evaluating inter-observer variability in histological diagnosis of CD between different pathologists and clinical settings. Results indicate that CD-related histological findings are underdiagnosed in community-based hospital and practice settings when compared to academic settings.

No SRs on how method of diagnosis affects patient adherence or clinical decisionmaking have been published. Very few studies have addressed these issues; we found insufficient evidence to answer Key Questions on this topic.

Applicability

Several factors affect the applicability of this review.

To increase generalizability, this report limited accuracy studies to those that included consecutive patients or a random sample. Several studies were excluded because we could not determine enrollment based on the information available.

Only one general population screening study met the criteria that all subjects, regardless of serology results, undergo biopsy. The cost of performing biopsies in all subjects and the low rate of acceptance of biopsy in seronegative, asymptomatic individuals makes the conduct of such studies challenging. Thus, the evidence on accuracy of diagnostic screening in the general asymptomatic population with no risk factors for CD is categorized as low strength.

Although this report is limited to diagnostic methods currently used in the U.S., study location was not a basis for exclusion. Many studies were conducted in Europe, the Middle East, and South Asia. Due to differences in genetics and disease prevalence, the applicability of these studies to the U.S. population is uncertain.

No studies stratified accuracy results by racial or ethnic group. Few studies focused on populations of special interest.

Most studies were conducted by gastroenterologists in academic settings. This report found a significant difference in interpretation of biopsy results between academic and nonacademic physicians. The majority of accuracy studies included in this report used Marsh classification to categorize biopsy results (Marsh III or higher is classified as celiac disease.) In contrast, many community physicians base their diagnosis on a simple qualitative assessment of villous atrophy or elevation of intraepithelial lymphocytes.

Accuracy of serology assays may vary by both laboratory and manufacturer. For example, Li and colleagues (2009)¹⁰⁰used 150 samples from participants of known CD status to compare accuracy of tTG tests at 20 laboratories in the US and Europe. Sensitivity was less than 75% at

four laboratories. Using a similar research design, Rozenberg and colleagues (2011)¹⁰¹found differences in performance of tTG across various manufacturers.

Finally, VCE is not a first line diagnostic method—it is indicated for adults who refuse biopsy. A 2012 SR of six studies reported very good sensitivity and excellent specificity. However, patient characteristics may differ between those who refuse a biopsy and those who accept. For example, those with more severe symptoms are hypothesized to be more likely to accept a biopsy.

Implications for Clinical and Policy Decisionmaking

The findings of this review support those of previous systematic reviews on the accuracy of individual diagnostic tests using immunoglobulin A (IgA). All IgA tests for celiac disease have excellent specificity; DGP IgA has slightly lower specificity than tTG IgA and EmA IgA. tTG IgA testing has a high positive predictive value for most clinical populations with a modest prevalence of CD. EmA IgA has good sensitivity, DGP IgA has very good sensitivity, and tTG IgA has excellent sensitivity. DGP IgG tests have very good sensitivity and excellent specificity, even in non-IgA deficient individuals.

Unfortunately, we were unable to determine which tests, if any, are more accurate in patients with specific symptoms or risk factors due to a dearth of studies meeting our inclusion criteria. Patients with symptoms associated with celiac disease would impact the pretest probability and as a result the likelihood of disease based on a positive result. No studies of test accuracy in patients with trisomy 21, Turner syndrome, and Williams syndrome were identified; the few studies of patients with Type 1 diabetes included small samples and were conducted in non-Western countries. Thus, no clinical implications for testing individuals with specific risks can be stated at this time. New research has found DGP tests more accurate than tTG in small children; strength of evidence is low but could increase if findings are replicated. tTG IgA had greater sensitivity than EmA IgA in the one study of the general (asymptomatic) population identified that met our inclusion criteria that all participants undergo biopsy, regardless of serology results. The quality of this general population study was high, the sample size was large (over 1,000) and it was conducted in a Western country (Sweden) with estimated celiac disease prevalence similar to the US.

This review found insufficient evidence to determine which populations would most benefit from diagnostic algorithms that combine a tTG test with an EmA or DGP test. A combination of positive serological testing with a threshold level at or several fold above the upper limit of normal for specific celiac tests may be accurate for diagnosing celiac disease without requiring histopathology specimens; however, the currently available evidence on comparative accuracy of algorithms is inconclusive, due to the wide range of results, heterogeneity of populations studied, and the lack of clinically significant increases in accuracy compared to individual tests. Future studies aimed at the diagnostic accuracy of multiple-test strategies would strengthen the evidence for this approach.

Finally, regarding biopsy, there is high strength evidence that multiple duodenal specimens should be taken from the duodenal bulb and the distal duodenum for optimal diagnostic yield in both the adult and pediatric population. There is moderate strength evidence that celiac disease is underdiagnosed by pathologists in community settings compared to academic settings; continued education on diagnostic protocols may be warranted for community physicians.

Limitations of the Comparative Effectiveness Review Process

At the request of AHRQ we conducted an assessment of the evidence on comparative effectiveness of various diagnostic methods currently used in the U.S. to diagnosis celiac disease. We conducted an extensive literature search; however, our consideration of unpublished literature was limited. Although a Scientific Resource Center (SRC) funded by AHRQ requested information from test manufacturers and major laboratories, no information was provided; we did not search FDA databases for such information ourselves.

In addition, this project was funded as a "small" systematic review and budgeted to include abstraction and analysis of fewer than 50 studies. Thus, the project protocol was to assess evidence from recent applicable systematic reviews and to abstract studies published thereafter. Data were not abstracted from individual studies included in prior SRs; we assumed the data presented in the SRs were abstracted accurately.

Limitations of the Evidence Base

The literature that addresses the diagnosis of celiac disease has numerous limitations that make it difficult to draw firm conclusions. These limitations can be divided into three categories: study volume, design, and reporting quality.

Volume

We identified many studies on the accuracy of tTG and EmA screens in symptomatic adults and children, including several recent systematic reviews. There were fewer studies of DGP antibody tests, as this diagnostic method is relatively new. There were also few studies assessing the accuracy of using algorithms such as those suggested by the most recent NICE and ESPGHAN guidelines.

No studies stratified accuracy results by race, ethnicity, or SES. Several studies in non-Caucasian populations were identified; however, these were not U.S. studies, and results may not be generalizable to populations in the U.S. We identified no studies of diagnostic accuracy in persons with Turner's syndrome of Trisomy 21. Literature was sparse on other populations of interest; several studies of accuracy in patients with Type 1 diabetes, iron deficiency anemia, or IgA deficiency were identified.

Almost no studies examined the impact of diagnostic method on decisionmaking or clinical or patient centered outcomes. Although the impact of living with undiagnosed celiac disease is well documented,^{102, 103} very few studies report outcomes of individuals who initially receive false positive or false negative results.

Design

Diagnostic accuracy is generally assessed through case-control and cohort studies; we included both designs. In studies employing a case-control design, a group of patients with known disease and a different group known not to have the disease undergo both the "index" test and the reference standard. Researchers are blinded to initial disease status. In a cohort design, a group of patients *suspected* of having the disease (but without a confirmed diagnosis) undergo both diagnostic methods. In a cohort design, the group is defined based on symptoms, while in a

case-control design, the group is based on disease status. The latter design is more subject to bias.

We used the QUADAS-2 instrument to assess the quality of studies of diagnostic accuracy. The ratings for each QUADAS item for each study are presented in the Evidence Tables (Appendix C); case control studies are identified. Strengths and weaknesses of individual studies are discussed in the results section of this report and taken into consideration in rating the strength of the evidence.

To lessen bias, the decision to perform the reference standard should ideally be independent of the results of the test being studied. Thus, we included only studies where all patients underwent both tests. Many studies were identified where patients first underwent serological testing and only those who tested positive underwent biopsy; although these studies provide data on false positives, they were excluded. In addition, to increase generalizability, we included only studies that enrolled a random or consecutive sample.

The use of biopsy results as the reference standard also presents concerns. As discussed in the results for Key Question 2, inter-rater reliability of interpretation is higher at academic centers than community settings. Most of the published accuracy studies included in this review took place in an academic setting.

Regarding comparative accuracy, conclusions are based primarily on indirect evidence; i.e. pooled results on accuracy of individual tests rather than head to head studies comparing accuracy of different tests in the same samples. However, strength of evidence is high, given the large numbers of studies, the consistency of results, and the precision of the confidence intervals.

Finally, most of the prior SRs described in this report were of moderate quality. Strength of evidence (SOE) was not rated by the authors; we took the strengths and weaknesses of these SRs (as we assessed using AMSTAR) into consideration when we graded the SOE of the body of evidence. An additional item we considered regarding prior SRs was the method of pooling sensitivity and specificity; pooling both jointly in a bivariate model is recommended.

Reporting Quality

Failure to report important study design details in publications is a further limitation. Some accuracy studies were vague regarding blinding of assessors and the time lapse between implementation of the index test and reference standard. Data on these items were abstracted as part of QUADAS-2 and are displayed in the Evidence Tables. Such weaknesses are discussed in the Results section and were taken into consideration in rating the strength of evidence.

Research Gaps

Although there is high strength of evidence of the accuracy of various serologic tests for celiac disease in symptomatic individuals, strength of evidence on the accuracy of algorithms such as recommended by organizations such as ESPGHAN is insufficient due to the small number of studies and inconsistent results. Appendix F contains details on the test combinations, populations, and the strength of evidence domains for each algorithm studied. Further studies should be conducted.

There is also insufficient evidence to recommend specific tests for particular at risk populations. Patient-level factors that have been hypothesized to test accuracy include race and ethnicity, but no studies stratified results by these characteristics.

Due to the inherent invasive nature of biopsy, the vast majority of studies of serologic test accuracy using biopsy as the reference standard have been conducted in patients presenting for

testing due to symptoms. The most common symptoms are gastrointestinal (diarrhea, constipation, pain, etc.) as well as malnutrition in children. High accuracy was found in the only general population screening study; however, despite the high scientific quality of this study, the strength evidence of accuracy in the asymptomatic general population is low because the study has never been replicated. This does not mean the tests are inaccurate in asymptomatic individuals; lack of evidence does not equal evidence of inaccuracy.

No studies addressing the key subquestion "What impact does the method of initial diagnosis have on how a physician follows up with a patient?" were identified. Retrospective analyses of existing databases may shed light in this area.

Finally, studies may be needed to investigate the long term impact of misdiagnosis. False positives and false negatives may be important "harms" due to a) huge lifestyle changes involved for positive diagnosis and b) potential health harm (malabsorption, intestinal damage) from undiagnosed CD.

Conclusions

New evidence on accuracy of tests used to diagnosis celiac disease supports the high sensitivity of IgA tTG tests and high specificity of both IgA tTG and IgA EmA tests reported in prior SRs. Regarding comparative accuracy, IgA EmA tests have lower sensitivity but equal specificity to IgA tTG tests. IgA DGP and IgG DGP tests are not as sensitive as IgA tTG tests in non IgA deficient adults. These conclusions are based primarily on indirect evidence; however, strength of evidence is high, given the large number of studies, the consistency of results, and the precision of the confidence intervals.

High strength of evidence of accuracy, particularly in children, was found for DGP tests in recent SRs. Algorithms combining tTG with either EmA or DGP tests appear to be accurate in both children and adults. Adding an EmA test to a tTG test resulted in increased specificity, with either no change or a slight decrease in sensitivity. In contrast, adding a DGP test to a tTG test resulted in increased sensitivity but decreased specificity. However, strength of evidence is insufficient given the low number of studies relative to single tests, heterogeneity of populations, and wide range of results. The increase in accuracy over individual tests is not consistently clinically significant. Additional studies of algorithms are needed.

Notably, current ESPGHAN guidelines state that if a patient demonstrates a tTG result greater than (10x) the normal limit, the patient should then undergo an EmA test and HLA typing; if the patient tests positive, then responds to gluten exclusion diet, a diagnosis of celiac disease can be made without use of biopsy. These guidelines have not been adopted by societies in the U.S. Evidence seems to support that a multiple-testing strategy without biopsy is accurate; however, additional studies are needed to confirm the test threshold levels that would optimize accuracy for general and specific populations.

VCE is a safe and fairly accurate means of diagnosing celiac disease in adults who wish to avoid biopsy; risk of retaining the capsule is approximately 4.6%. However, our pooled results reveal that serological tests have higher sensitivity and specificity. No data are available on how VCE accuracy varies by population characteristics or setting. Endoscopy with biopsy has a very low risk of adverse events; accuracy appears to be greater in academic settings.

Importantly, few applicable studies on the sequelae of false positive or false negative diagnoses were identified. Long-term follow-up of patients, regardless of diagnostic outcomes, should be encouraged.

References

- 1. Rashtak S, Murray JA. Review article: coeliac disease, new approaches to therapy. Aliment Pharmacol Ther. 2012 Apr;35(7):768-81. PMID: 22324389.
- Rubio-Tapia A, Ludvigsson JF, Brantner TL, et al. The prevalence of celiac disease in the United States. Am J Gastroenterol. 2012 Oct;107(10):1538-44; quiz 7, 45. PMID: 22850429.
- Ludvigsson JF, Rubio-Tapia A, van Dyke CT, et al. Increasing incidence of celiac disease in a North American population. Am J Gastroenterol. 2013 May;108(5):818-24. PMID: 23511460.
- 4. See J, Murray JA. Gluten-free diet: the medical and nutrition management of celiac disease. Nutr Clin Pract. 2006 Feb;21(1):1-15. PMID: 16439765.
- Rubio-Tapia A, Hill ID, Kelly CP, et al. ACG clinical guidelines: diagnosis and management of celiac disease. Am J Gastroenterol. 2013 May;108(5): 656-76; quiz 77. PMID: 23609613.
- Bottaro G, Rotolo N, Spina M, et al. [Evaluation of sensitivity and specificity of antigliadin antibodies for the diagnosis of celiac disease in childhood]. Minerva Pediatr. 1995 Dec;47(12):505-10. PMID: 8900559.
- Bai J, Zeballos E, Fried M, et al. WGO-OMGE practice guideline: celiac disease. World Gastroenterology Organisation (WGO-OMGE). 2007.
- National Institute for Health and Clinical Excellence (NICE). Coeliac disease: Recognition and assessment of coeliac disease. National Institute for Health and Clinical Excellence (NICE). London (UK): 2009 May.
- 9. Sollid LM. Coeliac disease: dissecting a complex inflammatory disorder. Nat Rev Immunol. 2002 Sep;2(9):647-55. PMID: 12209133.
- Rostom A, Murray JA, Kagnoff MF. American Gastroenterological Association (AGA) Institute technical review on the diagnosis and management of celiac disease. Gastroenterology. 2006 Dec;131(6):1981-2002. PMID: 17087937.
- Husby S, Koletzko S, Korponay-Szabo IR, et al. European Society for Pediatric Gastroenterology, Hepatology, and Nutrition guidelines for the diagnosis of coeliac disease. J Pediatr Gastroenterol Nutr. 2012 Jan;54(1):136-60. PMID: 22197856.
- 12. Corazza GR, Villanacci V. Coeliac disease. J Clin Pathol. 2005 Jun;58(6):573-4. PMID: 15917404.

- Marsh MN. Gluten, major histocompatibility complex, and the small intestine. A molecular and immunobiologic approach to the spectrum of gluten sensitivity ('celiac sprue'). Gastroenterology. 1992 Jan;102(1):330-54. PMID: 1727768.
- Oberhuber G, Granditsch G, Vogelsang H. The histopathology of coeliac disease: time for a standardized report scheme for pathologists. Eur J Gastroenterol Hepatol. 1999 Oct;11(10):1185-94. PMID: 10524652.
- Dickson BC, Streutker CJ, Chetty R. Coeliac disease: an update for pathologists. J Clin Pathol. 2006 Oct;59(10):1008-16. PMID: 17021129.
- Bai J, Zeballos E, Fried M, et al. Coeliac Disease. WGO-OMGE Practice Guideline World Gastroenterol News. 2005;10(2 suppl):S1-S8.
- Hill ID, Dirks MH, Liptak GS, et al. Guideline for the diagnosis and treatment of celiac disease in children: recommendations of the North American Society for Pediatric Gastroenterology, Hepatology and Nutrition. J Pediatr Gastroenterol Nutr. 2005 Jan;40(1):1-19. PMID: 15625418.
- Bai JC, Fried M, Corazza GR, et al. World Gastroenterology Organisation global guidelines on celiac disease. J Clin Gastroenterol. 2013 Feb;47(2):121-6. PMID: 23314668.
- Revised criteria for diagnosis of coeliac disease. Report of Working Group of European Society of Paediatric Gastroenterology and Nutrition. Arch Dis Child. 1990 Aug;65(8):909-11. PMID: 2205160.
- Rostom A, Dube C, Cranney A, et al. Celiac disease. Evid Rep Technol Assess (Summ). 2004 Jun(104): 1-6. PMID: 15346868.
- Whiting PF, Rutjes AW, Westwood ME, et al. QUADAS-2: a revised tool for the quality assessment of diagnostic accuracy studies. Ann Intern Med. 2011 Oct 18;155(8):529-36. PMID: 22007046.
- 22. Shea BJ, Grimshaw JM, Wells GA, et al. Development of AMSTAR: a measurement tool to assess the methodological quality of systematic reviews. BMC Med Res Methodol. 2007;7:10. PMID: 17302989.
- 23. Santaguida PL, P. R. The Development of the McHarm Quality Assessment Scale for adverse events: Delphi Consensus on important criteria for evaluating harms. Available at: http://hiru.mcmaster.ca/epc/mcharm.pdf. 2008.

- 24. Parikh R, Mathai A, Parikh S, et al. Understanding and using sensitivity, specificity and predictive values. Indian journal of ophthalmology. 2008;56(1):45.
- 25. McGee S. Simplifying likelihood ratios. J Gen Intern Med. 2002 Aug;17(8):646-9. PMID: 12213147.
- Deeks JJ, Altman DG. Diagnostic tests 4: likelihood ratios. BMJ. 2004 Jul 17;329(7458):168-9. PMID: 15258077.
- 27. Rutter CM, Gatsonis CA. A hierarchical regression approach to meta-analysis of diagnostic test accuracy evaluations. Stat Med. 2001 Oct 15;20(19):2865-84. PMID: 11568945.
- 28. Reitsma JB, Glas AS, Rutjes AW, et al. Bivariate analysis of sensitivity and specificity produces informative summary measures in diagnostic reviews. J Clin Epidemiol. 2005 Oct;58(10):982-90. PMID: 16168343.
- 29. Rector TS, Taylor BC, Wilt TJ. Systematic Review of Prognostic Tests. In: Chang SM, Matchar DB, Smetana GW, Umscheid CA, eds. Methods Guide for Medical Test Reviews. Rockville (MD); 2012.
- 30. Chang SM, Matchar DB, Smetana GW, et al. Methods Guide for Medical Test Reviews. AHRQ Publication No. 12-EC017 Agency for Healthcare Research and Quality. Rockville (MD): 2012. http://www.ncbi.nlm.nih.gov/pubmed/22834019.
- Barada K, Habib RH, Malli A, et al. Prediction of celiac disease at endoscopy. Endoscopy. 2014 Feb;46(2):110-9. PMID: 24477366.
- 32. Basso D, Guariso G, Bozzato D, et al. New screening tests enrich anti-transglutaminase results and support a highly sensitive two-test based strategy for celiac disease diagnosis. Clin Chim Acta. 2011 Aug 17;412(17-18):1662-7. PMID: 21640087.
- Bienvenu F, Anghel SI, Besson Duvanel C, et al. Early diagnosis of celiac disease in IgA deficient children: contribution of a point-of-care test. BMC Gastroenterol. 2014;14:186. PMID: 25376178.
- Cekin AH, Cekin Y, Sezer C. Celiac disease prevalence in patients with iron deficiency anemia. Turk J Gastroenterol. 2012;23(5):490-5. PMID: 23161292.
- 35. Dahlbom I, Korponay-Szabo IR, Kovacs JB, et al. Prediction of clinical and mucosal severity of coeliac disease and dermatitis herpetiformis by quantification of IgA/IgG serum antibodies to tissue transglutaminase. J Pediatr Gastroenterol Nutr. 2010 Feb;50(2):140-6. PMID: 19841593.

- 36. Dahle C, Hagman A, Ignatova S, et al. Antibodies against deamidated gliadin peptides identify adult coeliac disease patients negative for antibodies against endomysium and tissue transglutaminase. Aliment Pharmacol Ther. 2010 Jul;32(2):254-60. PMID: 20456302.
- DeGaetani M, Tennyson CA, Lebwohl B, et al. Villous atrophy and negative celiac serology: a diagnostic and therapeutic dilemma. Am J Gastroenterol. 2013 May;108(5):647-53. PMID: 23644957.
- Dutta AK, Chacko A, Avinash B. Suboptimal performance of IgG anti-tissue transglutaminase in the diagnosis of celiac disease in a tropical country. Dig Dis Sci. 2010 Mar;55(3):698-702. PMID: 19333755.
- 39. Emami MH, Karimi S, Kouhestani S. Is routine duodenal biopsy necessary for the detection of celiac disease in patients presenting with iron deficiency anemia? Int J Prev Med. 2012 Apr;3(4):273-7. PMID: 22624084.
- Harrison E, Li KK, Petchey M, et al. Selective measurement of anti-tTG antibodies in coeliac disease and IgA deficiency: an alternative pathway. Postgrad Med J. 2013 Jan;89(1047):4-7. PMID: 22872871.
- Kaukinen K, Collin P, Mykkanen AH, et al. Celiac disease and autoimmune endocrinologic disorders. Dig Dis Sci. 1999 Jul;44(7):1428-33. PMID: 10489930.
- 42. Mansour AA, Najeeb AA. Coeliac disease in Iraqi type 1 diabetic patients. Arab J Gastroenterol. 2011 Jun;12(2):103-5. PMID: 21684484.
- 43. Mozo L, Gomez J, Escanlar E, et al. Diagnostic value of anti-deamidated gliadin peptide IgG antibodies for celiac disease in children and IgA-deficient patients. J Pediatr Gastroenterol Nutr. 2012 Jul;55(1):50-5. PMID: 22197936.
- Nevoral J, Kotalova R, Hradsky O, et al. Symptom positivity is essential for omitting biopsy in children with suspected celiac disease according to the new ESPGHAN guidelines. Eur J Pediatr. 2013 Nov 15PMID: 24233405.
- 45. Olen O, Gudjonsdottir AH, Browaldh L, et al. Antibodies against deamidated gliadin peptides and tissue transglutaminase for diagnosis of pediatric celiac disease. J Pediatr Gastroenterol Nutr. 2012 Dec;55(6):695-700. PMID: 22722680.
- Sakly W, Mankai A, Ghdess A, et al. Performance of anti-deamidated gliadin peptides antibodies in celiac disease diagnosis. Clin Res Hepatol Gastroenterol. 2012 Dec;36(6):598-603. PMID: 22436429.

- 47. Srinivas M, Basumani P, Podmore G, et al. Utility of testing patients, on presentation, for serologic features of celiac disease. Clin Gastroenterol Hepatol. 2014 Jun;12(6):946-52. PMID: 24262940.
- Srinivas M, Basumani P, Podmore G, et al. Utility of Testing Patients, on Presentation, for Serologic Features of Celiac Disease. Clin Gastroenterol Hepatol. 2013 Nov 19PMID: 24262940.
- 49. Sugai E, Hwang HJ, Vazquez H, et al. New serology assays can detect gluten sensitivity among enteropathy patients seronegative for anti-tissue transglutaminase. Clin Chem. 2010 Apr;56(4):661-5. PMID: 20022983.
- 50. Swallow K, Wild G, Sargur R, et al. Quality not quantity for transglutaminase antibody 2: the performance of an endomysial and tissue transglutaminase test in screening coeliac disease remains stable over time. Clin Exp Immunol. 2013 Jan;171(1):100-6. PMID: 23199329.
- 51. Van Meensel B, Hiele M, Hoffman I, et al. Diagnostic accuracy of ten second-generation (human) tissue transglutaminase antibody assays in celiac disease. Clin Chem. 2004 Nov;50(11): 2125-35. PMID: 15388634.
- 52. Vermeersch P, Geboes K, Marien G, et al. Diagnostic performance of IgG anti-deamidated gliadin peptide antibody assays is comparable to IgA anti-tTG in celiac disease. Clin Chim Acta. 2010 Jul 4;411 (13-14):931-5. PMID: 20171961.
- 53. Vermeersch P, Coenen D, Geboes K, et al. Use of likelihood ratios improves clinical interpretation of IgA anti-tTG antibody testing for celiac disease. Clin Chim Acta. 2010 Jan;411(1-2):13-7. PMID: 19799890.
- 54. Vermeersch P, Geboes K, Marien G, et al. Serological diagnosis of celiac disease: comparative analysis of different strategies. Clin Chim Acta. 2012 Nov 12;413(21-22):1761-7. PMID: 22771970.
- 55. Wakim-Fleming J, Pagadala MR, McCullough AJ, et al. Prevalence of celiac disease in cirrhosis and outcome of cirrhosis on a gluten free diet: a prospective study. J Hepatol. 2014 Sep;61(3):558-63. PMID: 24842303.
- 56. Walker MM, Murray JA, Ronkainen J, et al. Detection of celiac disease and lymphocytic enteropathy by parallel serology and histopathology in a population-based study. Gastroenterology. 2010 Jul;139(1):112-9. PMID: 20398668.
- 57. Wolf J, Hasenclever D, Petroff D, et al. Antibodies in the diagnosis of coeliac disease: a biopsy-controlled, international, multicentre study of 376 children with coeliac disease and 695 controls. PLoS One. 2014;9(5):e97853. PMID: 24830313.

- 58. Zanini B, Magni A, Caselani F, et al. High tissuetransglutaminase antibody level predicts small intestinal villous atrophy in adult patients at high risk of celiac disease. Dig Liver Dis. 2012 Apr;44(4): 280-5. PMID: 22119616.
- 59. Lewis NR, Scott BB. Meta-analysis: deamidated gliadin peptide antibody and tissue transglutaminase antibody compared as screening tests for coeliac disease. Aliment Pharmacol Ther. 2010 Jan;31(1): 73-81. PMID: 19664074.
- Giersiepen K, Lelgemann M, Stuhldreher N, et al. Accuracy of diagnostic antibody tests for coeliac disease in children: summary of an evidence report. J Pediatr Gastroenterol Nutr. 2012 Feb;54(2):229-41. PMID: 22266486.
- Schyum AC, Rumessen JJ. Serological testing for celiac disease in adults. United European Gastroenterol J. 2013 Oct;1(5):319-25. PMID: 24917978.
- 62. Basso D, Gallo N, Guariso G, et al. Role of antitransglutaminase (anti-tTG), anti-gliadin, and antiendomysium serum antibodies in diagnosing celiac disease: a comparison of four different commercial kits for anti-tTG determination. J Clin Lab Anal. 2001;15(3):112-5. PMID: 11344524.
- 63. Rokkas T, Niv Y. The role of video capsule endoscopy in the diagnosis of celiac disease: a metaanalysis. Eur J Gastroenterol Hepatol. 2012 Mar;24(3):303-8. PMID: 22266837.
- 64. El-Matary W, Huynh H, Vandermeer B. Diagnostic characteristics of given video capsule endoscopy in diagnosis of celiac disease: a meta-analysis. J Laparoendosc Adv Surg Tech A. 2009 Dec;19(6):815-20. PMID: 19405806.
- 65. Hall NJ, Rubin G, Charnock A. Systematic review: adherence to a gluten-free diet in adult patients with coeliac disease. Aliment Pharmacol Ther. 2009 Aug 15;30(4):315-30. PMID: 19485977.
- Shamir R, Yehezkely-Schildkraut V, Hartman C, et al. Population screening for celiac disease: follow up of patients identified by positive serology. J Gastroenterol Hepatol. 2007 Apr;22(4):532-5. PMID: 17376047.
- 67. Lebwohl B, Genta RM, Kapel RC, et al. Procedure volume influences adherence to celiac disease guidelines. Eur J Gastroenterol Hepatol. 2013 Nov;25(11):1273-8. PMID: 23995767.
- Mubarak A, Nikkels P, Houwen R, et al. Reproducibility of the histological diagnosis of celiac disease. Scand J Gastroenterol. 2011 Sep;46(9): 1065-73. PMID: 21668407.

- Arguelles-Grande C, Tennyson CA, Lewis SK, et al. Variability in small bowel histopathology reporting between different pathology practice settings: impact on the diagnosis of coeliac disease. J Clin Pathol. 2012 Mar;65(3):242-7. PMID: 22081783.
- Pinto Sanchez MI, Smecuol E, Vazquez H, et al. Very high rate of misdiagnosis of celiac disease in clinical practice. Acta Gastroenterol Latinoam. 2009 Dec;39(4):250-3. PMID: 20178253.
- Prasad KK, Thapa BR, Nain CK, et al. The frequency of histologic lesion variability of the duodenal mucosa in children with celiac disease. World J Pediatr. 2010 Feb;6(1):60-4. PMID: 20143213.
- Weir DC, Glickman JN, Roiff T, et al. Variability of histopathological changes in childhood celiac disease. Am J Gastroenterol. 2010 Jan;105(1): 207-12. PMID: 19809405.
- Bonamico M, Mariani P, Thanasi E, et al. Patchy villous atrophy of the duodenum in childhood celiac disease. J Pediatr Gastroenterol Nutr. 2004 Feb;38(2):204-7. PMID: 14734885.
- 74. Bonamico M, Thanasi E, Mariani P, et al. Duodenal bulb biopsies in celiac disease: a multicenter study. J Pediatr Gastroenterol Nutr. 2008 Nov;47(5):618-22. PMID: 18979585.
- Lebwohl B, Kapel RC, Neugut AI, et al. Adherence to biopsy guidelines increases celiac disease diagnosis. Gastrointest Endosc. 2011 Jul;74(1): 103-9. PMID: 21601201.
- 76. Pais WP, Duerksen DR, Pettigrew NM, et al. How many duodenal biopsy specimens are required to make a diagnosis of celiac disease? Gastrointest Endosc. 2008 Jun;67(7):1082-7. PMID: 18308317.
- Hopper AD, Cross SS, Sanders DS. Patchy villous atrophy in adult patients with suspected glutensensitive enteropathy: is a multiple duodenal biopsy strategy appropriate? Endoscopy. 2008 Mar;40(3):219-24. PMID: 18058655.
- Nenna R, Pontone S, Pontone P, et al. Duodenal bulb in celiac adults: the "whether biopsying" dilemma. J Clin Gastroenterol. 2012 Apr;46(4):302-7. PMID: 21934529.
- 79. Caruso R, Marafini I, Del Vecchio Blanco G, et al. Sampling of proximal and distal duodenal biopsies in the diagnosis and monitoring of celiac disease. Dig Liver Dis. 2014 Apr;46(4):323-9. PMID: 24394601.
- Evans KE, Aziz I, Cross SS, et al. A prospective study of duodenal bulb biopsy in newly diagnosed and established adult celiac disease. Am J Gastroenterol. 2011 Oct;106(10):1837-742. PMID: 21606978.

- Ravelli A, Bolognini S, Gambarotti M, et al. Variability of histologic lesions in relation to biopsy site in gluten-sensitive enteropathy. Am J Gastroenterol. 2005 Jan;100(1):177-85. PMID: 15654798.
- 82. Sharma A, Mews C, Jevon G, et al. Duodenal bulb biopsy in children for the diagnosis of coeliac disease: experience from Perth, Australia. J Paediatr Child Health. 2013 Mar;49(3):210-4.
 PMID: 23432775.
- Levinson-Castiel R, Hartman C, Morgenstern S, et al. The role of duodenal bulb biopsy in the diagnosis of celiac disease in children. J Clin Gastroenterol. 2011 Jan;45(1):26-9. PMID: 20628309.
- Bruins MJ. The clinical response to gluten challenge: a review of the literature. Nutrients. 2013 Nov;5(11):4614-41. PMID: 24284613.
- 85. Leffler D, Schuppan D, Pallav K, et al. Kinetics of the histological, serological and symptomatic responses to gluten challenge in adults with coeliac disease. Gut. 2013 Jul;62(7):996-1004.
 PMID: 22619366.
- 86. Ford AC, Chey WD, Talley NJ, et al. Yield of Diagnostic Tests for Celiac Disease in Individuals With Symptoms Suggestive of Irritable Bowel Syndrome Systematic Review and Meta-analysis. Archives of Internal Medicine. 2009 Apr;169(7): 651-8. PMID: WOS:000265105700002.
- 87. van der Windt DA, Jellema P, Mulder CJ, et al. Diagnostic testing for celiac disease among patients with abdominal symptoms: a systematic review. JAMA. 2010 May 5;303(17):1738-46. PMID: 20442390.
- Altobelli E, Paduano R, Gentile T, et al. Healthrelated quality of life in children and adolescents with celiac disease: survey of a population from central Italy. Health Qual Life Outcomes. 2013;11:204. PMID: 24304679.
- Cave D, Legnani P, de Franchis R, et al. ICCE consensus for capsule retention. Endoscopy. 2005 Oct;37(10):1065-7. PMID: 16189792.
- 90. Vere CC, Rogoveanu I, Streba CT, et al. The role of capsule endoscopy in the detection of small bowel disease. Chirurgia (Bucur). 2012 May-Jun;107(3):352-60. PMID: 22844834.
- 91. Gomez V, Cheesman AR, Heckman MG, et al. Safety of capsule endoscopy in the octogenarian as compared with younger patients. Gastrointestinal Endoscopy. 2013 Nov;78(5):744-9. PMID: WOS:000325485100013.

- 92. Atay O, Mahajan L, Kay M, et al. Risk of capsule endoscope retention in pediatric patients: a large single-center experience and review of the literature. J Pediatr Gastroenterol Nutr. 2009 Aug;49(2): 196-201. PMID: 19561547.
- 93. Liao Z, Gao R, Xu C, et al. Indications and detection, completion, and retention rates of small-bowel capsule endoscopy: a systematic review. Gastrointest Endosc. 2010 Feb;71(2):280-6. PMID: 20152309.
- 94. Persliden J, Pettersson HB, Falth-Magnusson K. Small intestinal biopsy in children with coeliac disease: measurement of radiation dose and analysis of risk. Acta Paediatr. 1993 Mar;82(3):296-9. PMID: 8495087.
- 95. Pettersson HB, Falth-Magnusson K, Persliden J, et al. Radiation risk and cost-benefit analysis of a paediatric radiology procedure: results from a national study. Br J Radiol. 2005 Jan;78(925): 34-8. PMID: 15673527.
- 96. Romagnuolo J, Cotton PB, Eisen G, et al. Identifying and reporting risk factors for adverse events in endoscopy. Part II: noncardiopulmonary events. Gastrointest Endosc. 2011 Mar;73(3):586-97. PMID: 21353858.
- 97. Kurppa K, Rasanen T, Collin P, et al. Endomysial antibodies predict celiac disease irrespective of the titers or clinical presentation. World J Gastroenterol. 2012 May 28;18(20):2511-6. PMID: 22654448.
- 98. Kurppa K, Paavola A, Collin P, et al. Benefits of a gluten-free diet for asymptomatic patients with serologic markers of celiac disease.
 Gastroenterology. 2014 Sep;147(3):610-7 e1. PMID: 24837306.
- 99. Smecuol E, Maurino E, Vazquez H, et al. Gynaecological and obstetric disorders in coeliac disease: frequent clinical onset during pregnancy or the puerperium. Eur J Gastroenterol Hepatol. 1996 Jan;8(1):63-89. PMID: 8900911.
- 100. Li M, Yu LP, Tiberti C, et al. A Report on the International Transglutaminase Autoantibody Workshop for Celiac Disease. American Journal of Gastroenterology. 2009 Jan;104(1):154-63. PMID: WOS:000262265800026.
- 101. Rozenberg O, Lerner A, Pacht A, et al. A novel algorithm for the diagnosis of celiac disease and a comprehensive review of celiac disease diagnostics. Clin Rev Allergy Immunol. 2012 Jun;42(3):331-41. PMID: 21279475.
- 102. Green PH, Cellier C. Celiac disease. N Engl J Med. 2007 Oct 25;357(17):1731-43. PMID: 17960014.

 Goddard CJR, Gillett HR. Complications of coeliac disease: are all patients at risk? Postgraduate Medical Journal. 2006 Nov;82(973):705-12. PMID: WOS:000241945900003.

Abbreviations/Acronyms

ACG - American College of Gastroenterology **AEs - Adverse Events** AGA - Anti-Gliadin Antibodies AHRQ - Agency for Healthcare Research and Quality CD - Celiac Disease CI - Confidence Interval COI - Conflict of Interest DGP - Deamidated Gliadin Peptide (DGP) Antibodies ELISA - Enzyme-Linked Immunosorbent Assay EmA - Endomysial antibodies EPC - Southern California Evidence-based Practice Center ESPGHAN - European Society for Pediatric Gastroenterology, Hepatology, and Nutrition FN - False Negative FP - False Positive GI - Gastrointestinal HLA - Human Leukocyte Antigen IBS - Irritable Bowel Syndrome IgA - Immunoglobulin A IgG - Immunoglobulin G IU - International Unit KI - Key Informants **KQ** - Key Question LR - Likelihood Ratio NA - Not Applicable NASPGHAN - North American Society for Pediatric Gastroenterology, Hepatology, and Nutrition NICE - National Institute for Clinical Excellence NPV - Negative Predictive Value NR - Not Reported PICOTS - Populations, Interventions, Comparators, Outcomes, and Timing PPV - Positive Predictive Value QOL - Quality of life RBA - Radio blinding assay **ROC** - Receiver Operating Characteristic SES - Socioeconomic status SR - Systematic Review tTG - Tissue Transglutaminase VCE - Video Capsule Endoscopy

WGO - World Gastroenterology Organization

Appendix A. Search Strategy

KQ1 (DIAGNOSTIC METHODS):

SEARCH #1 (DIAGNOSTIC ACCURACY) DATABASE SEARCHED & TIME PERIOD COVERED:

PubMed - 1/1/2010-1/07/2015

LANGUAGE:

English

SEARCH STRATEGY:

"celiac disease "[Mesh] OR "celiac disease"[tiab] OR "coeliac disease"[tiab] AND

diagnosis OR diagnoses OR diagnostic OR diagnose OR diagnosing OR endomysial OR transglutaminase* OR serolog* OR antibody OR antibodies OR leucocyte* OR hla OR biopsy OR biopsies OR test OR tests OR testing OR screen OR screening OR screened OR

"Transglutaminases" [Mesh] OR "HLA Antigens" [Mesh] OR iga OR ttg OR dgp OR IGG OR mass screening OR diagnosis [mh] OR biopsy [mh] OR biopsies [mh] OR "deamidated gliadin peptide antibodies" OR Human leukocyte antigen* OR "video capsule endoscopy" OR endoscop* AND

Accura* OR Sensitivity and specificity[mh] OR Sensitivity[tiab] OR Specificity[tiab] OR False positive reactions[mh] OR false positive* OR False negative reactions[mh] OR False negative* OR Predictive value OR predictive value of tests[mh] OR Distinguish* OR Differential* OR Identif* OR Detect* OR valid* OR reliab* OR reproducibility of results

SEARCH #2 (INTERMEDIATE OUTCOMES): DATABASE SEARCHED & TIME PERIOD COVERED: PubMed – 1/1/2010-1/07/2015

LANGUAGE:

English

SEARCH STRATEGY:

"celiac disease "[Mesh] OR "celiac disease"[tiab] OR "coeliac disease"[tiab] AND

diagnosis OR diagnoses OR diagnostic OR diagnose OR diagnosing OR endomysial OR transglutaminase* OR serolog* OR antibody OR antibodies OR leucocyte* OR hla OR biopsy OR biopsies OR test OR tests OR testing OR screen OR screening OR screened OR

"Transglutaminases" [Mesh] OR "HLA Antigens" [Mesh] OR iga OR ttg OR dgp OR IGG OR mass screening OR diagnosis [mh] OR biopsy [mh] OR biopsies [mh] OR "deamidated gliadin peptide antibodies" OR Human leukocyte antigen* OR "video capsule endoscopy" OR endoscop* OR misdiagnos* OR undiagnos*

AND

intermediate outcome* OR decision* OR dietary OR diet OR nutrition* OR eating OR food OR foods OR compliance OR comply OR complying OR patient compliance OR adherence OR (("Decision Making"[Mesh]) OR "Decision Support Systems, Clinical"[Mesh]) OR "Food Habits"[Mesh]

SEARCH #3 (CLINICAL OUTCOMES/COMPLICATIONS): DATABASE SEARCHED & TIME PERIOD COVERED:

PubMed - 1/1/2010-1/07/2015

LANGUAGE:

English

SEARCH STRATEGY:

"celiac disease "[Mesh] OR "celiac disease"[tiab] OR "coeliac disease"[tiab] AND

diagnosis OR diagnoses OR diagnostic OR diagnose OR diagnosing OR endomysial OR transglutaminase* OR serolog* OR antibody OR antibodies OR leucocyte* OR hla OR biopsy OR biopsies OR test OR tests OR testing OR screen OR screening OR screened OR "Transglutaminases"[Mesh] OR "HLA Antigens"[Mesh] OR iga OR ttg OR dgp OR IGG OR mass screening OR diagnosis[mh] OR biopsy[mh] OR biopsies[mh] OR "deamidated gliadin peptide antibodies" OR Human leukocyte antigen* OR "video capsule endoscopy" OR endoscop* AND

clinical outcome* OR complication* OR adverse event* OR adverse effect* OR harm* OR enteropathy OR "quality of life" OR villous atrophy OR abdominal OR anemia OR anemic OR (deficien* AND (folic acid OR folate)) OR "Outcome and Process Assessment (Health Care)"[Mesh] OR "complications" [Subheading] OR "adverse effects" [Subheading] OR "Quality of Life"[Mesh] OR "Folic Acid Deficiency"[Mesh]

SEARCH #4 (ADD TERMS "MISDIAGNOS* OR "UNDIAGNOS*"): DATABASE SEARCHED & TIME PERIOD COVERED:

PubMed - 1/1/2010-1/07/2015

LANGUAGE:

English

SEARCH STRATEGY:

"celiac disease "[Mesh] OR "celiac disease"[tiab] OR "coeliac disease"[tiab] AND misdiagnos* OR undiagnos*

NOTE – THESE RESULTS WERE INCORPORATED INTO THE PREVIOUS RESULT SETS

SEARCH #1 (DIAGNOSTIC ACCURACY): DATABASE SEARCHED & TIME PERIOD COVERED:

Web of Science Indexes=SCI-EXPANDED, SSCI, A&HCI, CPCI-S, CPCI-SSH - 1/1/2010-107/2015

LANGUAGE:

English

SEARCH STRATEGY: ("ts"= topical search)

ts=("celiac disease" OR "coeliac disease")
AND

ts=(diagnosis OR diagnoses OR diagnostic OR diagnose OR diagnosing OR endomysial OR transglutaminase* OR serolog* OR antibody OR antibodies OR leucocyte* OR hla OR biopsy OR biopsies OR test OR tests OR testing OR screen OR screening OR screened OR Transglutaminase* OR iga OR ttg OR dgp OR IGG OR mass screening OR "Human leukocyte antigen*" OR endoscop* OR misdiagnos* OR undiagnos*)

AND

ts=(Accura* OR Sensitivity OR Specificity OR false positive* OR False negative* OR Predictive value OR Distinguish* OR Differential* OR Identif* OR Detect* OR valid* OR reliab* OR reproducib*)

SEARCH #2 (INTERMEDIATE OUTCOMES): DATABASE SEARCHED & TIME PERIOD COVERED:

Web of Science Indexes=SCI-EXPANDED, SSCI, A&HCI, CPCI-S, CPCI-SSH – 1/1/2010-1/07/2015

LANGUAGE:

English

SEARCH STRATEGY:

ts=("celiac disease" OR "coeliac disease")

AND

ts=(diagnosis OR diagnoses OR diagnostic OR diagnose OR diagnosing OR endomysial OR transglutaminase* OR serolog* OR antibody OR antibodies OR leucocyte* OR hla OR biopsy OR biopsies OR test OR tests OR testing OR screen OR screening OR screened OR Transglutaminase* OR iga OR ttg OR dgp OR IGG OR mass screening OR "Human leukocyte antigen*" OR endoscop* OR misdiagnos* OR undiagnos*)

AND

ts=(intermediate outcome* OR decision* OR dietary OR diet OR nutrition* OR eating OR food OR foods OR compliance OR comply OR complying OR adherence)

SEARCH #3 (CLINICAL OUTCOMES/COMPLICATIONS): DATABASE SEARCHED & TIME PERIOD COVERED:

Web of Science Indexes=SCI-EXPANDED, SSCI, A&HCI, CPCI-S, CPCI-SSH - 1/1/2010-1/07/2015

LANGUAGE:

English

SEARCH STRATEGY:

ts=("celiac disease" OR "coeliac disease")

AND

ts=(diagnosis OR diagnoses OR diagnostic OR diagnose OR diagnosing OR endomysial OR transglutaminase* OR serolog* OR antibody OR antibodies OR leucocyte* OR hla OR biopsy OR biopsies OR test OR tests OR testing OR screen OR screening OR screened OR Transglutaminase* OR iga OR ttg OR dgp OR IGG OR mass screening OR "Human leukocyte antigen*" OR endoscop* OR misdiagnos* OR undiagnos*)

AND

ts=("adverse effect*" OR "adverse event*" OR "clinical outcome*" OR complication* OR harm* OR enteropathy OR "quality of life" OR villous atrophy OR abdominal OR anemia OR anemic OR "folic acid" OR folate)

DATABASE SEARCHED & TIME PERIOD COVERED:

Cochrane Databases - 1/1/2010-1/07/2015

LANGUAGE:

English

SEARCH STRATEGY:

"celiac disease" OR "coeliac disease" in Title, Abstract, Keywords

NUMBER OF RESULTS: 65

By database: Cochrane Reviews (0) Other Reviews (5) Trials (54) Methods Studies (0) Technology Assessments (3) Economic Evaluations (3) Cochrane Groups (0)

KQ2 (ENDOSCOPY WITH DUODENAL BIOPSY)

DATABASE SEARCHED & TIME PERIOD COVERED:

PubMed - 1/1/1990-1/07/2015

LANGUAGE:

English

SEARCH STRATEGY:

"celiac disease "[Mesh] OR "celiac disease"[tiab] OR "coeliac disease"[tiab] AND endoscopy AND duodenal or duodenum AND biopsy OR biopsies

DATABASE SEARCHED & TIME PERIOD COVERED:

Web of Science Indexes=SCI-EXPANDED, SSCI, A&HCI, CPCI-S, CPCI-SSH – 1/1/1990-1/07/2015

LANGUAGE:

English

SEARCH STRATEGY:

TOPIC: ("celiac disease" OR "coeliac disease") AND TOPIC: (endoscop* AND (duodenal or duodenum) AND (biopsy OR biopsies))

KQ3 (POPULATION): DATABASE SEARCHED & TIME PERIOD COVERED: Data 1/1/1000 1/07/2015

PubMed - 1/1/1990-1/07/2015

LANGUAGE:

English

SEARCH STRATEGY:

"celiac disease "[Mesh] OR "celiac disease"[tiab] OR "coeliac disease"[tiab] AND

diagnosis OR diagnoses OR diagnostic OR diagnose OR diagnosing OR endomysial OR transglutaminase* OR serolog* OR antibody OR antibodies OR leucocyte* OR hla OR biopsy OR biopsies OR test OR tests OR testing OR screen OR screening OR screened OR "Transglutaminases"[Mesh] OR "HLA Antigens"[Mesh] OR iga OR ttg OR dgp OR IGG OR mass screening OR diagnosis[mh] OR biopsy[mh] OR biopsies[mh] OR "deamidated gliadin peptide

antibodies" OR Human leukocyte antigen* OR "video capsule endoscopy" OR endoscop* OR misdiagnos* OR undiagnos*

AND

"Ethnic Groups" [Mesh] OR "Minority Groups" [Mesh] OR "Socioeconomic Factors" [Mesh] OR "Continental Population Groups" [Mesh] OR "Demography" [Mesh] OR population* OR symptomatic OR nonsymptomatic OR non-symptomatic OR child OR children OR infant OR infants OR pediatric* OR paediatric* OR demograph* OR race OR racial OR ethnic OR ethnicit* OR minority OR minorities OR genetic* OR geograph* OR region OR regions OR regional OR socioeconom* OR socio-econom* OR economic* OR income OR (iga AND deficien*) OR negative OR country [tiab] OR countries [tiab]

ĀND

"outcome assessment health care" [MeSH Terms] OR Accura* OR Sensitivity and specificity[mh] OR Sensitivity[tiab] OR Specificity[tiab] OR False positive reactions[mh] OR false positive* OR False negative reactions[mh] OR False negative* OR Predictive value OR predictive value of tests[mh] OR Distinguish* OR Differential* OR Identif* OR Detect* OR valid* OR reliab* OR reproducibility of results OR outcome OR outcomes OR treatment outcome OR treatment outcomes NOT

case report* OR case reports[pt])

DATABASE SEARCHED & TIME PERIOD COVERED:

Web of Science Indexes=SCI-EXPANDED, SSCI, A&HCI, CPCI-S, CPCI-SSH – 1/1/1990-1/07/2015

LANGUAGE: English

SEARCH STRATEGY:

ts=("celiac disease" OR "coeliac disease") AND

ts=(diagnosis OR diagnoses OR diagnostic OR diagnose OR diagnosing OR endomysial OR transglutaminase* OR serolog* OR antibody OR antibodies OR leucocyte* OR hla OR biopsy OR biopsies OR test OR tests OR testing OR screen OR screening OR screened OR Transglutaminase* OR iga OR ttg OR dgp OR IGG OR mass screening OR "Human leukocyte antigen*" OR endoscop* OR misdiagnos* OR undiagnos*)

AND

ts=(population* OR symptomatic OR nonsymptomatic OR non-symptomatic OR child OR children OR infant OR infants OR pediatric* OR paediatric* OR demograph* OR race OR racial OR ethnic OR ethnicit* OR minority OR minorities OR genetic* OR geograph* OR region OR regions OR regional OR socioeconom* OR socio-econom* OR economic* OR income OR (iga AND deficien*) OR negative OR country OR countries)

AND

ts=(Accura* OR Sensitivity OR Specificity OR false positive* OR False negative* OR "predictive value" OR Distinguish* OR Differential* OR Identif* OR Detect* OR valid* OR reliab* OR reproducib* OR outcome* OR prevalen*)

KQ4 (ADVERSE EVENTS):

DATABASE SEARCHED & TIME PERIOD COVERED:

PubMed - 1/1/2003-1/07/2015

LANGUAGE:

English

SEARCH STRATEGY:

"celiac disease "[Mesh] OR "celiac disease"[tiab] OR "coeliac disease"[tiab] AND

diagnosis OR diagnoses OR diagnostic OR diagnose OR diagnosing OR endomysial OR transglutaminase* OR serolog* OR antibody OR antibodies OR leucocyte* OR hla OR biopsy OR biopsies OR test OR tests OR testing OR screen OR screening OR screened OR "Transglutaminases"[Mesh] OR "HLA Antigens"[Mesh] OR iga OR ttg OR dgp OR IGG OR mass screening OR diagnosis[mh] OR biopsy[mh] OR biopsies[mh] OR "deamidated gliadin peptide antibodies" OR Human leukocyte antigen* OR "video capsule endoscopy" OR endoscop* OR misdiagnos* OR undiagnos*

AND

adverse effect* OR adverse event* OR harm* OR bleeding OR perforat* OR danger* OR safe*[tiab] OR safety[tiab] OR patient safety OR accident*

DATABASE SEARCHED & TIME PERIOD COVERED:

Web of Science Indexes=SCI-EXPANDED, SSCI, A&HCI, CPCI-S, CPCI-SSH - 1/1/2003-1/07/2015

LANGUAGE:

English

SEARCH STRATEGY:

ts=("celiac disease" OR "coeliac disease")

AND

ts=(diagnosis OR diagnoses OR diagnostic OR diagnose OR diagnosing OR endomysial OR transglutaminase* OR serolog* OR antibody OR antibodies OR leucocyte* OR hla OR biopsy OR biopsies OR test OR tests OR testing OR screen OR screening OR screened OR Transglutaminase* OR iga OR ttg OR dgp OR IGG OR mass screening OR "Human leukocyte antigen*" OR endoscop* OR misdiagnos* OR undiagnos*)

AND

ts=(adverse OR harm* OR danger* OR bleed* OR perforat* OR OR safe* OR accident*)

Appendix B. List of Excluded Studies

Not English – N=1

1. Nisihara RM, Kotze LM, Utiyama SR, et al. Celiac disease in children and adolescents with Down syndrome. J Pediatr (Rio J). 2005 Sep-Oct;81(5):373-6. PMID: 16247538.

Not Human – N=1

1. Kilmartin C, Lynch S, Abuzakouk M, et al. Avenin fails to induce a Th1 response in coeliac tissue following in vitro culture. Gut. 2003 Jan;52(1):47-52. PMID: 12477758.

Not About Celiac Disease (CD) - N=12

1. Agostoni M, Fanti L, Gemma M, et al. Adverse events during monitored anesthesia care for GI endoscopy: an 8-year experience. Gastrointest Endosc. 2011 Aug;74(2):266-75. PMID: 21704990.

2. Barkin J, O'Loughlin C. Capsule endoscopy contraindications: complications and how to avoid their occurrence. Gastrointest Endosc Clin N Am. 2004;14:61-5.

3. Bednarska O, Ignatova S, Dahle C, et al. Intraepithelial lymphocyte distribution differs between the bulb and the second part of duodenum. BMC Gastroenterol. 2013;13:111. PMID: 23841671.

4. Kakar S, Nehra V, Murray JA, et al. Significance of intraepithelial lymphocytosis in small bowel biopsy samples with normal mucosal architecture. Am J Gastroenterol. 2003 Sep;98(9):2027-33. PMID: 14499783.

5. Kyriakos N, Karagiannis S, Galanis P, et al. Evaluation of four time-saving methods of reading capsule endoscopy videos. Eur J Gastroenterol Hepatol. 2012 Nov;24(11):1276-80. PMID: 22825645.

6. Lakatos L, Pandur T, David G, et al. Association of extraintestinal manifestations of inflammatory bowel disease in a province of western Hungary with disease phenotype: results of a 25-year follow-up study. World J Gastroenterol. 2003 Oct;9(10):2300-7. PMID: 14562397.

7. Pimentel M, Hwang L, Melmed GY, et al. New clinical method for distinguishing D-IBS from other gastrointestinal conditions causing diarrhea: the LA/IBS diagnostic strategy. Dig Dis Sci. 2010 Jan;55(1):145-9. PMID: 19169820.

8. Rogers MA, Levine DA, Blumberg N, et al. Antigenic challenge in the etiology of autoimmune disease in women. J Autoimmun. 2012 May;38(2-3):J97-J102. PMID: 21880464.

9. Sheiko MA, Feinstein JA, Capocelli KE, et al. The concordance of endoscopic and histologic findings of 1000 pediatric EGDs. Gastrointest Endosc. 2014 Nov 1PMID: 25440693.

10. Sidhu R, Sanders DS, Kapur K, et al. Capsule endoscopy changes patient management in routine clinical practice. Dig Dis Sci. 2007 May;52(5):1382-6. PMID: 17357836.

11. Van Beers EH, Einerhand AWC, Taminiau J, et al. Pediatric duodenal biopsies: Mucosal morphology and glycohydrolase expression do not change along the duodenum. Journal of Pediatric Gastroenterology and Nutrition. 1998 Feb;26(2):186-93. PMID: WOS:000071735600012.

12. Wang SS, Flowers CR, Kadin ME, et al. Medical history, lifestyle, family history, and occupational risk factors for peripheral T-cell lymphomas: the InterLymph Non-Hodgkin Lymphoma Subtypes Project. J Natl Cancer Inst Monogr. 2014 Aug;2014(48):66-75. PMID: 25174027.

Not About Diagnostic Tests - N=150

1. Abbas Z, Raza S, Yakoob J, et al. Varied presentation of celiac disease in Pakistani adults. J Coll Physicians Surg Pak. 2013 Jul;23(7):522-4. PMID: 23823965.

2. Abdullah AM. Aetiology of chronic diarrhoea in children: experience at King Khalid University Hospital, Riyadh, Saudi Arabia. Ann Trop Paediatr. 1994;14(2):111-7. PMID: 7521625.

3. Abraham G, Tye-Din JA, Bhalala OG, et al. Accurate and robust genomic prediction of celiac disease using statistical learning. PLoS Genet. 2014 Feb;10(2):e1004137. PMID: 24550740.

4. Acar S, Yetkiner AA, Ersin N, et al. Oral findings and salivary parameters in children with celiac disease: a preliminary study. Med Princ Pract. 2012;21(2):129-33. PMID: 22024774.

5. Alakoski A, Salmi TT, Hervonen K, et al. Chronic Gastritis in Dermatitis Herpetiformis: A Controlled Study. Clinical & Developmental Immunology. 2012PMID: WOS:000303749900001.

6. Atteno M, Costa L, Cozzolino A, et al. The enthesopathy of celiac patients: effects of gluten-free diet. Clin Rheumatol. 2014 Apr;33(4):537-41. PMID: 24567238.

7. Aydogdu S, Cakir M, Yuksekkaya HA, et al. Helicobacter pylori infection in children with celiac disease. Scand J Gastroenterol. 2008;43(9):1088-93. PMID: 18609161.

8. Bakker SF, Pouwer F, Tushuizen ME, et al. Compromised quality of life in patients with both Type 1 diabetes mellitus and coeliac disease. Diabet Med. 2013 Jul;30(7):835-9. PMID: 23534496.

9. Ban L, West J, Abdul Sultan A, et al. Limited risks of major congenital anomalies in children of mothers with coeliac disease: a population-based cohort study. Bjog. 2014 Oct 7PMID: 25288361.

10. Bannister EG, Cameron DJ, Ng J, et al. Can celiac serology alone be used as a marker of duodenal mucosal recovery in children with celiac disease on a gluten-free diet? Am J Gastroenterol. 2014 Sep;109(9):1478-83. PMID: 25070050.

11. Barisani D, Ceroni S, Meneveri R, et al. IL-10 polymorphisms are associated with earlyonset celiac disease and severe mucosal damage in patients of Caucasian origin. Genet Med. 2006 Mar;8(3):169-74. PMID: 16540751.

12. Barnea L, Mozer-Glassberg Y, Hojsak I, et al. Pediatric Celiac Disease Patients Who Are Lost to Follow-Up Have a Poorly Controlled Disease. Digestion. 2014 Dec 19;90(4):248-53. PMID: 25531121.

13. Barratt SM, Leeds JS, Sanders DS. Factors influencing the type, timing and severity of symptomatic responses to dietary gluten in patients with biopsy-proven coeliac disease. J Gastrointestin Liver Dis. 2013 Dec;22(4):391-6. PMID: 24369320.

14. Basso MS, Luciano R, Ferretti F, et al. Association between celiac disease and primary lactase deficiency. Eur J Clin Nutr. 2012 Dec;66(12):1364-5. PMID: 23211657.

15. Bergamaschi G, Markopoulos K, Albertini R, et al. Anemia of chronic disease and defective erythropoietin production in patients with celiac disease. Haematologica. 2008 Dec;93(12):1785-91. PMID: 18815191.

16. Biagi F, Marchese A, Ferretti F, et al. A multicentre case control study on complicated coeliac disease: two different patterns of natural history, two different prognoses. BMC Gastroenterol. 2014;14:139. PMID: 25103857.

17. Biagi F, Trotta L, Alfano C, et al. Prevalence and natural history of potential celiac disease in adult patients. Scand J Gastroenterol. 2013 May;48(5):537-42. PMID: 23506211.

18. Biagi F, Vattiato C, Agazzi S, et al. A second duodenal biopsy is necessary in the follow-up of adult coeliac patients. Ann Med. 2014 Sep;46(6):430-3. PMID: 24857202.

19. Bode S, Gudmand-Hoyer E. Symptoms and haematologic features in consecutive adult coeliac patients. Scand J Gastroenterol. 1996 Jan;31(1):54-60. PMID: 8927941.

20. Boot H. Diagnosis and staging in gastrointestinal lymphoma. Best Pract Res Clin Gastroenterol. 2010 Feb;24(1):3-12. PMID: 20206103.

21. Borrelli M, Salvati VM, Maglio M, et al. Immunoregulatory pathways are active in the small intestinal mucosa of patients with potential celiac disease. Am J Gastroenterol. 2013 Nov;108(11):1775-84. PMID: 24060758.

22. Brandimarte G, Tursi A, Giorgetti GM. Changing trends in clinical form of celiac disease. Which is now the main form of celiac disease in clinical practice? Minerva Gastroenterol Dietol. 2002 Jun;48(2):121-30. PMID: 16489303.

23. Brar P, Kwon GY, Egbuna, II, et al. Lack of correlation of degree of villous atrophy with severity of clinical presentation of coeliac disease. Digestive and Liver Disease. 2007 Jan;39(1):26-9. PMID: WOS:000245931100004.

24. Bystrom IM, Hollen E, Falth-Magnusson K, et al. Health-related quality of life in children and adolescents with celiac disease: from the perspectives of children and parents. Gastroenterol Res Pract. 2012;2012:986475. PMID: 22548054.

25. Campisi G, Di Liberto C, Carroccio A, et al. Coeliac disease: oral ulcer prevalence, assessment of risk and association with gluten-free diet in children. Dig Liver Dis. 2008 Feb;40(2):104-7. PMID: 18063428.

26. Casella G, Antonelli E, Di Bella C, et al. Prevalence and causes of abnormal liver function in patients with coeliac disease. Liver Int. 2013 Aug;33(7):1128-31. PMID: 23601438.

27. Cassio A, Ricci G, Baronio F, et al. Long-term clinical significance of thyroid autoimmunity in children with celiac disease. J Pediatr. 2010 Feb;156(2):292-5. PMID: 19846116.

28. Cellier C, Delabesse E, Helmer C, et al. Refractory sprue, coeliac disease, and enteropathyassociated T-cell lymphoma. French Coeliac Disease Study Group. Lancet. 2000 Jul 15;356(9225):203-8. PMID: 10963198.

29. Chang M, Green PH. Genetic testing before serologic screening in relatives of patients with celiac disease as a cost containment method. J Clin Gastroenterol. 2009 Jan;43(1):43-50. PMID: 19020464.

30. Cheng J, Malahias T, Brar P, et al. The association between celiac disease, dental enamel defects, and aphthous ulcers in a United States cohort. J Clin Gastroenterol. 2010 Mar;44(3):191-4. PMID: 19687752.

31. Chow MA, Lebwohl B, Reilly NR, et al. Immunoglobulin A deficiency in celiac disease. J Clin Gastroenterol. 2012 Nov-Dec;46(10):850-4. PMID: 22476042.

32. Ciacci C, Cirillo M, Giorgetti G, et al. Low plasma cholesterol: A correlate of nondiagnosed celiac disease in adults with hypochromic anemia. American Journal of Gastroenterology. 1999 Jul;94(7):1888-91. PMID: WOS:000081224000030.

33. Cooper SE, Kennedy NP, Mohamed BM, et al. Immunological indicators of coeliac disease activity are not altered by long-term oats challenge. Clin Exp Immunol. 2013 Mar;171(3):313-8. PMID: 23379438.

34. Cummins AG, Alexander BG, Chung A, et al. Morphometric evaluation of duodenal biopsies in celiac disease. Am J Gastroenterol. 2011 Jan;106(1):145-50. PMID: 20736938.

35. de Graaf AP, Westerhof J, Weersma RK, et al. Correlation between predicted and actual consequences of capsule endoscopy on patient management. Digestive and Liver Disease. 2008 Sep;40(9):761-6. PMID: WOS:000259520800009.

36. De Marchi S, Chiarioni G, Prior M, et al. Young adults with coeliac disease may be at increased risk of early atherosclerosis. Aliment Pharmacol Ther. 2013 Jul;38(2):162-9. PMID: 23730933.

37. Di Simone N, Silano M, Castellani R, et al. Anti-tissue transglutaminase antibodies from celiac patients are responsible for trophoblast damage via apoptosis in vitro. Am J Gastroenterol. 2010 Oct;105(10):2254-61. PMID: 20571491.

38. Dickey W, Kenny BD, McMillan SA, et al. Gastric as well as duodenal biopsies may be useful in the investigation of iron deficiency anaemia. Scand J Gastroenterol. 1997 May;32(5):469-72. PMID: 9175209.

39. Egan LJ, Walsh SV, Stevens FM, et al. Celiac-associated lymphoma. A single institution experience of 30 cases in the combination chemotherapy era. J Clin Gastroenterol. 1995 Sep;21(2):123-9. PMID: 8583077.

40. Elfstrom P, Granath F, Ekstrom Smedby K, et al. Risk of lymphoproliferative malignancy in relation to small intestinal histopathology among patients with celiac disease. J Natl Cancer Inst. 2011 Mar 2;103(5):436-44. PMID: 21289299.

41. Elli L, Bonura A, Garavaglia D, et al. Immunological comorbity in coeliac disease: associations, risk factors and clinical implications. J Clin Immunol. 2012 Oct;32(5):984-90. PMID: 22526595.

42. Elli L, Contiero P, Tagliabue G, et al. Risk of intestinal lymphoma in undiagnosed coeliac disease: results from a registered population with different coeliac disease prevalence. Dig Liver Dis. 2012 Sep;44(9):743-7. PMID: 22677003.

43. Errichiello S, Esposito O, Di Mase R, et al. Celiac disease: predictors of compliance with a gluten-free diet in adolescents and young adults. J Pediatr Gastroenterol Nutr. 2010 Jan;50(1):54-60. PMID: 19644397.

44. Fabiani E, Catassi C, Villari A, et al. Dietary compliance in screening-detected coeliac disease adolescents. Acta Paediatr Suppl. 1996 May;412:65-7. PMID: 8783764.

45. Ford S, Howard R, Oyebode J. Psychosocial aspects of coeliac disease: a cross-sectional survey of a UK population. Br J Health Psychol. 2012 Nov;17(4):743-57. PMID: 22502725.

46. Fotoulaki M, Nousia-Arvanitakis S, Augoustidou-Savvopoulou P, et al. Clinical application of immunological markers as monitoring tests in celiac disease. Dig Dis Sci. 1999 Oct;44(10):2133-8. PMID: 10548368.

47. Freeman HJ. Clinical Spectrum of Biopsy-Defined Celiac-Disease in the Elderly. Canadian Journal of Gastroenterology. 1995 Jan-Feb;9(1):42-6. PMID: WOS:A1995QJ30600009.

48. Fuchs V, Kurppa K, Huhtala H, et al. Factors associated with long diagnostic delay in celiac disease. Scand J Gastroenterol. 2014 Nov;49(11):1304-10. PMID: 25139307.

49. Gasbarrini G, Ciccocioppo R, De Vitis I, et al. Coeliac Disease in the Elderly. A multicentre Italian study. Gerontology. 2001 Nov-Dec;47(6):306-10. PMID: 11721143.

50. Gobbi G, Bouquet F, Greco L, et al. Coeliac disease, epilepsy, and cerebral calcifications. The Italian Working Group on Coeliac Disease and Epilepsy. Lancet. 1992 Aug 22;340(8817):439-43. PMID: 1354781.

51. Goldacre MJ, Wotton CJ, Seagroatt V, et al. Cancers and immune related diseases associated with Down's syndrome: a record linkage study. Arch Dis Child. 2004 Nov;89(11):1014-7. PMID: 15499053.

52. Gopee E, van den Oever EL, Cameron F, et al. Coeliac disease, gluten-free diet and the development and progression of albuminuria in children with type 1 diabetes. Pediatr Diabetes. 2013 Sep;14(6):455-8. PMID: 23763501.

53. Gornowicz-Porowska J, Bowszyc-Dmochowska M, Seraszek-Jaros A, et al. Association between Levels of IgA Antibodies to Tissue Transglutaminase and Gliadin-Related Nonapeptides in Dermatitis Herpetiformis. Scientific World Journal. 2012PMID: WOS:000303244700001.

54. Gray AM, Papanicolas IN. Impact of symptoms on quality of life before and after diagnosis of coeliac disease: results from a UK population survey. BMC Health Serv Res. 2010;10:105. PMID: 20423498.

55. Green PH, Fleischauer AT, Bhagat G, et al. Risk of malignancy in patients with celiac disease. Am J Med. 2003 Aug 15;115(3):191-5. PMID: 12935825.

56. Green PHR, Stavropoulos SN, Panagi SG, et al. Characteristics of adult celiac disease in the USA: results of a national survey. Am J Gastroenterol. 2001 Jan;96(1):126-31. PMID: 11197241.

57. Hansen D, Brock-Jacobsen B, Lund E, et al. Clinical benefit of a gluten-free diet in type 1 diabetic children with screening-detected celiac disease: a population-based screening study with 2 years' follow-up. Diabetes Care. 2006 Nov;29(11):2452-6. PMID: 17065683.

58. Harewood GC, Holub JL, Lieberman DA. Variation in small bowel biopsy performance among diverse endoscopy settings: results from a national endoscopic database. Am J Gastroenterol. 2004 Sep;99(9):1790-4. PMID: 15330920.

59. Hogen Esch CE, Wolters VM, Gerritsen SA, et al. Specific celiac disease antibodies in children on a gluten-free diet. Pediatrics. 2011 Sep;128(3):547-52. PMID: 21859913.

60. Hull CM, Liddle M, Hansen N, et al. Elevation of IgA anti-epidermal transglutaminase antibodies in dermatitis herpetiformis. Br J Dermatol. 2008 Jul;159(1):120-4. PMID: 18503599.

61. Ilus T, Kaukinen K, Virta LJ, et al. Incidence of malignancies in diagnosed celiac patients: a population-based estimate. Am J Gastroenterol. 2014 Sep;109(9):1471-7. PMID: 25047399.

62. Ilyas M, Niedobitek G, Agathanggelou A, et al. Non-Hodgkin's lymphoma, coeliac disease, and Epstein-Barr virus: a study of 13 cases of enteropathy-associated T- and B-cell lymphoma. J Pathol. 1995 Oct;177(2):115-22. PMID: 7490676.

63. Jansen MA, Kiefte-de Jong JC, Gaillard R, et al. Growth Trajectories and Bone Mineral Density in Anti-Tissue Transglutaminase Antibody-positive Children: The Generation R Study. Clin Gastroenterol Hepatol. 2014 Sep 22PMID: 25245626.

64. Johnston SD, Rodgers C, Watson RG. Quality of life in screen-detected and typical coeliac disease and the effect of excluding dietary gluten. Eur J Gastroenterol Hepatol. 2004 Nov;16(12):1281-6. PMID: 15618833.

65. Kaila B, Orr K, Bernstein CN. The anti-Saccharomyces cerevisiae antibody assay in a province-wide practice: accurate in identifying cases of Crohn's disease and predicting inflammatory disease. Can J Gastroenterol. 2005 Dec;19(12):717-21. PMID: 16341311.

66. Khashan AS, Kenny LC, McNamee R, et al. Undiagnosed coeliac disease in a father does not influence birthweight and preterm birth. Paediatr Perinat Epidemiol. 2010 Jul 1;24(4):363-9. PMID: 20618726.

67. Koskela RM, Niemela SE, Karttunen TJ, et al. Clinical characteristics of collagenous and lymphocytic colitis. Scand J Gastroenterol. 2004 Sep;39(9):837-45. PMID: 15513381.

68. Krishnareddy S, Lewis SK, Green PH. Dermatitis herpetiformis: clinical presentations are independent of manifestations of celiac disease. Am J Clin Dermatol. 2014 Feb;15(1):51-6. PMID: 24293087.

69. Krupa-Kozak U. Pathologic bone alterations in celiac disease: etiology, epidemiology, and treatment. Nutrition. 2014 Jan;30(1):16-24. PMID: 24290593.

70. Kumar M, Rastogi A, Bhadada SK, et al. Effect of zoledronic acid on bone mineral density in patients of celiac disease: a prospective, randomized, pilot study. Indian J Med Res. 2013 Dec;138(6):882-7. PMID: 24521630.

71. Kuokkanen M, Myllyniemi M, Vauhkonen M, et al. A biopsy-based quick test in the diagnosis of duodenal hypolactasia in upper gastrointestinal endoscopy. Endoscopy. 2006 Jul;38(7):708-12. PMID: 16761211.

72. Lahdeaho ML, Kaukinen K, Collin P, et al. Celiac disease: from inflammation to atrophy: a long-term follow-up study. J Pediatr Gastroenterol Nutr. 2005 Jul;41(1):44-8. PMID: 15990629.

73. Lahdeaho ML, Kaukinen K, Laurila K, et al. The Glutenase ALV003 Attenuates Gluten-Induced Mucosal Injury in Patients with Celiac Disease. Gastroenterology. 2014 Feb 25PMID: 24583059.

74. Lebwohl B, Bhagat G, Markoff S, et al. Prior endoscopy in patients with newly diagnosed celiac disease: a missed opportunity? Dig Dis Sci. 2013 May;58(5):1293-8. PMID: 23361572.

75. Lebwohl B, Granath F, Ekbom A, et al. Mucosal healing and risk for lymphoproliferative malignancy in celiac disease: a population-based cohort study. Ann Intern Med. 2013 Aug 6;159(3):169-75. PMID: 23922062.

76. Lebwohl B, Stephansson O, Green PH, et al. Mucosal Healing in Patients With Celiac Disease and Outcomes of Pregnancy: A Nationwide Population-Based Study. Clin Gastroenterol Hepatol. 2014 Nov 21PMID: 25460563.

77. Leslie C, Mews C, Charles A, et al. Celiac disease and eosinophilic esophagitis: a true association. J Pediatr Gastroenterol Nutr. 2010 Apr;50(4):397-9. PMID: 19841598.

78. Leslie LA, Lebwohl B, Neugut AI, et al. Incidence of lymphoproliferative disorders in patients with celiac disease. Am J Hematol. 2012 Aug;87(8):754-9. PMID: 22641457.

79. Lewy H, Meirson H, Laron Z. Seasonality of birth month of children with celiac disease differs from that in the general population and between sexes and is linked to family history and environmental factors. J Pediatr Gastroenterol Nutr. 2009 Feb;48(2):181-5. PMID: 19179880.

80. Ludvigsson JF, Brandt L, Montgomery SM, et al. Validation study of villous atrophy and small intestinal inflammation in Swedish biopsy registers. BMC Gastroenterol. 2009;9:19. PMID: 19284576.

81. Ludvigsson JF, Ludvigsson J. Stressful life events, social support and confidence in the pregnant woman and risk of coeliac disease in the offspring. Scand J Gastroenterol. 2003 May;38(5):516-21. PMID: 12795462.

82. Ludvigsson JF, Ludvigsson J, Ekbom A, et al. Celiac disease and risk of subsequent type 1 diabetes: a general population cohort study of children and adolescents. Diabetes Care. 2006 Nov;29(11):2483-8. PMID: 17065689.

83. Ludvigsson JF, Montgomery SM, Ekbom A. Celiac disease and risk of adverse fetal outcome: a population-based cohort study. Gastroenterology. 2005 Aug;129(2):454-63. PMID: 16083702.

84. Ludvigsson JF, Montgomery SM, Ekbom A. Smoking and celiac disease: a populationbased cohort study. Clin Gastroenterol Hepatol. 2005 Sep;3(9):869-74. PMID: 16234024.

85. Ludvigsson JF, Montgomery SM, Ekbom A, et al. Small-intestinal histopathology and mortality risk in celiac disease. JAMA. 2009 Sep 16;302(11):1171-8. PMID: 19755695.

86. Ludvigsson JF, Sellgren C, Runeson B, et al. Increased suicide risk in coeliac disease--a Swedish nationwide cohort study. Dig Liver Dis. 2011 Aug;43(8):616-22. PMID: 21419726.

87. MacCulloch K, Rashid M. Factors affecting adherence to a gluten-free diet in children with celiac disease. Paediatr Child Health. 2014 Jun;19(6):305-9. PMID: 25332660.

88. Mackinder M, Allison G, Svolos V, et al. Nutritional status, growth and disease management in children with single and dual diagnosis of type 1 diabetes mellitus and coeliac disease. BMC Gastroenterol. 2014;14:99. PMID: 24885742.

89. Malamut G, Chandesris O, Verkarre V, et al. Enteropathy associated T cell lymphoma in celiac disease: a large retrospective study. Dig Liver Dis. 2013 May;45(5):377-84. PMID: 23313469.

90. Mazzeo T, Brambillasca F, Pellegrini N, et al. Evaluation of visual and taste preferences of some gluten-free commercial products in a group of celiac children. Int J Food Sci Nutr. 2014 Feb;65(1):112-6. PMID: 24079778.

91. Mohseninejad L, Feenstra T, van der Horst HE, et al. Targeted screening for Coeliac Disease among irritable bowel syndrome patients: analysis of cost-effectiveness and value of information. Eur J Health Econ. 2013 Dec;14(6):947-57. PMID: 23179163.

92. Monzani A, Rapa A, Fonio P, et al. Use of deamidated gliadin peptide antibodies to monitor diet compliance in childhood celiac disease. J Pediatr Gastroenterol Nutr. 2011 Jul;53(1):55-60. PMID: 21694536.

93. Nachman F, del Campo MP, Gonzalez A, et al. Long-term deterioration of quality of life in adult patients with celiac disease is associated with treatment noncompliance. Dig Liver Dis. 2010 Oct;42(10):685-91. PMID: 20399159.

94. Newnham E, Shepherd SJ, Hosking P, et al. Success and timecourse of reaching the therapeutic goals with a gluten-free diet in coeliac disease: A prospective five-year study from diagnosis. Journal of Gastroenterology and Hepatology. 2014 Oct;29:129-. PMID: WOS:000343863000253.

95. Nordyke K, Norstrom F, Lindholm L, et al. Health-related quality-of-life in children with coeliac disease, measured prior to receiving their diagnosis through screening. J Med Screen. 2011;18(4):187-92. PMID: 22106434.

96. Nordyke K, Rosen A, Emmelin M, et al. Internalizing the threat of risk--a qualitative study about adolescents' experience living with screening-detected celiac disease 5 years after diagnosis. Health Qual Life Outcomes. 2014;12:91. PMID: 24915870.

97. Osman M, Taha B, Al Duboni G. Assessment of the response to gluten-free diet in an Iraqi population with coeliac disease. A histological and serological follow-up study. Arch Med Sci. 2014 May 12;10(2):294-9. PMID: 24904663.

98. Pantaleoni S, Luchino M, Adriani A, et al. Bone mineral density at diagnosis of celiac disease and after 1 year of gluten-free diet. ScientificWorldJournal. 2014;2014:173082. PMID: 25379519.

99. Potter DD, Murray JA, Donohue JH, et al. The role of defective mismatch repair in small bowel adenocarcinoma in celiac disease. Cancer Res. 2004 Oct 1;64(19):7073-7. PMID: 15466202.

100. Pozo-Rubio T, Capilla A, Mujico JR, et al. Influence of breastfeeding versus formula feeding on lymphocyte subsets in infants at risk of coeliac disease: the PROFICEL study. Eur J Nutr. 2013 Mar;52(2):637-46. PMID: 22576041.

101. Pulido O, Zarkadas M, Dubois S, et al. Clinical features and symptom recovery on a gluten-free diet in Canadian adults with celiac disease. Can J Gastroenterol. 2013 Aug;27(8):449-53. PMID: 23936873.

102. Rajaguru P, Vaiphei K, Saikia B, et al. Increased accumulation of dendritic cells in celiac disease associates with increased expression of autophagy protein LC3. Indian J Pathol Microbiol. 2013 Oct-Dec;56(4):342-8. PMID: 24441219.

103. Rajani S, Sawyer-Bennett J, Shirton L, et al. Patient and parent satisfaction with a dietitianand nurse- led celiac disease clinic for children at the Stollery Children's Hospital, Edmonton, Alberta. Can J Gastroenterol. 2013 Aug;27(8):463-6. PMID: 23936876.

104. Reunala T, Salmi TT, Hervonen K, et al. IgA anti-epidermal transglutaminase antibodies in dermatitis herpetiformis: a significant but not complete response to a gluten-free diet treatment. Br J Dermatol. 2014 Sep 5PMID: 25196300.

105. Riddle MS, Murray JA, Porter CK. The incidence and risk of celiac disease in a healthy US adult population. Am J Gastroenterol. 2012 Aug;107(8):1248-55. PMID: 22584218.

106. Roma E, Roubani A, Kolia E, et al. Dietary compliance and life style of children with coeliac disease. J Hum Nutr Diet. 2010 Apr;23(2):176-82. PMID: 20163513.

107. Roos S, Karner A, Hallert C. Psychological well-being of adult coeliac patients treated for 10 years. Dig Liver Dis. 2006 Mar;38(3):177-80. PMID: 16461026.

108. Sainsbury K, Mullan B, Sharpe L. A randomized controlled trial of an online intervention to improve gluten-free diet adherence in celiac disease. Am J Gastroenterol. 2013 May;108(5):811-7. PMID: 23458849.

109. Sategna-Guidetti C, Pulitano R, Grosso S, et al. Serum IgA antiendomysium antibody titers as a marker of intestinal involvement and diet compliance in adult celiac sprue. J Clin Gastroenterol. 1993 Sep;17(2):123-7. PMID: 8409314.

110. Sategna-Guidetti C, Volta U, Ciacci C, et al. Prevalence of thyroid disorders in untreated adult celiac disease patients and effect of gluten withdrawal: an Italian multicenter study. Am J Gastroenterol. 2001 Mar;96(3):751-7. PMID: 11280546.

111. Sedghizadeh PP, Shuler CF, Allen CM, et al. Celiac disease and recurrent aphthous stomatitis: a report and review of the literature. Oral Surg Oral Med Oral Pathol Oral Radiol Endod. 2002 Oct;94(4):474-8. PMID: 12374923.

112. Selimoglu MA, Kelles M, Erdem T, et al. Craniofacial features of children with celiac disease. Eur J Gastroenterol Hepatol. 2013 Oct;25(10):1206-11. PMID: 23799417.

113. Shah S, Akbari M, Vanga R, et al. Patient Perception of Treatment Burden Is High in Celiac Disease Compared With Other Common Conditions. American Journal of Gastroenterology. 2014 Sep;109(9):1304-11. PMID: WOS:000344459800001.

114. Shahnaz A, Maguire G, Parker R, et al. Tissue transglutaminase antibody levels predict IgA deficiency. Arch Dis Child. 2013 Nov;98(11):873-6. PMID: 23928648.

115. Shaoul R, Marcon MA, Okada Y, et al. Gastric metaplasia: a frequently overlooked feature of duodenal biopsy specimens in untreated celiac disease. J Pediatr Gastroenterol Nutr. 2000 Apr;30(4):397-403. PMID: 10776950.

116. Sharaiha RZ, Lebwohl B, Reimers L, et al. Increasing incidence of enteropathy-associated T-cell lymphoma in the United States, 1973-2008. Cancer. 2012 Aug 1;118(15):3786-92. PMID: 22169928.

117. Sharkey LM, Corbett G, Currie E, et al. Optimising delivery of care in coeliac disease - comparison of the benefits of repeat biopsy and serological follow-up. Aliment Pharmacol Ther. 2013 Nov;38(10):1278-91. PMID: 24117503.

118. Silano M, Volta U, Mecchia AM, et al. Delayed diagnosis of coeliac disease increases cancer risk. BMC Gastroenterol. 2007;7:8. PMID: 17349035.

119. Smecuol E, Hwang HJ, Sugai E, et al. Exploratory, randomized, double-blind, placebocontrolled study on the effects of Bifidobacterium infantis natren life start strain super strain in active celiac disease. J Clin Gastroenterol. 2013 Feb;47(2):139-47. PMID: 23314670.

120. Smukalla S, Lebwohl B, Mears JG, et al. How often do hematologists consider celiac disease in iron-deficiency anemia? Results of a national survey. Clin Adv Hematol Oncol. 2014 Feb;12(2):100-5. PMID: 24892255.

121. Solaymani-Dodaran M, West J, Logan RF. Long-term mortality in people with celiac disease diagnosed in childhood compared with adulthood: a population-based cohort study. Am J Gastroenterol. 2007 Apr;102(4):864-70. PMID: 17324126.

122. Sponzilli I, Chiari G, Iovane B, et al. Celiac disease in children with type 1 diabetes: impact of gluten free diet on diabetes management. Acta Biomed. 2010 Dec;81(3):165-70. PMID: 22530453.

123. Stagi S, Rigante D, Lepri G, et al. Evaluation of autoimmune phenomena in patients with pediatric autoimmune neuropsychiatric disorders associated with streptococcal infections (PANDAS). Autoimmunity Reviews. 2014 Dec;13(12):1236-40. PMID: WOS:000345203800010.

124. Sugai E, Nachman F, Vaquez H, et al. Dynamics of celiac disease-specific serology after initiation of a gluten-free diet and use in the assessment of compliance with treatment. Dig Liver Dis. 2010 May;42(5):352-8. PMID: 19679520.

125. Szaflarska-Poplawska A, Krakowska A. Influence of specific dietary interventions on clinical manifestation of coeliac disease. Przeglad Gastroenterologiczny. 2010;5(1):24-30. PMID: WOS:000276471400004.

126. Taavela J, Kurppa K, Collin P, et al. Degree of damage to the small bowel and serum antibody titers correlate with clinical presentation of patients with celiac disease. Clin Gastroenterol Hepatol. 2013 Feb;11(2):166-71 e1. PMID: 23063678.

127. Tapsas D, Falth-Magnusson K, Hogberg L, et al. Swedish children with celiac disease comply well with a gluten-free diet, and most include oats without reporting any adverse effects: a long-term follow-up study. Nutrition Research. 2014 May;34(5):436-41. PMID: WOS:000337715700008.

128. Teufel A, Weinmann A, Kahaly GJ, et al. Concurrent autoimmune diseases in patients with autoimmune hepatitis. J Clin Gastroenterol. 2010 Mar;44(3):208-13. PMID: 20087196.

129. Toft-Hansen H, Nielsen C, Biagini M, et al. Lectin staining shows no evidence of involvement of glycocalyx/mucous layer carbohydrate structures in development of celiac disease. Nutrients. 2013 Nov;5(11):4540-52. PMID: 24253051.

130. Tontini GE, Rondonotti E, Saladino V, et al. Impact of gluten withdrawal on healthrelated quality of life in celiac subjects: an observational case-control study. Digestion. 2010;82(4):221-8. PMID: 20588037.

131. Tortora R, Capone P, Imperatore N, et al. Predictive value of "Marsh 1" type histology in subjects with suspected cealic disease. Scand J Gastroenterol. 2014 Jul;49(7):801-6. PMID: 24958090.

132. Trotta L, Biagi F, Bianchi PI, et al. Dental enamel defects in adult coeliac disease: prevalence and correlation with symptoms and age at diagnosis. Eur J Intern Med. 2013 Dec;24(8):832-4. PMID: 23571066.

133. Tursi A, Brandimarte G, Giorgetti GM, et al. Endoscopic and histological findings in the duodenum of adults with celiac disease before and after changing to a gluten-free diet: a 2-year prospective study. Endoscopy. 2006 Jul;38(7):702-7. PMID: 16810593.

134. Ukkola A, Maki M, Kurppa K, et al. Diet improves perception of health and well-being in symptomatic, but not asymptomatic, patients with celiac disease. Clin Gastroenterol Hepatol. 2011 Feb;9(2):118-23. PMID: 21029791.

135. Ukkola A, Maki M, Kurppa K, et al. Changes in body mass index on a gluten-free diet in coeliac disease: a nationwide study. Eur J Intern Med. 2012 Jun;23(4):384-8. PMID: 22560391.

136. Uspenskaya ID, Shirokova NY. Duodenal mucosa in children with coeliac disease in catamnesis and varying compliance with the gluten-free diet. Bratisl Lek Listy. 2014;115(3):150-5. PMID: 24579684.

137. Vakiani E, Arguelles-Grande C, Mansukhani MM, et al. Collagenous sprue is not always associated with dismal outcomes: a clinicopathological study of 19 patients. Mod Pathol. 2010 Jan;23(1):12-26. PMID: 19855376.

138. van Doorn RK, Winkler LM, Zwinderman KH, et al. CDDUX: a disease-specific healthrelated quality-of-life questionnaire for children with celiac disease. J Pediatr Gastroenterol Nutr. 2008 Aug;47(2):147-52. PMID: 18664865.

139. Vecsei E, Steinwendner S, Kogler H, et al. Follow-up of pediatric celiac disease: value of antibodies in predicting mucosal healing, a prospective cohort study. BMC Gastroenterol. 2014;14:28. PMID: 24524430.

140. Vermeersch P, Richter T, Hauer AC, et al. Use of likelihood ratios improves clinical interpretation of IgG and IgA anti-DGP antibody testing for celiac disease in adults and children. Clinical Biochemistry. 2011 Feb;44(2-3):248-50. PMID: WOS:000286959100019.

141. Viitasalo L, Niemi L, Ashorn M, et al. Early Microbial Markers of Celiac Disease. J Clin Gastroenterol. 2014 Feb 10PMID: 24518796.

142. Viljamaa M, Kaukinen K, Huhtala H, et al. Coeliac disease, autoimmune diseases and gluten exposure. Scand J Gastroenterol. 2005 Apr;40(4):437-43. PMID: 16028438.

143. Volta U, Caio G, Stanghellini V, et al. The changing clinical profile of celiac disease: a 15year experience (1998-2012) in an Italian referral center. BMC Gastroenterol. 2014;14:194. PMID: 25404189.

144. Vriezinga SL, Auricchio R, Bravi E, et al. Randomized feeding intervention in infants at high risk for celiac disease. N Engl J Med. 2014 Oct 2;371(14):1304-15. PMID: 25271603.

145. Wacklin P, Kaukinen K, Tuovinen E, et al. The duodenal microbiota composition of adult celiac disease patients is associated with the clinical manifestation of the disease. Inflamm Bowel Dis. 2013 Apr;19(5):934-41. PMID: 23478804.

146. Wei L, Spiers E, Reynolds N, et al. The association between coeliac disease and cardiovascular disease. Aliment Pharmacol Ther. 2008 Mar 15;27(6):514-9. PMID: 18162081.

147. Weinstock LB, Walters AS, Mullin GE, et al. Celiac disease is associated with restless legs syndrome. Dig Dis Sci. 2010 Jun;55(6):1667-73. PMID: 19731029.

148. Welander A, Tjernberg AR, Montgomery SM, et al. Infectious disease and risk of later celiac disease in childhood. Pediatrics. 2010 Mar;125(3):e530-6. PMID: 20176673.

149. Weston WL, Morelli JG, Huff JC. Misdiagnosis, treatments, and outcomes in the immunobullous diseases in children. Pediatric Dermatology. 1997 Jul-Aug;14(4):264-72. PMID: WOS:A1997XP81800002.

150. Zanini B, Basche R, Ferraresi A, et al. Factors that contribute to hypertransaminasemia in patients with celiac disease or functional gastrointestinal syndromes. Clin Gastroenterol Hepatol. 2014 May;12(5):804-10.e2. PMID: 24211290.

No Original Data - N=20

1. Alfonsi P, Iannuzzi V. Re: Scoglio et al. Is intestinal biopsy always needed for diagnosis of celiac disease? Am J Gastroenterol. 2004 May;99(5):963. PMID: 15128375.

2. Bizzaro N, Villalta D, Tonutti E, et al. Association of celiac disease with connective tissue diseases and autoimmune diseases of the digestive tract. Autoimmun Rev. 2003 Oct;2(6):358-63. PMID: 14550877.

3. Brigic E, Hadzic D, Mladina N. Early and correct diagnosis of celiac disease in the prevention of growth disorders and child development. Mater Sociomed. 2012;24(4):242-7. PMID: 23678328.

4. Cave D, Legnani P, de Franchis R, et al. ICCE consensus for capsule retention. Endoscopy. 2005 Oct;37(10):1065-7. PMID: 16189792.

5. Committee ASoP, Ben-Menachem T, Decker GA, et al. Adverse events of upper GI endoscopy. Gastrointest Endosc. 2012 Oct;76(4):707-18. PMID: 22985638.

6. Davies-Shaw J, Marcon MA, Assor E, et al. A randomized controlled trial to evaluate the efficacy and safety of a gluten-free diet in patients with asymptomatic celiac disease and type 1 diabetes. Celiac disease and diabetes-dietary intervention and evaluation trial (CD-DIET). Pediatric diabetes; 2014. p. 114.

7. Delvaux M, Gay G. Capsule endoscopy: technique and indications. Best Pract Res Clin Gastroenterol. 2008;22(5):813-37. PMID: 18790434.

8. Egner W, Shrimpton A, Sargur R, et al. ESPGHAN guidance on coeliac disease 2012: multiples of ULN for decision making do not harmonise assay performance across centres. J Pediatr Gastroenterol Nutr. 2012 Dec;55(6):733-5. PMID: 22744189.

9. Freeman HJ. Survey of gastroenterologists on the diagnosis and treatment of adult patients with celiac disease in British Columbia. Can J Gastroenterol. 1998 Mar;12(2):149-52. PMID: 9559209.

10. Husby S, Koletzko S, Korbonay-Szabo I. Can We Really Skip the Biopsy in Diagnosing Symptomatic Children With Celiac Disease Reply. Journal of Pediatric Gastroenterology and Nutrition. 2013 Oct;57(4)PMID: WOS:000326745500004.

11. Malamut G, Cellier C. Is refractory celiac disease more severe in old Europe? Am J Gastroenterol. 2011 May;106(5):929-32. PMID: 21540899.

12. Murray JA, Green PH. Biopsy is the gold standard of diagnosis of celiac sprue. Gastroenterology. 1999 May;116(5):1273-4. PMID: 10220528.

13. Parakkal D, Du H, Semer R, et al. Do gastroenterologists adhere to diagnostic and treatment guidelines for celiac disease? J Clin Gastroenterol. 2012 Feb;46(2):e12-20. PMID: 21959324.

14. Radaelli F, Minoli G, Bardella MT, et al. Celiac disease among patients referred for routine upper gastrointestinal endoscopy: prevalence and diagnostic accuracy of duodenal endoscopic markers. Am J Gastroenterol. 2000 Apr;95(4):1089-90. PMID: 10763970.

15. Riestra S, Dominguez F, Fernandez-Ruiz E, et al. Usefulness of duodenal biopsy during routine upper gastrointestinal endoscopy for diagnosis of celiac disease. World J Gastroenterol. 2006 Aug 21;12(31):5028-32. PMID: 16937500.

16. Rostom A, Murray JA, Kagnoff MF. American Gastroenterological Association (AGA) Institute technical review on the diagnosis and management of celiac disease. Gastroenterology. 2006 Dec;131(6):1981-2002. PMID: 17087937. 17. Samasca G, Iancu M, Baican A, et al. Romanian experience in child celiac disease diagnosis. Roum Arch Microbiol Immunol. 2011 Oct-Dec;70(4):178-85. PMID: 22568266.

18. Shamir R, Eliakim R. Capsule endoscopy in pediatric patients. World J Gastroenterol. 2008 Jul 14;14(26):4152-5. PMID: 18636660.

19. Simpson SM, Ciaccio EJ, Case S, et al. Celiac disease in patients with type 1 diabetes: screening and diagnostic practices. Diabetes Educ. 2013 Jul-Aug;39(4):532-40. PMID: 23674375.

20. Urbanski SJ. Invited review controversial issue: Can duodenal mucosa appear normal in gluten-sensitive enteropathy (celiac disease)? International Journal of Surgical Pathology. 1998 Jan;6(1):49-54. PMID: WOS:000075463200009.

Individual Case Report - N=19

1. Biagi F, Bianchi PI, Zilli A, et al. The significance of duodenal mucosal atrophy in patients with common variable immunodeficiency: a clinical and histopathologic study. Am J Clin Pathol. 2012 Aug;138(2):185-9. PMID: 22904128.

2. Brar P, Lee AR, Lewis SK, et al. Celiac disease in African-Americans. Dig Dis Sci. 2006 May;51(5):1012-5. PMID: 16642428.

3. Brocchi E, Tomassetti P, Volta U, et al. Adult coeliac disease diagnosed by endoscopic biopsies in the duodenal bulb. Eur J Gastroenterol Hepatol. 2005 Dec;17(12):1413-5. PMID: 16292098.

4. Chang MS, Rubin M, Lewis SK, et al. Diagnosing celiac disease by video capsule endoscopy (VCE) when esophagogastroduodenoscopy (EGD) and biopsy is unable to provide a diagnosis: a case series. BMC Gastroenterol. 2012;12:90. PMID: 22812595.

5. Ciccocioppo R, Bernardo ME, Russo ML, et al. Allogeneic hematopoietic stem cell transplantation may restore gluten tolerance in patients with celiac disease. J Pediatr Gastroenterol Nutr. 2013 Apr;56(4):422-7. PMID: 23531481.

6. Cohen SA, Ephrath H, Lewis JD, et al. Pediatric capsule endoscopy: review of the small bowel and patency capsules. J Pediatr Gastroenterol Nutr. 2012 Mar;54(3):409-13. PMID: 21760541.

7. Cuenca-Abente F, Nachman F, Bai JC. Diagnosis of celiac disease during pre-operative workup for bariatric surgery. Acta Gastroenterol Latinoam. 2012 Dec;42(4):321-4. PMID: 23383526.

8. da Silva Kotze LM, Nisihara R, Kotze LR, et al. Celiac disease and dermatitis herpetiformis in Brazilian twins: a long-term follow-up and screening of their relatives. J Pediatr Endocrinol Metab. 2013;26(1-2):71-5. PMID: 23329745.

9. Dickey W, Hughes D. Erosions in the second part of the duodenum in patients with villous atrophy. Gastrointest Endosc. 2004 Jan;59(1):116-8. PMID: 14722564.

10. Dorum S, Bodd M, Fallang LE, et al. HLA-DQ molecules as affinity matrix for identification of gluten T cell epitopes. J Immunol. 2014 Nov 1;193(9):4497-506. PMID: 25261484.

11. Friedel DM. Big bleed after endoscopic mucosal biopsy. South Med J. 2009 Feb;102(2):129. PMID: 19139710.

12. Gora-Gebka M, Wozniak M, Cielecka-Kuszyk J, et al. Graves' disease, celiac disease and liver function abnormalities in a patient--clinical manifestation and diagnostic difficulties. Acta Biochim Pol. 2014;61(2):281-4. PMID: 24904927.

13. Hu WT, Murray JA, Greenaway MC, et al. Cognitive impairment and celiac disease. Arch Neurol. 2006 Oct;63(10):1440-6. PMID: 17030661.

14. Kurppa K, Collin P, Lindfors K, et al. Spontaneous negative seroconversion of endomysial antibodies does not exclude subsequent celiac disease. J Pediatr Gastroenterol Nutr. 2011 Nov;53(5):576-9. PMID: 22020541.

15. Lurie Y, Landau DA, Pfeffer J, et al. Celiac disease diagnosed in the elderly. Journal of Clinical Gastroenterology. 2008 Jan;42(1):59-61. PMID: WOS:000252160900012.

16. Nenna R, Mennini M, Petrarca L, et al. Immediate effect on fertility of a gluten-free diet in women with untreated coeliac disease. Gut. 2011 Jul;60(7):1023-4. PMID: 21193444.

17. Tack GJ, van de Water JM, Bruins MJ, et al. Consumption of gluten with gluten-degrading enzyme by celiac patients: a pilot-study. World J Gastroenterol. 2013 Sep 21;19(35):5837-47. PMID: 24124328.

18. Taylor E, Dickson-Swift V, Anderson K. Coeliac disease: the path to diagnosis and the reality of living with the disease. J Hum Nutr Diet. 2013 Aug;26(4):340-8. PMID: 23190398.

19. Venkatesh K, Abou-Taleb A, Cohen M, et al. Role of confocal endomicroscopy in the diagnosis of celiac disease. J Pediatr Gastroenterol Nutr. 2010 Sep;51(3):274-9. PMID: 20531027.

Prevalence Only - N=41

1. Alessandrini S, Giacomoni E, Muccioli F. Mass population screening for celiac disease in children: the experience in Republic of San Marino from 1993 to 2009. Ital J Pediatr. 2013;39:67. PMID: 24152602.

2. Armes J, Gee DC, Macrae FA, et al. Collagenous colitis: jejunal and colorectal pathology. J Clin Pathol. 1992 Sep;45(9):784-7. PMID: 1401208.

3. Assiri AM. Isolated short stature as a presentation of celiac disease in Saudi children. Pediatr Rep. 2010;2(1):e4. PMID: 21589840.

4. Bhatnagar S, Gupta SD, Mathur M, et al. Celiac disease with mild to moderate histologic changes is a common cause of chronic diarrhea in Indian children. J Pediatr Gastroenterol Nutr. 2005 Aug;41(2):204-9. PMID: 16056100.

5. Bybrant MC, Ortqvist E, Lantz S, et al. High prevalence of celiac disease in Swedish children and adolescents with type 1 diabetes and the relation to the Swedish epidemic of celiac disease: a cohort study. Scand J Gastroenterol. 2014 Jan;49(1):52-8. PMID: 24164443.

6. Cannings-John R, Butler CC, Prout H, et al. A case-control study of presentations in general practice before diagnosis of coeliac disease. Br J Gen Pract. 2007 Aug;57(541):636-42. PMID: 17688758.

7. Cristofori F, Fontana C, Magista A, et al. Increased Prevalence of Celiac Disease Among Pediatric Patients With Irritable Bowel Syndrome: A 6-Year Prospective Cohort Study. JAMA Pediatr. 2014 Apr 21PMID: 24756157.

8. Fluge G, Olesen HV, Gilljam M, et al. Co-morbidity of cystic fibrosis and celiac disease in Scandinavian cystic fibrosis patients. J Cyst Fibros. 2009 May;8(3):198-202. PMID: 19303374.

9. Giannotti A, Tiberio G, Castro M, et al. Coeliac disease in Williams syndrome. J Med Genet. 2001 Nov;38(11):767-8. PMID: 11694549.

10. Greco L, Timpone L, Abkari A, et al. Burden of celiac disease in the Mediterranean area. World J Gastroenterol. 2011 Dec 7;17(45):4971-8. PMID: 22174546.

11. Grodzinsky E, Ivarsson A, Juto P, et al. New automated immunoassay measuring immunoglobulin A antigliadin antibodies for prediction of celiac disease in childhood. Clin Diagn Lab Immunol. 2001 May;8(3):564-70. PMID: 11329459.

12. Hadithi M, de Boer H, Meijer JW, et al. Coeliac disease in Dutch patients with Hashimoto's thyroiditis and vice versa. World J Gastroenterol. 2007 Mar 21;13(11):1715-22. PMID: 17461476.

13. Isikay S, Hizli S, Yilmaz K. Prevalence of Celiac Disease in Turkish Children with Idiopathic Epilepsy. Iranian Journal of Pediatrics. 2014 Jun;24(3):280-4. PMID: WOS:000337731700008.

14. Kingham JG, Parker DR. The association between primary biliary cirrhosis and coeliac disease: a study of relative prevalences. Gut. 1998 Jan;42(1):120-2. PMID: 9518232.

15. Liu E, Lee HS, Aronsson CA, et al. Risk of pediatric celiac disease according to HLA haplotype and country. N Engl J Med. 2014 Jul 3;371(1):42-9. PMID: 24988556.

16. Lu W, Gwee KA, Siah KT, et al. Prevalence of Anti-deamidated Gliadin Peptide Antibodies in Asian Patients With Irritable Bowel Syndrome. J Neurogastroenterol Motil. 2014 Apr 30;20(2):236-41. PMID: 24840376.

17. Maiwall R, Goel A, Pulimood AB, et al. Investigation into celiac disease in Indian patients with portal hypertension. Indian J Gastroenterol. 2014 Sep 18PMID: 25231910.

18. Makovicky P, Rimarova K, Boor A, et al. Correlation between antibodies and histology in celiac disease: incidence of celiac disease is higher than expected in the pediatric population. Mol Med Rep. 2013 Oct;8(4):1079-83. PMID: 23942815.

19. Mathew J, Kaur G. Risk of pediatric celiac disease according to HLA haplotype and country. Indian Pediatrics. 2014 Sep;51(9):733-7. PMID: WOS:000342430000013.

20. Pooni PA, Chhina RS, Jaina BK, et al. Clinical and anthropometric profile of children with celiac disease in Punjab (North India). J Trop Pediatr. 2006 Feb;52(1):30-3. PMID: 15947010.

21. Ress K, Annus T, Putnik U, et al. Celiac disease in children with atopic dermatitis. Pediatr Dermatol. 2014 Jul-Aug;31(4):483-8. PMID: 24831884.

22. Rodrigo L, Blanco I, Bobes J, et al. Remarkable prevalence of coeliac disease in patients with irritable bowel syndrome plus fibromyalgia in comparison with those with isolated irritable bowel syndrome: a case-finding study. Arthritis Res Ther. 2013;15(6):R201. PMID: 24283458.

23. Rostami-Nejad M, Villanacci V, Mashayakhi R, et al. Celiac disease and Hp infection association in Iran. Rev Esp Enferm Dig. 2009 Dec;101(12):850-4. PMID: 20082545.

24. Sanders DS, Hurlstone DP, Stokes RO, et al. Changing face of adult coeliac disease: experience of a single university hospital in South Yorkshire. Postgrad Med J. 2002 Jan;78(915):31-3. PMID: 11796869.

25. Savilahti E, Kolho KL, Westerholm-Ormio M, et al. Clinics of coeliac disease in children in the 2000s. Acta Paediatr. 2010 Jul;99(7):1026-30. PMID: 20199495.

26. Selva-O'Callaghan A, Casellas F, de Torres I, et al. Celiac disease and antibodies associated with celiac disease in patients with inflammatory myopathy. Muscle Nerve. 2007 Jan;35(1):49-54. PMID: 16967485.

27. Stagi S, Lapi E, D'Avanzo MG, et al. Coeliac disease and risk for other autoimmune diseases in patients with Williams-Beuren syndrome. BMC Med Genet. 2014;15:61. PMID: 24885139.

28. Stagi S, Simonini G, Ricci L, et al. Coeliac disease in patients with Kawasaki disease. Is there a link? Rheumatology (Oxford). 2006 Jul;45(7):847-50. PMID: 16418194.

29. Szodoray P, Barta Z, Lakos G, et al. Coeliac disease in Sjogren's syndrome--a study of 111 Hungarian patients. Rheumatol Int. 2004 Sep;24(5):278-82. PMID: 13680146.

30. Thevenot T, Denis J, Jouannaud V, et al. Coeliac disease in chronic hepatitis C: a French multicentre prospective study. Aliment Pharmacol Ther. 2007 Nov 1;26(9):1209-16. PMID: 17944735.

31. Toftedal P, Hansen DG, Nielsen C, et al. Questionnaire-based case finding of celiac disease in a population of 8- to 9-year-old children. Pediatrics. 2010 Mar;125(3):e518-24. PMID: 20123763.

32. Trovato CM, Albanese CV, Leoni S, et al. Lack of clinical predictors for low mineral density in children with celiac disease. J Pediatr Gastroenterol Nutr. 2014 Dec;59(6):799-802. PMID: 25162363.

33. Tucci F, Astarita L, Abkari A, et al. Celiac disease in the Mediterranean area. BMC Gastroenterol. 2014;14:24. PMID: 24517104.

34. Ucardag D, Guliter S, Ceneli O, et al. Celiac disease prevalence in patients with iron deficiency anemia of obscure origin. Turk J Gastroenterol. 2009 Dec;20(4):266-70. PMID: 20084570.

35. van der Pals M, Myleus A, Norstrom F, et al. Body mass index is not a reliable tool in predicting celiac disease in children. BMC Pediatr. 2014;14:165. PMID: 24981433.

36. van Gerven NM, Bakker SF, de Boer YS, et al. Seroprevalence of celiac disease in patients with autoimmune hepatitis. European Journal of Gastroenterology & Hepatology. 2014 Oct;26(10):1104-7. PMID: WOS:000341980200006.

37. Varma S, Malhotra P, Kochhar R, et al. Celiac disease presenting as iron-deficiency anemia in northern India. Indian J Gastroenterol. 2001 Nov-Dec;20(6):234-6. PMID: 11817777.

38. Ventura A, Ronsoni MF, Shiozawa MB, et al. Prevalence and clinical features of celiac disease in patients with autoimmune thyroiditis: cross-sectional study. Sao Paulo Med J. 2014 Dec;132(6):364-71. PMID: 25351758.

39. White LE, Bannerman E, McGrogan P, et al. Childhood coeliac disease diagnoses in Scotland 2009-2010: the SPSU project. Arch Dis Child. 2013 Jan;98(1):52-6. PMID: 23184350.

40. Wingren CJ, Bjorck S, Lynch KF, et al. Coeliac disease in children: a social epidemiological study in Sweden. Acta Paediatr. 2012 Feb;101(2):185-91. PMID: 21824189.

41. Wroblova K, Kolorz M, Pav I, et al. Frequencies of HLA-DQ2 and HLA-DQ8 haplotypes in Czech and Slovak coeliac patients and the healthy population. Acta Biochim Pol. 2014;61(1):191-3. PMID: 24660172.

Diagnostic Method Outside the Scope of Study - N=155

1. Al Saidi SS, Al Harthi SO, Mula-Abed WA. Diagnostic utility of coeliac disease: a descriptive study in a tertiary care hospital, oman. Oman Med J. 2013 Jul;28(4):232-6. PMID: 23904914.

2. al-Tawaty AI, Elbargathy SM. Coeliac disease in north-eastern Libya. Ann Trop Paediatr. 1998 Mar;18(1):27-30. PMID: 9691998.

3. Altuntas B, Filik B, Ensari A, et al. Can zinc deficiency be used as a marker for the diagnosis of celiac disease in Turkish children with short stature? Pediatr Int. 2000 Dec;42(6):682-4. PMID: 11192528.

4. Aoun JP, Moukarbel N, Aftimos G. Value of duodenal endoscopic markers of villous atrophy. J Med Liban. 2001 Nov-Dec;49(6):319-24. PMID: 12744633.

5. Araya M, Mondragon A, Perez-Bravo F, et al. Celiac disease in a Chilean population carrying Amerindian traits. J Pediatr Gastroenterol Nutr. 2000 Oct;31(4):381-6. PMID: 11045834.

6. Bardella MT, Minoli G, Radaelli F, et al. Reevaluation of duodenal endoscopic markers in the diagnosis of celiac disease. Gastrointest Endosc. 2000 Jun;51(6):714-6. PMID: 10840306.

7. Barera G, Bianchi C, Calisti L, et al. Screening of diabetic children for coeliac disease with antigliadin antibodies and HLA typing. Arch Dis Child. 1991 Apr;66(4):491-4. PMID: 2031607.

8. Bonamico M, Sabbatella L, Di Tola M, et al. Antiendomysial antibody detection in biopsy culture allows avoidance of gluten challenge in celiac children. J Pediatr Gastroenterol Nutr. 2005 Feb;40(2):165-9; discussion 22-4. PMID: 15699690.

9. Bonamico M, Scire G, Mariani P, et al. Short stature as the primary manifestation of monosymptomatic celiac disease. J Pediatr Gastroenterol Nutr. 1992 Jan;14(1):12-6. PMID: 1573504.

10. Burginwolff A, Gaze H, Hadziselimovic F, et al. Antigliadin and Antiendomysium Antibody Determination for Celiac-Disease. Archives of Disease in Childhood. 1991 Aug;66(8):941-7. PMID: WOS:A1991GA66700009.

11. Calero P, Ribes-Koninckx C, Albiach V, et al. IgA antigliadin antibodies as a screening method for nonovert celiac disease in children with insulin-dependent diabetes mellitus. J Pediatr Gastroenterol Nutr. 1996 Jul;23(1):29-33. PMID: 8811520.

12. Cammarota G, Cazzato A, Genovese O, et al. Water-immersion technique during standard upper endoscopy may be useful to drive the biopsy sampling of duodenal mucosa in children with celiac disease. J Pediatr Gastroenterol Nutr. 2009 Oct;49(4):411-6. PMID: 19581815.

13. Cammarota G, Cesaro P, Cazzato A, et al. The water immersion technique is easy to learn for routine use during EGD for duodenal villous evaluation: a single-center 2-year experience. J Clin Gastroenterol. 2009 Mar;43(3):244-8. PMID: 18813029.

14. Cammarota G, Cesaro P, Martino A, et al. High accuracy and cost-effectiveness of a biopsyavoiding endoscopic approach in diagnosing coeliac disease. Aliment Pharmacol Ther. 2006 Jan 1;23(1):61-9. PMID: 16393281.

15. Cammarota G, Martino A, Pirozzi GA, et al. Direct visualization of intestinal villi by high-resolution magnifying upper endoscopy: a validation study. Gastrointest Endosc. 2004 Nov;60(5):732-8. PMID: 15557949.

16. Campanella J, Biagi F, Bianchi PI, et al. Clinical response to gluten withdrawal is not an indicator of coeliac disease. Scand J Gastroenterol. 2008;43(11):1311-4. PMID: 18609173.

17. Card TR, Siffledeen J, West J, et al. An excess of prior irritable bowel syndrome diagnoses or treatments in Celiac disease: evidence of diagnostic delay. Scand J Gastroenterol. 2013 Jul;48(7):801-7. PMID: 23697749.

18. Carroccio A, Brusca I, Iacono G, et al. IgA anti-actin antibodies ELISA in coeliac disease: a multicentre study. Dig Liver Dis. 2007 Sep;39(9):818-23. PMID: 17652043.

19. Casella G, Villanacci V, Di-Bella C, et al. Colonoscopic findings in coeliac disease on a gluten-free diet. Rev Esp Enferm Dig. 2010 Sep;102(9):538-41. PMID: 20883070.

20. Castro M, Crino A, Papadatou B, et al. Downs-Syndrome and Celiac-Disease - the Prevalence of High Iga-Antigliadin Antibodies and Hla-Dr and Dq Antigens in Trisomy-21. Journal of Pediatric Gastroenterology and Nutrition. 1993 Apr;16(3):265-8. PMID: WOS:A1993KY38000007.

21. Ciaccio EJ, Tennyson CA, Bhagat G, et al. Quantitative estimates of motility from videocapsule endoscopy are useful to discern celiac patients from controls. Dig Dis Sci. 2012 Nov;57(11):2936-43. PMID: 22644741.

22. Collado MC, Donat E, Ribes-Koninckx C, et al. Specific duodenal and faecal bacterial groups associated with paediatric coeliac disease. J Clin Pathol. 2009 Mar;62(3):264-9. PMID: 18996905.

23. Collin P, Korpela M, Hallstrom O, et al. Rheumatic complaints as a presenting symptom in patients with coeliac disease. Scand J Rheumatol. 1992;21(1):20-3. PMID: 1570482.

24. Corazza GR, Caletti GC, Lazzari R, et al. Scalloped duodenal folds in childhood celiac disease. Gastrointest Endosc. 1993 Jul-Aug;39(4):543-5. PMID: 8365604.

25. Corazza GR, Villanacci V, Zambelli C, et al. Comparison of the interobserver reproducibility with different histologic criteria used in celiac disease. Clin Gastroenterol Hepatol. 2007 Jul;5(7):838-43. PMID: 17544877.

26. de Freitas IN, Sipahi AM, Damiao AO, et al. Celiac disease in Brazilian adults. J Clin Gastroenterol. 2002 Apr;34(4):430-4. PMID: 11907355.

27. De Luca L, Ricciardiello L, Rocchi MB, et al. Narrow band imaging with magnification endoscopy for celiac disease: results from a prospective, single-center study. Diagn Ther Endosc. 2013;2013:580526. PMID: 23983448.

28. Dell'Aquila P, Pietrini L, Barone M, et al. Small intestinal contrast ultrasonography-based scoring system: a promising approach for the diagnosis and follow-up of celiac disease. J Clin Gastroenterol. 2005 Aug;39(7):591-5. PMID: 16000926.

29. Dickey W. Diagnosis of coeliac disease at open-access endoscopy. Scand J Gastroenterol. 1998 Jun;33(6):612-5. PMID: 9669633.

30. Dickey W, Hughes D. Prevalence of celiac disease and its endoscopic markers among patients having routine upper gastrointestinal endoscopy. Am J Gastroenterol. 1999 Aug;94(8):2182-6. PMID: 10445547.

31. Dickey W, Hughes D. Disappointing sensitivity of endoscopic markers for villous atrophy in a high-risk population: implications for celiac disease diagnosis during routine endoscopy. Am J Gastroenterol. 2001 Jul;96(7):2126-8. PMID: 11467643.

32. Dutta AK, Sajith KG, Shah G, et al. Duodenal villous morphology assessed using magnification narrow band imaging correlates well with histology in patients with suspected malabsorption syndrome. Digestive Endoscopy. 2014 Nov;26(6):720-5. PMID: WOS:000344591400005.

33. El-Shabrawi M, El-Karaksy H, Mohsen N, et al. Celiac disease in children and adolescents with autoimmune hepatitis: a single-centre experience. J Trop Pediatr. 2011 Apr;57(2):104-8. PMID: 20571152.

34. Eltumi MA, Ong PS, Francis ND, et al. A comparison of endoscopic and capsule small intestinal biopsy techniques in children with upper gastrointestinal disorders. Journal of Paediatrics and Child Health. 1996 Jun;32(3):255-6. PMID: WOS:A1996UX85800014.

35. Emami MH, Karimi S, Nemati A. Do endoscopic markers still play a role in the diagnosis of celiac disease? Indian J Gastroenterol. 2008 Sep-Oct;27(5):183-5. PMID: 19112186.

36. Erriu M, Abbate GM, Pili FM, et al. Oral Signs and HLA-DQB1 *02 Haplotypes in the Celiac Paediatric Patient: A Preliminary Study. Autoimmune Dis. 2013;2013:389590. PMID: 24198965.

37. Erriu M, Sanna S, Nucaro A, et al. HLA-DQB1 Haplotypes and their Relation to Oral Signs Linked to Celiac Disease Diagnosis. Open Dent J. 2011;5:174-8. PMID: 22135701.

38. Fernandez-Banares F, Carrasco A, Garcia-Puig R, et al. Intestinal intraepithelial lymphocyte cytometric pattern is more accurate than subepithelial deposits of anti-tissue transglutaminase IgA for the diagnosis of celiac disease in lymphocytic enteritis. PLoS One. 2014;9(7):e101249. PMID: 25010214.

39. Freeman HJ. Detection of adult celiac disease with duodenal screening biopsies over a 30-year period. Can J Gastroenterol. 2013 Jul;27(7):405-8. PMID: 23862172.

40. Gadd S, Kamath KR, Silink M, et al. Co-existence of coeliac disease and insulin-dependent diabetes mellitus in children: screening sera using an ELISA test for gliadin antibody. Aust N Z J Med. 1992 Jun;22(3):256-60. PMID: 1497552.

41. Garampazzi A, Rapa A, Mura S, et al. Clinical pattern of celiac disease is still changing. J Pediatr Gastroenterol Nutr. 2007 Nov;45(5):611-4. PMID: 18030243.

42. Gasbarrini A, Ojetti V, Cuoco L, et al. Lack of endoscopic visualization of intestinal villi with the "immersion technique" in overt atrophic celiac disease. Gastrointest Endosc. 2003 Mar;57(3):348-51. PMID: 12612514.

43. Gianfrani C, Troncone R, Mugione P, et al. Celiac disease association with CD8+ T cell responses: identification of a novel gliadin-derived HLA-A2-restricted epitope. J Immunol. 2003 Mar 1;170(5):2719-26. PMID: 12594302.

44. Gonen C, Yilmaz N, Yalcin M, et al. Diagnostic yield of routine duodenal biopsies in iron deficiency anaemia: a study from Western Anatolia. Eur J Gastroenterol Hepatol. 2007 Jan;19(1):37-41. PMID: 17206075.

45. Goswami A, Dadhich S, Bhargava N. Use of narrow band imaging in assessing duodenal villous atrophy. Indian J Gastroenterol. 2014 Sep;33(5):440-4. PMID: 25015746.

46. Granito A, Muratori P, Cassani F, et al. Anti-actin IgA antibodies in severe coeliac disease. Clin Exp Immunol. 2004 Aug;137(2):386-92. PMID: 15270857.

47. Grodzinsky E, Hed J, Lieden G, et al. Presence of IgA and IgG antigliadin antibodies in healthy adults as measured by micro-ELISA. Effect of various cutoff levels on specificity and sensitivity when diagnosing coeliac disease. Int Arch Allergy Appl Immunol. 1990;92(2):119-23. PMID: 2242925.

48. Gunther U, Daum S, Heller F, et al. Diagnostic value of confocal endomicroscopy in celiac disease. Endoscopy. 2010 Mar;42(3):197-202. PMID: 20195989.

49. Guresci S, Hizli S, Simsek GG. The Value of Ensari's Proposal in Evaluating the Mucosal Pathology of Childhood Celiac Disease: Old Classification versus New Version. Balkan Medical Journal. 2012;29(3):281-4. PMID: WOS:000315506200011.

50. Hansson T, Dannaeus A, Kraaz W, et al. Production of antibodies to gliadin by peripheral blood lymphocytes in children with celiac disease: the use of an enzyme-linked immunospot technique for screening and follow-up. Pediatr Res. 1997 Apr;41(4 Pt 1):554-9. PMID: 9098859.

51. Hartman C, Hino B, Lerner A, et al. Bone quantitative ultrasound and bone mineral density in children with celiac disease. J Pediatr Gastroenterol Nutr. 2004 Nov;39(5):504-10. PMID: 15572890.

52. Heneghan MA, Stevens FM, Cryan EM, et al. Celiac sprue and immunodeficiency states: a 25-year review. J Clin Gastroenterol. 1997 Sep;25(2):421-5. PMID: 9412941.

53. Hutchinson JM, West NP, Robins GG, et al. Long-term histological follow-up of people with coeliac disease in a UK teaching hospital. QJM. 2010 Jul;103(7):511-7. PMID: 20519276.

54. Ikram MA, Sajid A, Hameed S, et al. Coeliac disease in children presenting with failure to thrive. J Ayub Med Coll Abbottabad. 2011 Oct-Dec;23(4):6-9. PMID: 23472398.

55. Jevon GP, Dimmick JE, Dohil R, et al. Spectrum of gastritis in celiac disease in childhood. Pediatric and Developmental Pathology. 1999 May-Jun;2(3):221-6. PMID: WOS:000079556000006.

56. Joda H, Beni V, Alakulppi N, et al. Medium-high resolution electrochemical genotyping of HLA-DQ2/DQ8 for detection of predisposition to coeliac disease. Anal Bioanal Chem. 2014 Mar 15PMID: 24633503.

57. Johnson JE, Johnson KE. Ambiguous chronic illness in women: community health nursing concern. J Community Health Nurs. 2006 Fall;23(3):159-67. PMID: 16863401.

58. Johnston SD, Watson RG, McMillan SA, et al. Serological markers for coeliac disease: changes with time and relationship to enteropathy. Eur J Gastroenterol Hepatol. 1998 Mar;10(3):259-64. PMID: 9585032.

59. Johnston SD, Watson RG, McMillan SA, et al. Preliminary results from follow-up of a large-scale population survey of antibodies to gliadin, reticulin and endomysium. Acta Paediatr Suppl. 1996 May;412:61-4. PMID: 8783763.

60. Johnston SD, Watson RG, Middleton D, et al. Genetic, morphometric and immunohistochemical markers of latent coeliac disease. Eur J Gastroenterol Hepatol. 1999 Nov;11(11):1283-8. PMID: 10563541.

61. Karinen H, Karkkainen P, Pihlajamaki J, et al. Gene dose effect of the DQB1*0201 allele contributes to severity of coeliac disease. Scandinavian Journal of Gastroenterology. 2006 Feb;41(2):191-9. PMID: WOS:000235275800011.

62. Kasirer Y, Turner D, Lerman L, et al. Scalloping is a reliable endoscopic marker for celiac disease. Dig Endosc. 2014 Mar;26(2):232-5. PMID: 23746050.

63. Kaukinen K, Peraaho M, Collin P, et al. Small-bowel mucosal transglutaminase 2-specific IgA deposits in coeliac disease without villous atrophy: a prospective and randomized clinical study. Scand J Gastroenterol. 2005 May;40(5):564-72. PMID: 16036509.

64. Kepczyk MT, Kadakia CSC. Prospective Evaluation of Gastrointestinal-Tract in Patients with Iron-Deficiency Anemia. Digestive Diseases and Sciences. 1995 Jun;40(6):1283-9. PMID: WOS:A1995RE07000021.

65. Knudtzon J, Fluge G, Aksnes L. Routine measurements of gluten antibodies in children of short stature. J Pediatr Gastroenterol Nutr. 1991 Feb;12(2):190-4. PMID: 2051271.

66. Kolho KL, Farkkila MA, Savilahti E. Undiagnosed coeliac disease is common in Finnish adults. Scand J Gastroenterol. 1998 Dec;33(12):1280-3. PMID: 9930391.

67. Korponay-Szabo IR, Kovacs JB, Lorincz M, et al. Prospective significance of antiendomysium antibody positivity in subsequently verified celiac disease. J Pediatr Gastroenterol Nutr. 1997 Jul;25(1):56-63. PMID: 9226528.

68. Koskinen L, Romanos J, Kaukinen K, et al. Cost-effective HLA typing with tagging SNPs predicts celiac disease risk haplotypes in the Finnish, Hungarian, and Italian populations. Immunogenetics. 2009 Apr;61(4):247-56. PMID: 19255754.

69. Koskinen O, Collin P, Lindfors K, et al. Usefulness of small-bowel mucosal transglutaminase-2 specific autoantibody deposits in the diagnosis and follow-up of celiac disease. J Clin Gastroenterol. 2010 Aug;44(7):483-8. PMID: 19779364.

70. Koulaouzidis A, Yung DE, Lam JH, et al. The use of small-bowel capsule endoscopy in iron-deficiency anemia alone; be aware of the young anemic patient. Scand J Gastroenterol. 2012 Sep;47(8-9):1094-100. PMID: 22852553.

71. Kumar PJ, O'Donoghue DP, Stenson K, et al. Reintroduction of gluten in adults and children with treated coeliac disease. Gut. 1979 Sep;20(9):743-9. PMID: 499912.

72. Kurppa K, Lauronen O, Collin P, et al. Factors associated with dietary adherence in celiac disease: a nationwide study. Digestion. 2012;86(4):309-14. PMID: 23095439.

73. Lankisch PG, Martinez Schramm A, Petersen F, et al. Diagnostic intervals for recognizing celiac disease. Z Gastroenterol. 1996 Aug;34(8):473-7. PMID: 8794542.

74. Laseta F, Salerno G, Buccellato A, et al. Radiographic Indicants of Adult Celiac-Disease Assessed by Double-Contrast Small-Bowel Enteroclysis. European Journal of Radiology. 1992 Sep;15(2):157-62. PMID: WOS:A1992JM91600015.

75. Lebwohl B, Tennyson CA, Holub JL, et al. Sex and racial disparities in duodenal biopsy to evaluate for celiac disease. Gastrointest Endosc. 2012 Oct;76(4):779-85. PMID: 22732871.

76. Lian L, Remzi FH, Kiran RP, et al. Clinical implication of false-positive celiac serology in patients with ileal pouch. Dis Colon Rectum. 2010 Oct;53(10):1446-51. PMID: 20847628.

77. Llorente-Alonso MJ, Fernandez-Acenero MJ, Sebastian M. Gluten intolerance: sex and agerelated features. Can J Gastroenterol. 2006 Nov;20(11):719-22. PMID: 17111054.

78. Lo A, Guelrud M, Essenfeld H, et al. Classification of villous atrophy with enhanced magnification endoscopy in patients with celiac disease and tropical sprue. Gastrointest Endosc. 2007 Aug;66(2):377-82. PMID: 17643717.

79. Lorini R, Scaramuzza A, Vitali L, et al. Clinical aspects of coeliac disease in children with insulin-dependent diabetes mellitus. J Pediatr Endocrinol Metab. 1996 Mar;9 Suppl 1:101-11. PMID: 8887160.

80. Ludvigsson JF, Aro P, Walker MM, et al. Celiac disease, eosinophilic esophagitis and gastroesophageal reflux disease, an adult population-based study. Scand J Gastroenterol. 2013 Jul;48(7):808-14. PMID: 23672638.

81. Magazzu G, Bottari M, Tuccari G, et al. Upper gastrointestinal endoscopy can be a reliable screening tool for celiac sprue in adults. J Clin Gastroenterol. 1994 Oct;19(3):255-7; discussion 7-8. PMID: 7806840.

82. Maglio M, Tosco A, Auricchio R, et al. Intestinal deposits of anti-tissue transglutaminase IgA in childhood celiac disease. Dig Liver Dis. 2011 Aug;43(8):604-8. PMID: 21342796.

83. Mahadev S, Simpson S, Lebwohl B, et al. Is dietitian use associated with celiac disease outcomes? Nutrients. 2013 May;5(5):1585-94. PMID: 23676548.

84. Marsh MN, Bjarnason I, Shaw J, et al. Studies of intestinal lymphoid tissue. XIV--HLA status, mucosal morphology, permeability and epithelial lymphocyte populations in first degree relatives of patients with coeliac disease. Gut. 1990 Jan;31(1):32-6. PMID: 2318429.

85. Matysiak-Budnik T, Coron E, Mosnier JF, et al. In vivo real-time imaging of human duodenal mucosal structures in celiac disease using endocytoscopy. Endoscopy. 2010 Mar;42(3):191-6. PMID: 20101565.

86. Maurino E, Capizzano H, Niveloni S, et al. Value of endoscopic markers in celiac disease. Dig Dis Sci. 1993 Nov;38(11):2028-33. PMID: 8223077.

87. McIntyre AS, Ng DP, Smith JA, et al. The endoscopic appearance of duodenal folds is predictive of untreated adult celiac disease. Gastrointest Endosc. 1992 Mar-Apr;38(2):148-51. PMID: 1568611.

88. Meijer JW, Wahab PJ, Mulder CJ. Small intestinal biopsies in celiac disease: duodenal or jejunal? Virchows Arch. 2003 Feb;442(2):124-8. PMID: 12596062.

89. Mina S, Riga C, Azcurra AI, et al. Oral ecosystem alterations in celiac children: a follow-up study. Arch Oral Biol. 2012 Feb;57(2):154-60. PMID: 21920498.

90. Mooney PD, Kurien M, Evans KE, et al. Point-of-care testing for celiac disease has a low sensitivity in endoscopy. Gastrointest Endosc. 2014 Mar 29PMID: 24685008.

91. Mustalahti K, Lohiniemi S, Collin P, et al. Gluten-free diet and quality of life in patients with screen-detected celiac disease. Eff Clin Pract. 2002 May-Jun;5(3):105-13. PMID: 12088289.

92. Myleus A, Hernell O, Gothefors L, et al. Early infections are associated with increased risk for celiac disease: an incident case-referent study. BMC Pediatr. 2012;12:194. PMID: 23249321.

93. Nieminen U, Kahri A, Savilahti E, et al. Duodenal disaccharidase activities in the follow-up of villous atrophy in coeliac disease. Scand J Gastroenterol. 2001 May;36(5):507-10. PMID: 11346204.

94. Nordyke K, Myleus A, Ivarsson A, et al. How do children experience participating in a coeliac disease screening? A qualitative study based on children's written narratives. Scand J Public Health. 2010 Jun;38(4):351-8. PMID: 20413585.

95. Norstrom F, Lindholm L, Sandstrom O, et al. Delay to celiac disease diagnosis and its implications for health-related quality of life. BMC Gastroenterol. 2011;11:118. PMID: 22060243.

96. O'Leary C, Wieneke P, Buckley S, et al. Celiac disease and irritable bowel-type symptoms. Am J Gastroenterol. 2002 Jun;97(6):1463-7. PMID: 12094866.

97. Olen O, Bihagen E, Rasmussen F, et al. Socioeconomic position and education in patients with coeliac disease. Dig Liver Dis. 2012 Jun;44(6):471-6. PMID: 22341742.

98. O'Loughlin EV, Dutt S, Kamath R, et al. Prospective peer-review audit of paediatric upper gastrointestinal endoscopy. J Paediatr Child Health. 2007 Jul-Aug;43(7-8):551-4. PMID: 17635684.

99. Ortiz M, Fragoso A, O'Sullivan CK. Detection of antigliadin autoantibodies in celiac patient samples using a cyclodextrin-based supramolecular biosensor. Anal Chem. 2011 Apr 15;83(8):2931-8. PMID: 21443170.

100. Oxentenko AS, Grisolano SW, Murray JA, et al. The insensitivity of endoscopic markers in celiac disease. American Journal of Gastroenterology. 2002 Apr;97(4):933-8. PMID: WOS:000175060000024.

101. Paarlahti P, Kurppa K, Ukkola A, et al. Predictors of persistent symptoms and reduced quality of life in treated coeliac disease patients: a large cross-sectional study. BMC Gastroenterol. 2013;13:75. PMID: 23631482.

102. Page SR, Lloyd CA, Hill PG, et al. The prevalence of coeliac disease in adult diabetes mellitus. QJM. 1994 Oct;87(10):631-7. PMID: 7987659.

103. Page SR, Lloyd CA, Hill PG, et al. The Prevalence of Celiac-Disease in Adult Diabetes-Mellitus. Quarterly Journal of Medicine. 1994 Oct;87(10):631-7. PMID: WOS:A1994PN04000007.

104. Paparo F, Petrone E, Tosco A, et al. Clinical, HLA, and small bowel immunohistochemical features of children with positive serum antiendomysium antibodies and architecturally normal small intestinal mucosa. Am J Gastroenterol. 2005 Oct;100(10):2294-8. PMID: 16181383.

105. Parnanen A, Kaukinen K, Helakorpi S, et al. Symptom-detected and screen-detected celiac disease and adult height: a large cohort study. Eur J Gastroenterol Hepatol. 2012 Sep;24(9):1066-70. PMID: 22664941.

106. Parry SD, Welfare MR, Cobden I, et al. Push enteroscopy in a UK district general hospital: experience of 51 cases over 2 years. Eur J Gastroenterol Hepatol. 2002 Mar;14(3):305-9. PMID: 11953697.

107. Pellegrini G, Scotta MS, Soardo S, et al. Elevated IgA anti-gliadin antibodies in juvenile chronic arthritis. Clin Exp Rheumatol. 1991 Nov-Dec;9(6):653-6. PMID: 1764847.

108. Periolo N, Guillen L, Bernardo D, et al. Altered expression of the lymphocyte activation antigen CD30 in active celiac disease. Autoimmunity. 2010 Jun;43(4):288-98. PMID: 20166880.

109. Persliden J, Pettersson HB, Falth-Magnusson K. Small intestinal biopsy in children with coeliac disease: measurement of radiation dose and analysis of risk. Acta Paediatr. 1993 Mar;82(3):296-9. PMID: 8495087.

110. Pettersson HB, Falth-Magnusson K, Persliden J, et al. Radiation risk and cost-benefit analysis of a paediatric radiology procedure: results from a national study. Br J Radiol. 2005 Jan;78(925):34-8. PMID: 15673527.

111. Piazzi L, Zancanella L, Chilovi F, et al. Diagnostic value of endoscopic markers for celiac disease in adults: a multicentre prospective Italian study. Minerva Gastroenterol Dietol. 2008 Dec;54(4):335-46. PMID: 19047974.

112. Picarelli A, Di Tola M, Marino M, et al. Usefulness of the organ culture system when villous height/crypt depth ratio, intraepithelial lymphocyte count, or serum antibody tests are not diagnostic for celiac disease. Transl Res. 2013 Mar;161(3):172-80. PMID: 23177794.

113. Picarelli A, Triglione P, Mariani P, et al. Use of a threshold serum level of anti-gliadin antibodies improves diagnostic efficiency of the test in adult coeliac disease but is unreliable as a screening test. Ital J Gastroenterol. 1996 Feb-Mar;28(2):70-5. PMID: 8781997.

114. Pironti A, Tadeu V, Pedroni A, et al. Role of routine small intestinal biopsy in adult patient with irritable bowel syndrome-like symptoms. Minerva Med. 2010 Jun;101(3):129-34. PMID: 20562801.

115. Pizzuti D, Bortolami M, Mazzon E, et al. Transcriptional downregulation of tight junction protein ZO-1 in active coeliac disease is reversed after a gluten-free diet. Dig Liver Dis. 2004 May;36(5):337-41. PMID: 15191203.

116. Pohl H, Rosch T, Tanczos BT, et al. Endocytoscopy for the detection of microstructural features in adult patients with celiac sprue: a prospective, blinded endocytoscopy-conventional histology correlation study. Gastrointest Endosc. 2009 Nov;70(5):933-41. PMID: 19560762.

117. Pohl H, Tanczos BT, Rudolph B, et al. Probe-based confocal laser microscopy identifies criteria predictive of active celiac sprue. Dig Dis Sci. 2012 Feb;57(2):451-7. PMID: 21901262.

118. Polivy AW, Marion SA, Firebaugh SL, et al. Design of a Capsule Endoscopy Device Less Susceptible to Tumbling. 2012 38th Annual Northeast Bioengineering Conference. New York: Ieee; 2012:235-6.

119. Popp A, Jinga M, Jurcut C, et al. Fingertip rapid point-of-care test in adult case-finding in coeliac disease. BMC Gastroenterol. 2013 Jul 12;13(1):115. PMID: 23849178.

120. Prause C, Richter T, Koletzko S, et al. New developments in serodiagnosis of childhood celiac disease: assay of antibodies against deamidated gliadin. Ann N Y Acad Sci. 2009 Sep;1173:28-35. PMID: 19758128.

121. Qari FA. Clinical presentation of adult celiac disease in Western Saudi Arabia. Saudi Medical Journal. 2002 Dec;23(12):1514-7. PMID: WOS:000180310800017.

122. Reyes H, Niveloni S, Moreno ML, et al. A prospective evaluation of endoscopic markers for identifying celiac disease in patients with high and low probability of having the disease. Acta Gastroenterol Latinoam. 2008 Sep;38(3):178-86. PMID: 18979897.

123. Rolles CJ, McNeish AS. Standardised approach to gluten challenge in diagnosing childhood coeliac disease. Br Med J. 1976 May 29;1(6021):1309-11. PMID: 1268678.

124. Rosen A, Ivarsson A, Nordyke K, et al. Balancing health benefits and social sacrifices: a qualitative study of how screening-detected celiac disease impacts adolescents' quality of life. BMC Pediatr. 2011;11:32. PMID: 21569235.

125. Rosen A, Sandstrom O, Carlsson A, et al. Usefulness of symptoms to screen for celiac disease. Pediatrics. 2014 Feb;133(2):211-8. PMID: 24420802.

126. Sacchetti L, Calcagno G, Ferrajolo A, et al. Discrimination between celiac and other gastrointestinal disorders in childhood by rapid human lymphocyte antigen typing. Clin Chem. 1998 Aug;44(8 Pt 1):1755-7. PMID: 9702970.

127. Saleem N, Ali S, Ahmed TA, et al. HLA-DR alleles among Pakistani patients of coeliac disease. J Pak Med Assoc. 2013 Oct;63(10):1271-4. PMID: 24392558.

128. Salmi TT, Hervonen K, Laurila K, et al. Small bowel transglutaminase 2-specific IgA deposits in dermatitis herpetiformis. Acta Derm Venereol. 2014 Jul;94(4):393-7. PMID: 24352382.

129. Salur L, Uibo O, Talvik I, et al. The high frequency of coeliac disease among children with neurological disorders. Eur J Neurol. 2000 Nov;7(6):707-11. PMID: 11136360.

130. Saukkonen T, Savilahti E, Reijonen H, et al. Coeliac disease: frequent occurrence after clinical onset of insulin-dependent diabetes mellitus. Childhood Diabetes in Finland Study Group. Diabet Med. 1996 May;13(5):464-70. PMID: 8737029.

131. Schweizer JJ, Oren A, Mearin ML. Cancer in children with celiac disease: a survey of the European Society of Paediatric Gastroenterology, Hepatology and Nutrition. J Pediatr Gastroenterol Nutr. 2001 Jul;33(1):97-100. PMID: 11479418.

132. Sheiko MA, Feinstein JA, Capocelli KE, et al. Diagnostic yield of EGD in children: a retrospective single-center study of 1000 cases. Gastrointestinal Endoscopy. 2013 Jul;78(1):47-U224. PMID: WOS:000321490500008.

133. Shirts BH, Bennett ST, Jackson BR. Using patients like my patient for clinical decision support: institution-specific probability of celiac disease diagnosis using simplified near-neighbor classification. J Gen Intern Med. 2013 Dec;28(12):1565-72. PMID: 23645451.

134. Singh P, Arora S, Makharia GK. Presence of anemia in patients with celiac disease suggests more severe disease. Indian J Gastroenterol. 2014 Mar;33(2):161-4. PMID: 24243078.

135. Stenman SM, Lindfors K, Korponay-Szabo IR, et al. Secretion of celiac disease autoantibodies after in vitro gliadin challenge is dependent on small-bowel mucosal transglutaminase 2-specific IgA deposits. BMC Immunol. 2008;9:6. PMID: 18312620.

136. Storm W. Prevalence and diagnostic significance of gliadin antibodies in children with Down syndrome. Eur J Pediatr. 1990 Sep;149(12):833-4. PMID: 2146127.
137. Szakal DN, Gyorffy H, Arato A, et al. Mucosal expression of claudins 2, 3 and 4 in proximal and distal part of duodenum in children with coeliac disease. Virchows Arch. 2010 Mar;456(3):245-50. PMID: 20143085.

138. Tanpowpong P, Obuch JC, Jiang H, et al. Multicenter study on season of birth and celiac disease: evidence for a new theoretical model of pathogenesis. J Pediatr. 2013 Mar;162(3):501-4. PMID: 23084709.

139. Tanure MG, Silva IN, Bahia M, et al. Prevalence of celiac disease in Brazilian children with type 1 diabetes mellitus. J Pediatr Gastroenterol Nutr. 2006 Feb;42(2):155-9. PMID: 16456407.

140. Thijs WJ, van Baarlen J, Kleibeuker JH, et al. Duodenal versus jejunal biopsies in suspected celiac disease. Endoscopy. 2004 Nov;36(11):993-6. PMID: WOS:000225024900010.

141. Thomas AG, Phillips AD, Walker-Smith JA. The value of proximal small intestinal biopsy in the differential diagnosis of chronic diarrhoea. Arch Dis Child. 1992 Jun;67(6):741-3; discussion 3-4. PMID: 1626996.

142. Tortora R, Russo I, De Palma GD, et al. In vitro gliadin challenge: diagnostic accuracy and utility for the difficult diagnosis of celiac disease. Am J Gastroenterol. 2012 Jan;107(1):111-7. PMID: 21946279.

143. Troncone R, Starita A, Coletta S, et al. Antigliadin antibody, D-xylose, and cellobiose/mannitol permeability tests as indicators of mucosal damage in children with coeliac disease. Scand J Gastroenterol. 1992 Aug;27(8):703-6. PMID: 1439555.

144. Uibo O. Childhood celiac disease in Estonia: efficacy of the IgA-class antigliadin antibody test in the search for new cases. J Pediatr Gastroenterol Nutr. 1994 Jan;18(1):53-5. PMID: 8126618.

145. Uibo O, Uibo R, Kleimola V, et al. Serum IgA anti-gliadin antibodies in an adult population sample. High prevalence without celiac disease. Dig Dis Sci. 1993 Nov;38(11):2034-7. PMID: 8223078.

146. Ukkola A, Kurppa K, Collin P, et al. Use of health care services and pharmaceutical agents in coeliac disease: a prospective nationwide study. BMC Gastroenterol. 2012;12:136. PMID: 23016889.

147. Ukkola A, Maki M, Kurppa K, et al. Patients' experiences and perceptions of living with coeliac disease - implications for optimizing care. J Gastrointestin Liver Dis. 2012 Mar;21(1):17-22. PMID: 22457855.

148. Valitutti F, Oliva S, Iorfida D, et al. Narrow band imaging combined with water immersion technique in the diagnosis of celiac disease. Dig Liver Dis. 2014 Dec;46(12):1099-102. PMID: 25224697.

149. Vecsei A, Fuhrmann T, Liedlgruber M, et al. Automated classification of duodenal imagery in celiac disease using evolved Fourier feature vectors. Computer Methods and Programs in Biomedicine. 2009 Aug;95(2):S68-S78. PMID: WOS:000269602100008.

150. Verkasalo MA, Raitakari OT, Viikari J, et al. Undiagnosed silent coeliac disease: a risk for underachievement? Scand J Gastroenterol. 2005 Dec;40(12):1407-12. PMID: 16293555.

151. Villanacci V, Magazzu G, Pellegrino S, et al. Comparison of the Marsh-Oberhuber classification with a new grading system in identifying patients with latent celiac disease. Minerva Gastroenterol Dietol. 2010 Dec;56(4):371-5. PMID: 21139535.

152. Vitoria JC, Arrieta A, Astigarraga I, et al. Use of serological markers as a screening test in family members of patients with celiac disease. J Pediatr Gastroenterol Nutr. 1994 Oct;19(3):304-9. PMID: 7815262.

153. Volta U, Molinaro N, De Franchis R, et al. Correlation between IgA antiendomysial antibodies and subtotal villous atrophy in dermatitis herpetiformis. J Clin Gastroenterol. 1992 Jun;14(4):298-301. PMID: 1607605.

154. Wapenaar MC, Monsuur AJ, Poell J, et al. The SPINK gene family and celiac disease susceptibility. Immunogenetics. 2007 May;59(5):349-57. PMID: 17333166.

155. Zubillaga P, Vitoria JC, Arrieta A, et al. Down's syndrome and celiac disease. J Pediatr Gastroenterol Nutr. 1993 Feb;16(2):168-71. PMID: 8450384.

Test Processing Issue - N=20

1. Cammarota G, Cesaro P, La Mura R, et al. Role of the "immersion technique" in diagnosing celiac disease with villous atrophy limited to the duodenal bulb. J Clin Gastroenterol. 2007 Jul;41(6):571-5. PMID: 17577113.

2. Candon S, Mauvais FX, Garnier-Lengline H, et al. Monitoring of anti-transglutaminase autoantibodies in pediatric celiac disease using a sensitive radiobinding assay. J Pediatr Gastroenterol Nutr. 2012 Mar;54(3):392-6. PMID: 21900830.

3. Ciaccio EJ, Bhagat G, Naiyer AJ, et al. Quantitative assessment of the degree of villous atrophy in patients with coeliac disease. J Clin Pathol. 2008 Oct;61(10):1089-93. PMID: 18641407.

4. Ciaccio EJ, Tennyson CA, Bhagat G, et al. Robust spectral analysis of videocapsule images acquired from celiac disease patients. Biomed Eng Online. 2011;10:78. PMID: 21906318.

5. Ciaccio EJ, Tennyson CA, Bhagat G, et al. Transformation of videocapsule images to detect small bowel mucosal differences in celiac versus control patients. Comput Methods Programs Biomed. 2012 Oct;108(1):28-37. PMID: 22284703.

6. Ciaccio EJ, Tennyson CA, Bhagat G, et al. Implementation of a polling protocol for predicting celiac disease in videocapsule analysis. World J Gastrointest Endosc. 2013 Jul 16;5(7):313-22. PMID: 23858375.

7. Ciaccio EJ, Tennyson CA, Bhagat G, et al. Use of basis images for detection and classification of celiac disease. Biomed Mater Eng. 2014;24(6):1913-23. PMID: 25226887.

8. Cronin CC, Jackson LM, Feighery C, et al. Coeliac disease and epilepsy. QJM. 1998 Apr;91(4):303-8. PMID: 9666954.

9. Dahlbom I, Agardh D, Hansson T. Protein A and protein G ELISA for the detection of IgG autoantibodies against tissue transglutaminase in childhood celiac disease. Clin Chim Acta. 2008 Sep;395(1-2):72-6. PMID: 18514068.

10. Di Pisa M, Buccato P, Sabatino G, et al. Epitope mapping of the N-terminal portion of tissue transglutaminase protein antigen to identify linear epitopes in celiac disease. J Pept Sci. 2014 Sep;20(9):689-95. PMID: 24831711.

11. Dickey W. Endoscopy, serology and histology in the diagnosis of coeliac disease. Dig Liver Dis. 2002 Mar;34(3):172-4. PMID: 11990387.

12. Green PH. Celiac disease: how many biopsies for diagnosis? Gastrointest Endosc. 2008 Jun;67(7):1088-90. PMID: 18513550.

13. Gunther U, Daum S, Zeitz M, et al. Capsule endoscopy: comparison of two different reading modes. Int J Colorectal Dis. 2012 Apr;27(4):521-5. PMID: 22065113.

14. Hammerle-Uhl J, Holler Y, Uhl A, et al. Endoscope distortion correction does not (easily) improve mucosa-based classification of celiac disease. Med Image Comput Comput Assist Interv. 2012;15(Pt 3):574-81. PMID: 23286177.

15. Liu E, Li M, Bao F, et al. Need for quantitative assessment of transglutaminase autoantibodies for celiac disease in screening-identified children. J Pediatr. 2005 Apr;146(4):494-9. PMID: 15812452.

16. Megiorni F, Mora B, Bonamico M, et al. A rapid and sensitive method to detect specific human lymphocyte antigen (HLA) class II alleles associated with celiac disease. Clin Chem Lab Med. 2008;46(2):193-6. PMID: 18076355.

17. Ruiz-Ortiz E, Montraveta M, Cabre E, et al. HLA-DQ2/DQ8 and HLA-DQB1*02 homozygosity typing by real-time polymerase chain reaction for the assessment of celiac disease

genetic risk: evaluation of a Spanish celiac population. Tissue Antigens. 2014 Dec;84(6):545-53. PMID: 25413104.

18. Stern M. Comparative evaluation of serologic tests for celiac disease: a European initiative toward standardization. J Pediatr Gastroenterol Nutr. 2000 Nov;31(5):513-9. PMID: 11144436.

19. Taavela J, Koskinen O, Huhtala H, et al. Validation of morphometric analyses of smallintestinal biopsy readouts in celiac disease. PLoS One. 2013;8(10):e76163. PMID: 24146832.

20. van Beek EM, Roelandse-Koop EA, Vijzelaar R, et al. A multiplex assay to rapidly exclude HLA-DQ2.5 and HLA-DQ8 expression in patients at risk for celiac disease. Clin Chem Lab Med. 2013 Jun;51(6):1191-8. PMID: 23314539.

Included in a Prior Systematic Review on Topic - N=7

1. Aleanzi M, Demonte AM, Esper C, et al. Celiac disease: antibody recognition against native and selectively deamidated gliadin peptides. Clin Chem. 2001 Nov;47(11):2023-8. PMID: 11673371.

2. Arrowsmith JB, Gerstman BB, Fleischer DE, et al. Results from the American Society for Gastrointestinal Endoscopy/U.S. Food and Drug Administration collaborative study on complication rates and drug use during gastrointestinal endoscopy. Gastrointest Endosc. 1991 Jul-Aug;37(4):421-7. PMID: 1833259.

3. Lahdeaho ML, Maki M, Laurila K, et al. Small- bowel mucosal changes and antibody responses after low- and moderate-dose gluten challenge in celiac disease. BMC Gastroenterol. 2011;11:129. PMID: 22115041.

4. Li F, Gurudu SR, De Petris G, et al. Retention of the capsule endoscope: a single-center experience of 1000 capsule endoscopy procedures. Gastrointest Endosc. 2008 Jul;68(1):174-80. PMID: 18513723.

5. Mergener K, Enns R, JJ B, et al. Complications and problems with capsule endoscopy: results from two referral centers. Gastrointest Endosc 2003;57(AB171).

6. Roma E, Panayiotou J, Karantana H, et al. Changing pattern in the clinical presentation of pediatric celiac disease: a 30-year study. Digestion. 2009;80(3):185-91. PMID: 19776583.

7. Rostami K, Kerckhaert J, von Blomberg BM, et al. SAT and serology in adult coeliacs, seronegative coeliac disease seems a reality. Neth J Med. 1998 Jul;53(1):15-9. PMID: 9718937.

Does Not Assess Accuracy or Effectiveness - N=133

1. Adornetto G, Volpe G, De Stefano A, et al. An ELIME assay for the rapid diagnosis of coeliac disease. Anal Bioanal Chem. 2012 May;403(4):1191-4. PMID: 22258206.

2. Aguiar FM, Melo SB, Galvao LC, et al. Serological testing for celiac disease in women with endometriosis. A pilot study. Clin Exp Obstet Gynecol. 2009;36(1):23-5. PMID: 19400413.

3. Ahmed OI, Qasem SA, Abdulsattar JA, et al. Esophageal Eosinophilia in Pediatric Patients with Celiac Disease; Is it a Causal or an Incidental Association? J Pediatr Gastroenterol Nutr. 2014 Nov 25PMID: 25438025.

4. Ansaldi N, Palmas T, Corrias A, et al. Autoimmune thyroid disease and celiac disease in children. J Pediatr Gastroenterol Nutr. 2003 Jul;37(1):63-6. PMID: 12827007.

5. Ashabani A, Errabtea H, Shapan A, et al. Serologic markers of untreated celiac disease in Libyan children: antigliadin, antitransglutaminase, antiendomysial, and anticalreticulin antibodies. J Pediatr Gastroenterol Nutr. 2001 Sep;33(3):276-82. PMID: 11593122.

6. Balihar K, Kozeluhova J, Hejda V, et al. Diagnosing celiac disease in patients with a history of lymphoma: factors that matter. Wien Klin Wochenschr. 2013 Nov;125(21-22):696-703. PMID: 24149983.

7. Bao F, Yu L, Babu S, et al. One third of HLA DQ2 homozygous patients with type 1 diabetes express celiac disease-associated transglutaminase autoantibodies. J Autoimmun. 1999 Aug;13(1):143-8. PMID: 10441179.

8. Bardella MT, Elli L, De Matteis S, et al. Autoimmune disorders in patients affected by celiac sprue and inflammatory bowel disease. Ann Med. 2009;41(2):139-43. PMID: 18777226.

9. Bardella MT, Velio P, Cesana BM, et al. Coeliac disease: a histological follow-up study. Histopathology. 2007 Mar;50(4):465-71. PMID: WOS:000244518600008.

10. Blackwell PJ, Hill PG, Holmes GK. Autoantibodies to human tissue transglutaminase: superior predictors of coeliac disease. Scand J Gastroenterol. 2002 Nov;37(11):1282-5. PMID: 12465726.

11. Book L, Hart A, Black J, et al. Prevalence and clinical characteristics of celiac disease in Downs syndrome in a US study. Am J Med Genet. 2001 Jan 1;98(1):70-4. PMID: 11426458.

12. Borhani Haghighi A, Ansari N, Mokhtari M, et al. Multiple sclerosis and gluten sensitivity. Clin Neurol Neurosurg. 2007 Oct;109(8):651-3. PMID: 17537569.

13. Bottaro G, Cataldo F, Rotolo N, et al. The clinical pattern of subclinical/silent celiac disease: an analysis on 1026 consecutive cases. Am J Gastroenterol. 1999 Mar;94(3):691-6. PMID: 10086653.

14. Brett PM, Yiannakou JY, Morris MA, et al. A pedigree-based linkage study of coeliac disease: failure to replicate previous positive findings. Ann Hum Genet. 1998 Jan;62(Pt 1):25-32. PMID: 9659975.

15. Cabanne A, Vazquez H, Argonz J, et al. Clinical utility of counting intraepithelial lymphocytes in celiac disease intestinal mucosa. Acta Gastroenterol Latinoam. 2007 Mar;37(1):20-8. PMID: 17486742.

16. Caprai S, Vajro P, Ventura A, et al. Autoimmune liver disease associated with celiac disease in childhood: a multicenter study. Clin Gastroenterol Hepatol. 2008 Jul;6(7):803-6. PMID: 18258488.

17. Chumpitazi BP, Mysore K, Tsai CM, et al. Interprovider variation of celiac disease testing in childhood chronic abdominal pain. BMC Gastroenterol. 2013;13:150. PMID: 24124697.

18. Collin P, Mustalahti K, Kyronpalo S, et al. Should we screen reflux oesophagitis patients for coeliac disease? Eur J Gastroenterol Hepatol. 2004 Sep;16(9):917-20. PMID: 15316418.

19. Collin P, Rondonotti E, Lundin KE, et al. Video capsule endoscopy in celiac disease: current clinical practice. J Dig Dis. 2012 Feb;13(2):94-9. PMID: 22257477.

20. Corazza GR, Andreani ML, Biagi F, et al. The smaller size of the 'coeliac iceberg' in adults. Scand J Gastroenterol. 1997 Sep;32(9):917-9. PMID: 9299671.

21. Corazza GR, Di Sario A, Sacco G, et al. Subclinical coeliac disease: an anthropometric assessment. J Intern Med. 1994 Aug;236(2):183-7. PMID: 8046318.

22. da Rosa Utiyama SR, da Silva Kotze LM, Nisihara RM, et al. Spectrum of autoantibodies in celiac patients and relatives. Dig Dis Sci. 2001 Dec;46(12):2624-30. PMID: 11768251.

23. da Silva Kotze LM, Nisihara RM, da Rosa Utiyama SR, et al. Thyroid disorders in Brazilian patients with celiac disease. J Clin Gastroenterol. 2006 Jan;40(1):33-6. PMID: 16340631.

24. D'Archivio M, Silano M, Fagnani C, et al. Clinical evolution of celiac disease in Italy 1982-2002. J Clin Gastroenterol. 2004 Nov-Dec;38(10):877-9. PMID: 15492604.

25. Deja G, Myrda A, Jarosz-Chobot P, et al. The assessment of autoimmunological status and prevalence of different forms of celiac disease among children with type 1 diabetes mellitus and celiac disease. Mediators Inflamm. 2008;2008:285989. PMID: 18437226.

26. Diaconu G, Burlea M, Grigore I, et al. Celiac disease with neurologic manifestations in children. Rev Med Chir Soc Med Nat Iasi. 2013 Jan-Mar;117(1):88-94. PMID: 24505898.

27. Dixit R, Lebwohl B, Ludvigsson JF, et al. Celiac Disease Is Diagnosed Less Frequently in Young Adult Males. Dig Dis Sci. 2014 Jan 21PMID: 24445731.

28. Durante-Mangoni E, Iardino P, Resse M, et al. Silent celiac disease in chronic hepatitis C: impact of interferon treatment on the disease onset and clinical outcome. J Clin Gastroenterol. 2004 Nov-Dec;38(10):901-5. PMID: 15492610.

29. Ehsani-Ardakani MJ, Fallahian M, Rostami K, et al. Celiac disease and dysfunctional uterine bleeding; the efficiency of gluten free diet. Bratisl Lek Listy. 2014;115(1):19-21. PMID: 24471897.

30. Esteve M, Rosinach M, Fernandez-Banares F, et al. Spectrum of gluten-sensitive enteropathy in first-degree relatives of patients with coeliac disease: clinical relevance of lymphocytic enteritis. Gut. 2006 Dec;55(12):1739-45. PMID: 16709658.

31. Fasano A, Berti I, Gerarduzzi T, et al. Prevalence of celiac disease in at-risk and not-at-risk groups in the United States: a large multicenter study. Arch Intern Med. 2003 Feb 10;163(3):286-92. PMID: 12578508.

32. Ford AC, Ching E, Moayyedi P. Meta-analysis: yield of diagnostic tests for coeliac disease in dyspepsia. Alimentary Pharmacology & Therapeutics. 2009 Jul;30(1):28-36. PMID: WOS:000267069100003.

33. Freeman HJ. Biopsy-defined adult celiac disease in Asian-Canadians. Can J Gastroenterol. 2003 Jul;17(7):433-6. PMID: 12915916.

34. Gabrielli M, Cremonini F, Fiore G, et al. Association between migraine and Celiac disease: results from a preliminary case-control and therapeutic study. Am J Gastroenterol. 2003 Mar;98(3):625-9. PMID: 12650798.

35. Galvan JA, Lemos G, Fernandez de Cossio ME, et al. Silent celiac disease in a cohort of healthy adults. Autoimmunity. 2009;42(8):705-8. PMID: 19886741.

36. Garcia-Leiva JM, Carrasco JL, Slim M, et al. Celiac symptoms in patients with fibromyalgia: a cross-sectional study. Rheumatol Int. 2014 Aug 15PMID: 25119831.

37. Garnier-Lengline H, Brousse N, Candon S, et al. Have serological tests changed the face of childhood coeliac disease? A retrospective cohort study. BMJ Open. 2012;2(6)PMID: 23180388.

38. Gatti S, Caporelli N, Galeazzi T, et al. Oats in the diet of children with celiac disease: preliminary results of a double-blind, randomized, placebo-controlled multicenter Italian study. Nutrients. 2013 Nov;5(11):4653-64. PMID: 24264227.

39. Germenis AE, Yiannaki EE, Zachou K, et al. Prevalence and clinical significance of immunoglobulin A antibodies against tissue transglutaminase in patients with diverse chronic liver diseases. Clin Diagn Lab Immunol. 2005 Aug;12(8):941-8. PMID: 16085912.

40. Ghawil M, Miotti V, Tonutti E, et al. HLA-DQ types of celiac disease in Libyan children with type 1 diabetes mellitus. Eur J Gastroenterol Hepatol. 2012 Jan;24(1):59-63. PMID: 22002004.

41. Ghazzawi Y, Tapia AR, Murray JA, et al. Mucosal Healing in Children With Treated Celiac Disease. J Pediatr Gastroenterol Nutr. 2014 Mar 31PMID: 24691402.

42. Giangreco E, D'Agate C, Barbera C, et al. Prevalence of celiac disease in adult patients with refractory functional dyspepsia: value of routine duodenal biopsy. World J Gastroenterol. 2008 Dec 7;14(45):6948-53. PMID: 19058330.

43. Grainge MJ, West J, Card TR, et al. Causes of death in people with celiac disease spanning the pre- and post-serology era: a population-based cohort study from Derby, UK. Am J Gastroenterol. 2011 May;106(5):933-9. PMID: 21245833.

44. Granot E, Korman SM, Sallon S, et al. "Early" vs. "late" diagnosis of celiac disease in two ethnic groups living in the same geographic area. Isr J Med Sci. 1994 Apr;30(4):271-5. PMID: 8175328.

45. Green PH, Yang J, Cheng J, et al. An association between microscopic colitis and celiac disease. Clin Gastroenterol Hepatol. 2009 Nov;7(11):1210-6. PMID: 19631283.

46. Gulen H, Kasirga E, Yildirim SA, et al. Diagnostic yield of upper gastrointestinal endoscopy in the evaluation of iron deficiency anemia in older children and adolescents. Pediatr Hematol Oncol. 2011 Nov;28(8):694-701. PMID: 21728721.

47. Habior A, Lewartowska A, Orlowska J, et al. Association of coeliac disease with primary biliary cirrhosis in Poland. Eur J Gastroenterol Hepatol. 2003 Feb;15(2):159-64. PMID: 12560760.

48. Hansson T, Dahlbom I, Tuvemo T, et al. Silent coeliac disease is over-represented in children with type 1 diabetes and their siblings. Acta Paediatr. 2014 Oct 5PMID: 25283799.

49. Hariz MB, Laadhar L, Kallel-Sellami M, et al. Celiac disease in Tunisian children: a second screening study using a "new generation" rapid test. Immunol Invest. 2013;42(4):356-68. PMID: 23883201.

50. Harmon GS, Lebeck LK, Weidner N. Gluten-dependent enteropathy and atypical human leukocyte antigen alleles. Hum Pathol. 2011 Aug;42(8):1112-6. PMID: 21292306.

51. Hauser W, Gold J, Stein J, et al. Health-related quality of life in adult coeliac disease in Germany: results of a national survey. Eur J Gastroenterol Hepatol. 2006 Jul;18(7):747-54. PMID: 16772832.

52. Hill I, Fasano A, Schwartz R, et al. The prevalence of celiac disease in at-risk groups of children in the United States. J Pediatr. 2000 Jan;136(1):86-90. PMID: 10636980.

53. Holding S, Wilson F, Spradbery D. Clinical evaluation of the BioPlex 2200 Celiac IgA and IgG Kits - A novel multiplex screen incorporating an integral check for IgA deficiency. J Immunol Methods. 2014 Mar;405:29-34. PMID: 24424297.

54. Huang Y, Don-Wauchope AC, Grey VL, et al. Improving serological test ordering patterns for the diagnosis of celiac disease through clinical laboratory audit of practice. Clin Biochem. 2012 Apr;45(6):455-9. PMID: 22285379.

55. Isikay S, Kocamaz H. Prevalence of celiac disease in children with idiopathic epilepsy in southeast Turkey. Pediatr Neurol. 2014 May;50(5):479-81. PMID: 24656466.

56. Jansen MA, Tromp, II, Kiefte-de Jong JC, et al. Infant feeding and anti-tissue transglutaminase antibody concentrations in the Generation R Study. Am J Clin Nutr. 2014 Oct;100(4):1095-101. PMID: 25240074.

57. Johnston SD, Watson RG, McMillan SA, et al. Coeliac disease detected by screening is not silent--simply unrecognized. QJM. 1998 Dec;91(12):853-60. PMID: 10024951.

58. Jones S, D'Souza C, Haboubi NY. Patterns of clinical presentation of adult coeliac disease in a rural setting. Nutr J. 2006;5:24. PMID: 16972991.

59. Kaistha A, Castells S. Celiac disease in African American children with type 1 diabetes mellitus in inner city Brooklyn. Pediatr Endocrinol Rev. 2008 Aug;5 Suppl 4:994-8. PMID: 18806716.

60. Kalhan S, Joseph P, Sharma S, et al. Comparative study of histopathological Marsh grading with clinical and serological parameters in celiac iceberg of north India. Indian J Pathol Microbiol. 2011 Apr-Jun;54(2):279-83. PMID: 21623074.

61. Karavanaki K, Kakleas K, Paschali E, et al. Screening for associated autoimmunity in children and adolescents with type 1 diabetes mellitus (T1DM). Horm Res. 2009;71(4):201-6. PMID: 19258711.

62. Kautto E, Ryden PJ, Ivarsson A, et al. What happens to food choices when a gluten-free diet is required? A prospective longitudinal population-based study among Swedish adolescent with coeliac disease and their peers. J Nutr Sci. 2014;3:e2. PMID: 25191610.

63. Khashan AS, Henriksen TB, Mortensen PB, et al. The impact of maternal celiac disease on birthweight and preterm birth: a Danish population-based cohort study. Hum Reprod. 2010 Feb;25(2):528-34. PMID: 19939833.

64. Kinos S, Kurppa K, Ukkola A, et al. Burden of illness in screen-detected children with celiac disease and their families. J Pediatr Gastroenterol Nutr. 2012 Oct;55(4):412-6. PMID: 22614110.

65. Kochhar R, Dutta U, Miglani A, et al. Celiac disease suspected at endoscopy in patients with chronic liver disease. Indian J Gastroenterol. 2011 Jul;30(4):166-9. PMID: 21847607.

66. Koskinen O, Lindfors K, Collin P, et al. Intestinal transglutaminase 2 specific antibody deposits in non-responsive coeliac disease. Dig Liver Dis. 2010 Oct;42(10):692-7. PMID: 20409763.

67. Kotze LM, Utiyama SR, Nisihara RM, et al. Antiendomysium antibodies in Brazilian patients with celiac disease and their first-degree relatives. Arq Gastroenterol. 2001 Apr-Jun;38(2):94-103. PMID: 11793949.

68. Lakos G, Norman GL, Mahler M, et al. Analytical and clinical comparison of two fully automated immunoassay systems for the diagnosis of celiac disease. J Immunol Res. 2014;2014:371263. PMID: 24741592.

69. Larsson K, Carlsson A, Cederwall E, et al. Annual screening detects celiac disease in children with type 1 diabetes. Pediatr Diabetes. 2008 Aug;9(4 Pt 2):354-9. PMID: 18774995.

70. Levine A, Domanov S, Sukhotnik I, et al. Celiac-associated peptic disease at upper endoscopy: how common is it? Scand J Gastroenterol. 2009;44(12):1424-8. PMID: 19883278.

71. Lindh E, Ljunghall S, Larsson K, et al. Screening for antibodies against gliadin in patients with osteoporosis. J Intern Med. 1992 Apr;231(4):403-6. PMID: 1588266.

72. Lionetti E, Castellaneta S, Francavilla R, et al. Introduction of gluten, HLA status, and the risk of celiac disease in children. N Engl J Med. 2014 Oct 2;371(14):1295-303. PMID: 25271602.

73. Lionetti E, Francavilla R, Maiuri L, et al. Headache in pediatric patients with celiac disease and its prevalence as a diagnostic clue. J Pediatr Gastroenterol Nutr. 2009 Aug;49(2):202-7. PMID: 19543115.

74. Luostarinen L, Dastidar P, Collin P, et al. Association between coeliac disease, epilepsy and brain atrophy. Eur Neurol. 2001;46(4):187-91. PMID: 11721124.

75. Mandal AK, Mehdi I, Munshi SK, et al. Value of routine duodenal biopsy in diagnosing coeliac disease in patients with iron deficiency anaemia. Postgrad Med J. 2004 Aug;80(946):475-7. PMID: 15299158.

76. Mankai A, Ben Hamouda H, Amri F, et al. Screening by anti-endomysium antibodies for celiac disease in Tunisian children with type 1 diabetes mellitus. Gastroenterol Clin Biol. 2007 May;31(5):462-6. PMID: 17541335.

77. Mankai A, Chadli-Chaieb M, Saad F, et al. Screening for celiac disease in Tunisian patients with Graves' disease using anti-endomysium and anti-tissue transglutaminase antibodies. Gastroenterol Clin Biol. 2006 Aug-Sep;30(8-9):961-4. PMID: 17075442.

78. Matysiak-Budnik T, Malamut G, de Serre NP, et al. Long-term follow-up of 61 coeliac patients diagnosed in childhood: evolution toward latency is possible on a normal diet. Gut. 2007 Oct;56(10):1379-86. PMID: 17303598.

79. Metzger MH, Heier M, Maki M, et al. Mortality excess in individuals with elevated IgA anti-transglutaminase antibodies: the KORA/MONICA Augsburg cohort study 1989-1998. Eur J Epidemiol. 2006;21(5):359-65. PMID: 16649072.

80. Midhagen G, Aberg AK, Olcen P, et al. Antibody levels in adult patients with coeliac disease during gluten-free diet: a rapid initial decrease of clinical importance. J Intern Med. 2004 Dec;256(6):519-24. PMID: 15554953.

81. Myleus A, Petersen S, Carlsson A, et al. Health-related quality of life is not impaired in children with undetected as well as diagnosed celiac disease: a large population based cross-sectional study. BMC Public Health. 2014;14:425. PMID: 24884747.

82. Nakachi K, Swift G, Wilmot D, et al. Antibodies to tissue transglutaminase: comparison of ELISA and immunoprecipitation assay in the presence and in the absence of calcium ions. Clin Chim Acta. 2001 Feb;304(1-2):75-84. PMID: 11165201.

83. Nenna R, Tiberti C, Petrarca L, et al. Anti-transglutaminase immunoreactivity and histological lesions of the duodenum in coeliac patients. International Immunology. 2013 Jun;25(6):389-94. PMID: WOS:000319467000007.

84. Neves M, Gonzalez-Garcia MB, Delerue-Matos C, et al. Multiplexed electrochemical immunosensor for detection of celiac disease serological markers. Sensors and Actuators B-Chemical. 2013 Oct;187:33-9. PMID: WOS:000324298300007.

85. Norouzinia M, Rostami K, Amini M, et al. Coeliac disease; Prevalence and Outcome in Pregnancy. Healthmed. 2011;5(6):1537-41. PMID: WOS:000298663500024.

86. Nwosu BU, Snook RI, Maranda L. The relationship between adiposity and stature in prepubertal children with celiac disease. J Pediatr Endocrinol Metab. 2013;26(9-10):819-24. PMID: 23729610.

87. Ozgenc F, Aksu G, Aydogdu S, et al. Association between anti-endomysial antibody and total intestinal villous atrophy in children with coeliac disease. J Postgrad Med. 2003 Jan-Mar;49(1):21-4; discussion 4. PMID: 12865566.

88. Palabykoglu M, Botoman VA, Coban S, et al. A tale of two cities: typical celiac sprue presenting symptoms are significantly more common in Turkish than in US Patients. J Clin Gastroenterol. 2008 Jan;42(1):62-5. PMID: 18097292.

89. Picarelli A, Sabbatella L, Di TM, et al. Antiendomysial antibody detection in fecal supernatants: in vivo proof that small bowel mucosa is the site of antiendomysial antibody production. Am J Gastroenterol. 2002 Jan;97(1):95-8. PMID: 11808976.

90. Pierucci A, Fofi C, Bartoli B, et al. Antiendomysial antibodies in Berger's disease. Am J Kidney Dis. 2002 Jun;39(6):1176-82. PMID: 12046028.

91. Pittschieler K, Gentili L, Niederhofer H. Onset of coeliac disease: a prospective longitudinal study. Acta Paediatr. 2003 Oct;92(10):1149-52. PMID: 14632329.

92. Poddar U, Thapa BR, Singh K. Clinical features of celiac disease in Indian children: are they different from the West? J Pediatr Gastroenterol Nutr. 2006 Sep;43(3):313-7. PMID: 16954952.

93. Prasad KK, Thapa BR, Nain CK, et al. Brush border enzyme activities in relation to histological lesion in pediatric celiac disease. J Gastroenterol Hepatol. 2008 Aug;23(8 Pt 2):e348-52. PMID: 18070009.

94. Rajani S, Huynh HQ, Turner J. The changing frequency of celiac disease diagnosed at the Stollery Children's Hospital. Can J Gastroenterol. 2010 Feb;24(2):109-12. PMID: 20151069.

95. Rami B, Sumnik Z, Schober E, et al. Screening detected celiac disease in children with type 1 diabetes mellitus: effect on the clinical course (a case control study). J Pediatr Gastroenterol Nutr. 2005 Sep;41(3):317-21. PMID: 16131986.

96. Rawashdeh MO, Khalil B, Raweily E. Celiac disease in Arabs. J Pediatr Gastroenterol Nutr. 1996 Nov;23(4):415-8. PMID: 8956178.

97. Respondek W, Tomasiuk R, Jarosz M, et al. Is it reasonable to perform serological tests for celiac disease in patients with irritable bowel syndrome? Przeglad Gastroenterologiczny. 2013;8(3):184-90. PMID: WOS:000321721100007.

98. Rostami K, Mulder CJ, Stapel S, et al. Autoantibodies and histogenesis of celiac disease. Rom J Gastroenterol. 2003 Jun;12(2):101-6. PMID: 12853995.

99. Rostami-Nejad M, Romanos J, Rostami K, et al. Allele and haplotype frequencies for HLA-DQ in Iranian celiac disease patients. World J Gastroenterol. 2014 May 28;20(20):6302-8. PMID: 24876751.

100. Rostami-Nejad M, Villanacci V, Hogg-Kollars S, et al. Endoscopic and histological pitfalls in the diagnosis of celiac disease: A multicentre study assessing the current practice. Rev Esp Enferm Dig. 2013 Jul;105(6):326-33. PMID: 24090014.

101. Rubio-Tapia A, Kyle RA, Kaplan EL, et al. Increased prevalence and mortality in undiagnosed celiac disease. Gastroenterology. 2009 Jul;137(1):88-93. PMID: 19362553.

102. Rubio-Tapia A, Rahim MW, See JA, et al. Mucosal recovery and mortality in adults with celiac disease after treatment with a gluten-free diet. Am J Gastroenterol. 2010 Jun;105(6):1412-20. PMID: 20145607.

103. Rubio-Tapia A, Van Dyke CT, Lahr BD, et al. Predictors of family risk for celiac disease: a population-based study. Clin Gastroenterol Hepatol. 2008 Sep;6(9):983-7. PMID: 18585974.

104. Saadah OI, Al-Agha AE, Al Nahdi HM, et al. Prevalence of celiac disease in children with type 1 diabetes mellitus screened by anti-tissue transglutaminase antibody from Western Saudi Arabia. Saudi Med J. 2012 May;33(5):541-6. PMID: 22588816.

105. Salmi TT, Hervonen K, Laurila K, et al. Small Bowel Transglutaminase 2-specific IgA Deposits in Dermatitis Herpetiformis. Acta Derm Venereol. 2013 Dec 17PMID: 24352382.

106. Samasca G, Iancu M, Butnariu A, et al. Controversies in the laboratory diagnosis of celiac disease in children; new haplotypes discovered. Roum Arch Microbiol Immunol. 2010 Jul-Sep;69(3):119-24. PMID: 21434588.

107. Sategna-Guidetti C, Bruno M, Mazza E, et al. Autoimmune thyroid diseases and coeliac disease. Eur J Gastroenterol Hepatol. 1998 Nov;10(11):927-31. PMID: 9872614.

108. Savas N, Akbulut S, Saritas U, et al. Correlation of clinical and histopathological with endoscopic findings of celiac disease in the Turkish population. Dig Dis Sci. 2007 May;52(5):1299-303. PMID: 17356915.

109. Schober E, Bittmann B, Granditsch G, et al. Screening by anti-endomysium antibody for celiac disease in diabetic children and adolescents in Austria. J Pediatr Gastroenterol Nutr. 2000 Apr;30(4):391-6. PMID: 10776949.

110. Sharma A, Poddar U, Yachha SK. Time to recognize atypical celiac disease in Indian children. Indian J Gastroenterol. 2007 Nov-Dec;26(6):269-73. PMID: 18431009.

111. Sharma M, Singh P, Agnihotri A, et al. Celiac disease: a disease with varied manifestations in adults and adolescents. J Dig Dis. 2013 Oct;14(10):518-25. PMID: 23906112.

112. Shmidt E, Smyrk TC, Faubion WA, et al. Duodenal intraepithelial lymphocytosis with normal villous architecture in pediatric patients: Mayo Clinic experience, 2000-2009. J Pediatr Gastroenterol Nutr. 2013 Jan;56(1):51-5. PMID: 22785416.

113. Singh P, Kurray L, Agnihotri A, et al. Titers of Anti-tissue Transglutaminase Antibody Correlate Well With Severity of Villous Abnormalities in Celiac Disease. J Clin Gastroenterol. 2014 Feb 27PMID: 24583754.

114. Singh R, Nind G, Tucker G, et al. Narrow-band imaging in the evaluation of villous morphology: a feasibility study assessing a simplified classification and observer agreement. Endoscopy. 2010 Nov;42(11):889-94. PMID: 21072704.

115. Sperandeo MP, Tosco A, Izzo V, et al. Potential celiac patients: a model of celiac disease pathogenesis. PLoS One. 2011;6(7):e21281. PMID: 21760890.

116. Stagi S, Giani T, Simonini G, et al. Thyroid function, autoimmune thyroiditis and coeliac disease in juvenile idiopathic arthritis. Rheumatology (Oxford). 2005 Apr;44(4):517-20. PMID: 15695302.

117. Stenberg R, Kaukinen K, Bengtsson M, et al. Early developing celiac disease in children with cerebral palsy. J Pediatr Gastroenterol Nutr. 2011 Dec;53(6):674-8. PMID: 21697743.

118. Stenson WF, Newberry R, Lorenz R, et al. Increased prevalence of celiac disease and need for routine screening among patients with osteoporosis. Arch Intern Med. 2005 Feb 28;165(4):393-9. PMID: 15738367.

119. Szaflarska-Poplawska A. Patients with serological markers of coeliac disease but without features of atrophy concerning villi of the small bowel mucosa - own observations. Przeglad Gastroenterologiczny. 2009;4(3):152-8. PMID: WOS:000269188000008.

120. Szaflarska-Poplawska A, Parzecka M, Muller L, et al. Screening for celiac disease in Poland. Med Sci Monit. 2009 Mar;15(3):PH7-11. PMID: 19247256.

121. Tanpowpong P, Broder-Fingert S, Katz AJ, et al. Age-related patterns in clinical presentations and gluten-related issues among children and adolescents with celiac disease. Clin Transl Gastroenterol. 2012;3:e9. PMID: 23238134.

122. Taubman B, Mamula P, Sherry DD. Prevalence of asymptomatic celiac disease in children with fibromyalgia: a pilot study. Pediatr Rheumatol Online J. 2011;9:11. PMID: 21668956.

123. Tighe MR, Hall MA, Cardi E, et al. Associations between alleles of the major histocompatibility complex-encoded ABC transporter gene TAP2, HLA class II alleles, and celiac disease susceptibility. Hum Immunol. 1994 Jan;39(1):9-16. PMID: 8181966.

124. Tursi A, Brandimarte G. The symptomatic and histologic response to a gluten-free diet in patients with borderline enteropathy. J Clin Gastroenterol. 2003 Jan;36(1):13-7. PMID: 12488700.

125. Uibo O, Teesalu K, Metskula K, et al. Screening for celiac disease in Down's syndrome patients revealed cases of subtotal villous atrophy without typical for celiac disease HLA-DQ and tissue transglutaminase antibodies. World J Gastroenterol. 2006 Mar 7;12(9):1430-4. PMID: 16552815.

126. Unsworth DJ, Lock RJ, Harvey RF. Improving the diagnosis of coeliac disease in anaemic women. Br J Haematol. 2000 Dec;111(3):898-901. PMID: 11122153.

127. Velluzzi F, Caradonna A, Boy MF, et al. Thyroid and celiac disease: clinical, serological, and echographic study. Am J Gastroenterol. 1998 Jun;93(6):976-9. PMID: 9647032.

128. Viljamaa M, Collin P, Huhtala H, et al. Is coeliac disease screening in risk groups justified? A fourteen-year follow-up with special focus on compliance and quality of life. Aliment Pharmacol Ther. 2005 Aug 15;22(4):317-24. PMID: 16097998.

129. Vogelsang H, Panzer S, Mayr WR, et al. Distribution of HLA class I alleles differs in celiac disease patients according to age of onset. Dig Dis Sci. 2003 Mar;48(3):611-4. PMID: 12757179.

130. Watanabe C, Komoto S, Hokari R, et al. Prevalence of serum celiac antibody in patients with IBD in Japan. J Gastroenterol. 2013 Jun 12PMID: 23754511.

131. Westerhof J, Weersma RK, Koornstra JJ. Risk factors for incomplete small-bowel capsule endoscopy. Gastrointestinal Endoscopy. 2009 Jan;69(1):74-80. PMID: WOS:000262261900013.

132. Wierdsma NJ, van Bokhorst-de van der Schueren MA, Berkenpas M, et al. Vitamin and mineral deficiencies are highly prevalent in newly diagnosed celiac disease patients. Nutrients. 2013 Oct;5(10):3975-92. PMID: 24084055.

133. Zwolinska-Wcislo M, Tomaszewska R, Rozpondek P, et al. Mucosal lesions of the gastric mucosa in adult patients with coeliac disease. Przeglad Gastroenterologiczny. 2012;7(5):291-8. PMID: WOS:000315003200007.

Index Test Not Compared to Biopsy - N=62

1. Barbero EM, McNally SL, Donohue MC, et al. Barriers impeding serologic screening for celiac disease in clinically high-prevalence populations. BMC Gastroenterol. 2014;14:42. PMID: 24592899.

2. Barret M, Malamut G, Rahmi G, et al. Diagnostic yield of capsule endoscopy in refractory celiac disease. Am J Gastroenterol. 2012 Oct;107(10):1546-53. PMID: 22964554.

3. Barshack I, Goldberg I, Chowers Y, et al. Immunohistochemical analysis of candidate gene product expression in the duodenal epithelium of children with coeliac sprue. Journal of Clinical Pathology. 2001 Sep;54(9):684-8. PMID: WOS:000170833400007.

4. Burgin-Wolff A, Dahlbom I, Hadziselimovic F, et al. Antibodies against human tissue transglutaminase and endomysium in diagnosing and monitoring coeliac disease. Scand J Gastroenterol. 2002 Jun;37(6):685-91. PMID: 12126247.

5. Canavan C, Logan RF, Khaw KT, et al. No difference in mortality in undetected coeliac disease compared with the general population: a UK cohort study. Aliment Pharmacol Ther. 2011 Oct;34(8):1012-9. PMID: 21848796.

6. Catassi C, Kryszak D, Bhatti B, et al. Natural history of celiac disease autoimmunity in a USA cohort followed since 1974. Ann Med. 2010 Oct;42(7):530-8. PMID: 20868314.

7. Chomeili B, Aminzadeh M, Hardani AK, et al. Prevalence of celiac disease in siblings of Iranian patients with celiac disease. Arq Gastroenterol. 2011 Apr-Jun;48(2):131-5. PMID: 21709955.

8. Conrad K, Roggenbuck D, Ittenson A, et al. A new dot immunoassay for simultaneous detection of celiac specific antibodies and IgA-deficiency. Clin Chem Lab Med. 2012 Feb;50(2):337-43. PMID: 22505544.

9. Cooper SJ, Lovatt TJ. Highs and lows of coeliac screening. Br J Biomed Sci. 2009;66(2):79-84. PMID: 19637648.

10. Dahlbom I, Olsson M, Forooz NK, et al. Immunoglobulin G (IgG) anti-tissue transglutaminase antibodies used as markers for IgA-deficient celiac disease patients. Clin Diagn Lab Immunol. 2005 Feb;12(2):254-8. PMID: 15699419.

11. Dickerson F, Stallings C, Origoni A, et al. Markers of gluten sensitivity and celiac disease in bipolar disorder. Bipolar Disord. 2011 Feb;13(1):52-8. PMID: 21320252.

12. Diniz-Santos DR, Brandao F, Adan L, et al. Bone mineralization in young patients with type 1 diabetes mellitus and screening-identified evidence of celiac disease. Dig Dis Sci. 2008 May;53(5):1240-5. PMID: 17939041.

13. Duerksen DR, Leslie WD. Positive celiac disease serology and reduced bone mineral density in adult women. Can J Gastroenterol. 2010 Feb;24(2):103-7. PMID: 20151068.

14. Duerksen DR, Leslie WD. Longitudinal evaluation of bone mineral density and body composition in patients with positive celiac serology. J Clin Densitom. 2011 Oct-Dec;14(4):478-83. PMID: 21852167.

15. Ertekin V, Selimoglu MA, Kardas F, et al. Prevalence of celiac disease in Turkish children. J Clin Gastroenterol. 2005 Sep;39(8):689-91. PMID: 16082278.

16. Fernandez-Cavada-Pollo MJ, Alcala-Pena MI, Vargas-Perez ML, et al. Celiac disease and HLA-DQ genotype: Diagnosis of different genetic risk profiles related to the age in Badajoz, southwestern Spain. Rev Esp Enferm Dig. 2013 Sep;105(8):469-76. PMID: 24274444.

17. Fisher AH, Lomasky SJ, Fisher MJ, et al. Celiac disease and the endocrinologist: a diagnostic opportunity. Endocr Pract. 2008 Apr;14(3):381-8. PMID: 18463048.

18. Foucher B, Johanet C, Jego-Desplat S, et al. Are immunoglobulin A anti-gliadin antibodies helpful in diagnosing coeliac disease in children younger than 2 years? J Pediatr Gastroenterol Nutr. 2012 Jan;54(1):110-2. PMID: 21857243.

19. Gabriel S, Mihaela I, Angela B, et al. Prevalence of IgA antitissue transglutaminase antibodies in children with type 1 diabetes mellitus. J Clin Lab Anal. 2011;25(3):156-61. PMID: 21567461.

20. Gheita TA, Fawzy SM, Nour El-Din AM, et al. Asymptomatic celiac sprue in juvenile rheumatic diseases children. Int J Rheum Dis. 2012 Apr;15(2):220-6. PMID: 22212536.

21. Godfrey JD, Brantner TL, Brinjikji W, et al. Morbidity and mortality among older individuals with undiagnosed celiac disease. Gastroenterology. 2010 Sep;139(3):763-9. PMID: 20685275.

22. Goldberg D, Kryszak D, Fasano A, et al. Screening for celiac disease in family members: Is follow-up testing necessary? Digestive Diseases and Sciences. 2007 Apr;52(4):1082-6. PMID: WOS:000245131900036.

23. Grodzinsky E, Jansson G, Skogh T, et al. Anti-endomysium and anti-gliadin antibodies as serological markers for coeliac disease in childhood: a clinical study to develop a practical routine. Acta Paediatr. 1995 Mar;84(3):294-8. PMID: 7780251.

24. Hussain S, Sabir MU, Afzal M, et al. Coeliac disease--clinical presentation and diagnosis by anti tissue transglutaminase antibodies titre in children. J Pak Med Assoc. 2014 Apr;64(4):437-41. PMID: 24864640.

25. Hussain S, Sabir MUD, Afzal M, et al. Coeliac disease - clinical presentation and diagnosis by anti tissue transglutaminase antibodies titre in children. Journal of the Pakistan Medical Association. 2014 Apr;64(4):437-41. PMID: WOS:000333319300017.

26. Kaspers S, Kordonouri O, Schober E, et al. Anthropometry, metabolic control, and thyroid autoimmunity in type 1 diabetes with celiac disease: A multicenter survey. J Pediatr. 2004 Dec;145(6):790-5. PMID: 15580203.

27. Katsinelos P, Tziomalos K, Fasoulas K, et al. Can capsule endoscopy be used as a diagnostic tool in the evaluation of nonbleeding indications in daily clinical practice? A prospective study. Med Princ Pract. 2011;20(4):362-7. PMID: 21576998.

28. Kontiainen S, Schlenzka A, Koskimies S, et al. Autoantibodies and autoimmune diseases in young diabetics. Diabetes Res. 1990 Apr;13(4):151-6. PMID: 2134205.

29. Laadhar L, Kallel-Sellami M, Zitouni M, et al. Is the rapid whole blood test useful for diagnosis and monitoring celiac disease in children? Tunis Med. 2011 Jan;89(1):16-7. PMID: 21267821.

30. Laass MW, Koch T, Losel A, et al. Longitudinal follow-up examination of antigliadin antibody positive children and adults. Eur J Gastroenterol Hepatol. 2006 May;18(5):503-6. PMID: 16607144.

31. Laserna-Mendieta EJ, Pineda-Tenor D, Timon-Zapata J, et al. A proposed reference change value for an IgA anti-tissue transglutaminase immunoassay to improve interpretation of serial results in celiac patients. Clin Chim Acta. 2013 Jun 5;421:12-6. PMID: 23470429.

32. Lavant EH, Agardh DJ, Nilsson A, et al. A new PCR-SSP method for HLA DR-DQ risk assessment for celiac disease. Clin Chim Acta. 2011 Apr 11;412(9-10):782-4. PMID: 21219892.

33. Leboff MS, Cobb H, Gao LY, et al. Celiac disease is not increased in women with hip fractures and low vitamin D levels. J Nutr Health Aging. 2013;17(6):562-5. PMID: 23732553.

34. Leffler DA, Dennis M, Hyett B, et al. Etiologies and predictors of diagnosis in nonresponsive celiac disease. Clin Gastroenterol Hepatol. 2007 Apr;5(4):445-50. PMID: 17382600.

35. Leja M, Kojalo U, Frickauss G, et al. Changing patterns of serological testing for celiac disease in Latvia. J Gastrointestin Liver Dis. 2011 Jun;20(2):121-6. PMID: 21725506.

36. Lionetti E, Castellaneta S, Pulvirenti A, et al. Prevalence and natural history of potential celiac disease in at-family-risk infants prospectively investigated from birth. J Pediatr. 2012 Nov;161(5):908-14. PMID: 22704250.

37. Liu E, Li M, Emery L, et al. Natural history of antibodies to deamidated gliadin peptides and transglutaminase in early childhood celiac disease. J Pediatr Gastroenterol Nutr. 2007 Sep;45(3):293-300. PMID: 17873740.

38. Lochman I, Martis P, Burlingame RW, et al. Multiplex assays to diagnose celiac disease. Ann N Y Acad Sci. 2007 Aug;1109:330-7. PMID: 17785322.

39. Lohi S, Maki M, Montonen J, et al. Malignancies in cases with screening-identified evidence of coeliac disease: a long-term population-based cohort study. Gut. 2009 May;58(5):643-7. PMID: 18852259.

40. Makins R, Blanshard C. Guidelines for capsule endoscopy: diagnoses will be missed. Aliment Pharmacol Ther. 2006 Jul 15;24(2):293-7. PMID: 16842455.

41. Margaritte-Jeannin P, Babron MC, Bourgey M, et al. HLA-DQ relative risks for coeliac disease in European populations: a study of the European Genetics Cluster on Coeliac Disease. Tissue Antigens. 2004 Jun;63(6):562-7. PMID: 15140032.

42. Megiorni F, Mora B, Bonamico M, et al. HLA-DQ and susceptibility to celiac disease: evidence for gender differences and parent-of-origin effects. Am J Gastroenterol. 2008 Apr;103(4):997-1003. PMID: 18177450.

43. Miller A, Paspaliaris W, Elliott PR, et al. Anti-transglutaminase antibodies and coeliac disease. Aust N Z J Med. 1999 Apr;29(2):239-42. PMID: 10342024.

44. Murray JA, McLachlan S, Adams PC, et al. Association between celiac disease and iron deficiency in Caucasians, but not non-Caucasians. Clin Gastroenterol Hepatol. 2013 Jul;11(7):808-14. PMID: 23416278.

45. Nagui N, El Nabarawy E, Mahgoub D, et al. Estimation of (IgA) anti-gliadin, antiendomysium and tissue transglutaminase in the serum of patients with psoriasis. Clin Exp Dermatol. 2011 Apr;36(3):302-4. PMID: 21418272.

46. Nass FR, Kotze LM, Nisihara RM, et al. Serological and clinical follow-up of relatives of celiac disease patients from southern Brazil. Digestion. 2011;83(1-2):89-95. PMID: 21042020.

47. Nass FR, Kotze LM, Nisihara RM, et al. Autoantibodies in relatives of celiac disease patients: a follow-up of 6-10 years. Arq Gastroenterol. 2012 Jul-Sep;49(3):199-203. PMID: 23011242.

48. Neves MM, Gonzalez-Garcia MB, Nouws HP, et al. An electrochemical deamidated gliadin antibody immunosensor for celiac disease clinical diagnosis. Analyst. 2013 Apr 7;138(7):1956-8. PMID: 23400113.

49. Prince HE. Evaluation of the INOVA diagnostics enzyme-linked immunosorbent assay kits for measuring serum immunoglobulin G (IgG) and IgA to deamidated gliadin peptides. Clinical and Vaccine Immunology. 2006 Jan;13(1):150-1. PMID: WOS:000235369300019.

50. Rubio-Tapia A, Ludvigsson JF, Brantner TL, et al. The prevalence of celiac disease in the United States. Am J Gastroenterol. 2012 Oct;107(10):1538-44; quiz 7, 45. PMID: 22850429.

51. Samasca G, Iancu M, Farcau D, et al. IgA anti-tissue transglutaminase antibodies, first line in the diagnosis of celiac disease. Clin Lab. 2011;57(9-10):695-701. PMID: 22029184.

52. Sinclair D, Saas M, Turk A, et al. Do we need to measure total serum IgA to exclude IgA deficiency in coeliac disease? Journal of Clinical Pathology. 2006 Jul;59(7):736-9. PMID: WOS:000238592900013.

53. Teresi S, Crapisi M, Vallejo MD, et al. Celiac disease seropositivity in Saharawi children: a follow-up and family study. J Pediatr Gastroenterol Nutr. 2010 May;50(5):506-9. PMID: 20639708.

54. Tiberti C, Panimolle F, Bonamico M, et al. IgA anti-transglutaminase autoantibodies at type 1 diabetes onset are less frequent in adult patients and are associated with a general celiac-specific lower immune response in comparison with nondiabetic celiac patients at diagnosis. Diabetes Care. 2012 Oct;35(10):2083-5. PMID: 22815294.

55. Van Weyenberg SJ, Smits F, Jacobs MA, et al. Video capsule endoscopy in patients with nonresponsive celiac disease. J Clin Gastroenterol. 2013 May-Jun;47(5):393-9. PMID: 23164686.

56. Vidales MC, Zubillaga P, Zubillaga I, et al. Allele and haplotype frequencies for HLA class II (DQA1 and DQB1) loci in patients with celiac disease from Spain. Hum Immunol. 2004 Apr;65(4):352-8. PMID: 15120190.

57. Villalta D, Alessio MG, Tampoia M, et al. Diagnostic accuracy of IgA anti-tissue transglutaminase antibody assays in celiac disease patients with selective IgA deficiency. Ann N Y Acad Sci. 2007 Aug;1109:212-20. PMID: 17785308.

58. Villalta D, Tonutti E, Prause C, et al. IgG antibodies against deamidated gliadin peptides for diagnosis of celiac disease in patients with IgA deficiency. Clin Chem. 2010 Mar;56(3):464-8. PMID: 20022984.

59. Vitoria JC, Arrieta A, Arranz C, et al. Antibodies to gliadin, endomysium, and tissue transglutaminase for the diagnosis of celiac disease. J Pediatr Gastroenterol Nutr. 1999 Nov;29(5):571-4. PMID: 10554125.

60. Vojvodic S, Ademovic-Sazdanic D. Hla II Class Antigens and Susceptibility to Coeliac Disease. Genetika-Belgrade. 2011;43(3):517-26. PMID: WOS:000299730100009.

61. West J, Logan RFA, Hill PG, et al. The iceberg of celiac disease: What is below the waterline? Clinical Gastroenterology and Hepatology. 2007 Jan;5(1):59-62. PMID: WOS:000243781300011.

62. Wu J, Xia B, von Blomberg BM, et al. Coeliac disease in China, a field waiting for exploration. Rev Esp Enferm Dig. 2010 Jul;102(8):472-7. PMID: 20670067.

Not All Subjects Underwent Both Index Test and Reference Standard - N=410

1. Abbass R, Hopkins M, Dufour DR, et al. Celiac disease in an urban VA population with iron deficiency: the case against routine duodenal biopsy. Dig Dis Sci. 2011 Jul;56(7):2037-41. PMID: 21222157.

2. Abd El Dayem SM, Ahmed Aly A, Abd El Gafar E, et al. Screening for coeliac disease among Egyptian children. Arch Med Sci. 2010 Apr 30;6(2):226-35. PMID: 22371752.

3. Aberg AK, Olcen P. Serologic screening for celiac disease in children: a comparison between established assays and tests with deamidated gliadin-derived peptides plus conjugates for both IgA and IgG antibodies. APMIS. 2009 Nov;117(11):808-13. PMID: 19845531.

4. Abrantes-Lemos CP, Nakhle MC, Damiao AO, et al. Performance of two commercial ELISAs for detecting IgA anti-human and anti-guinea pig tissue transglutaminase antibodies. Clin Lab. 2010;56(1-2):29-35. PMID: 20380357.

5. Abu-Zekry M, Kryszak D, Diab M, et al. Prevalence of celiac disease in Egyptian children disputes the east-west agriculture-dependent spread of the disease. J Pediatr Gastroenterol Nutr. 2008 Aug;47(2):136-40. PMID: 18664863.

6. Acerini CL, Ahmed ML, Ross KM, et al. Coeliac disease in children and adolescents with IDDM: clinical characteristics and response to gluten-free diet. Diabet Med. 1998 Jan;15(1):38-44. PMID: 9472862.

7. Agardh D, Nilsson A, Carlsson A, et al. Tissue transglutaminase autoantibodies and human leucocyte antigen in Down's syndrome patients with coeliac disease. Acta Paediatr. 2002;91(1):34-8. PMID: 11883815.

8. Agardh D, Nilsson A, Tuomi T, et al. Prediction of silent celiac disease at diagnosis of childhood type 1 diabetes by tissue transglutaminase autoantibodies and HLA. Pediatr Diabetes. 2001 Jun;2(2):58-65. PMID: 15016199.

9. Akbari MR, Mohammadkhani A, Fakheri H, et al. Screening of the adult population in Iran for coeliac disease: comparison of the tissue-transglutaminase antibody and anti-endomysial antibody tests. Eur J Gastroenterol Hepatol. 2006 Nov;18(11):1181-6. PMID: 17033439.

10. Akbulut S, Gur G, Topal F, et al. Coeliac Disease-Associated Antibodies in Psoriasis. Annals of Dermatology. 2013 Aug;25(3):298-303. PMID: WOS:000323687000005.

11. Aktay AN, Lee PC, Kumar V, et al. The prevalence and clinical characteristics of celiac disease in juvenile diabetes in Wisconsin. J Pediatr Gastroenterol Nutr. 2001 Oct;33(4):462-5. PMID: 11698764.

12. Alarida K, Harown J, Ahmaida A, et al. Coeliac disease in Libyan children: a screening study based on the rapid determination of anti-transglutaminase antibodies. Dig Liver Dis. 2011 Sep;43(9):688-91. PMID: 21310672.

13. Alencar ML, Ortiz-Agostinho CL, Nishitokukado L, et al. Prevalence of celiac disease among blood donors in Sao Paulo: the most populated city in Brazil. Clinics (Sao Paulo). 2012 Sep;67(9):1013-8. PMID: 23018296.

14. Alessio MG, Tonutti E, Brusca I, et al. Correlation between IgA tissue transglutaminase antibody ratio and histological finding in celiac disease. J Pediatr Gastroenterol Nutr. 2012 Jul;55(1):44-9. PMID: 22197946.

15. Al-Hussaini A, Sulaiman N, Al-Zahrani M, et al. High prevalence of celiac disease among Saudi children with type 1 diabetes: a prospective cross-sectional study. BMC Gastroenterol. 2012;12:180. PMID: 23259699.

16. Altintas E, Senli MS, Sezgin O. Prevalence of celiac disease among dyspeptic patients: a community-based case-control study. Turk J Gastroenterol. 2008 Jun;19(2):81-4. PMID: 19110661.

17. Al-Toma A, Goerres MS, Meijer JW, et al. Human leukocyte antigen-DQ2 homozygosity and the development of refractory celiac disease and enteropathy-associated T-cell lymphoma. Clin Gastroenterol Hepatol. 2006 Mar;4(3):315-9. PMID: 16527694.

18. Amarri S, Alvisi P, De Giorgio R, et al. Antibodies to deamidated gliadin peptides: an accurate predictor of coeliac disease in infancy. J Clin Immunol. 2013 Jul;33(5):1027-30. PMID: 23558824.

19. Anderson RP, Henry MJ, Taylor R, et al. A novel serogenetic approach determines the community prevalence of celiac disease and informs improved diagnostic pathways. BMC Med. 2013 Aug 28;11(1):188. PMID: 23981538.

20. Ashabani A, Abushofa U, Abusrewill S, et al. The prevalence of coeliac disease in Libyan children with type 1 diabetes mellitus. Diabetes Metab Res Rev. 2003 Jan-Feb;19(1):69-75. PMID: 12592646.

21. Aygun C, Uraz S, Damci T, et al. Celiac disease in an adult Turkish population with type 1 diabetes mellitus. Dig Dis Sci. 2005 Aug;50(8):1462-6. PMID: 16110836.

22. Bakhshipour A, Kaykhaei MA, Moulaei N, et al. Prevalence of coeliac disease in patients with non-alcoholic fatty liver disease. Arab J Gastroenterol. 2013 Sep;14(3):113-5. PMID: 24206739.

23. Bakker SF, Tushuizen ME, Stokvis-Brantsma WH, et al. Frequent delay of coeliac disease diagnosis in symptomatic patients with type 1 diabetes mellitus: clinical and genetic characteristics. Eur J Intern Med. 2013 Jul;24(5):456-60. PMID: 23414771.

24. Barbato M, Viola F, Miglietta MR, et al. Association between insulin dependent diabetes mellitus and coeliac disease. A study on 175 diabetes patients. Minerva Gastroenterol Dietol. 1998 Mar;44(1):1-5. PMID: 16495876.

25. Bardella MT, Elli L, Velio P, et al. Silent celiac disease is frequent in the siblings of newly diagnosed celiac patients. Digestion. 2007;75(4):182-7. PMID: 17848794.

26. Bardella MT, Vecchi M, Conte D, et al. Chronic unexplained hypertransaminasemia may be caused by occult celiac disease. Hepatology. 1999 Mar;29(3):654-7. PMID: 10051464.

27. Bashiri H, Keshavarz A, Madani H, et al. Celiac disease in type-I diabetes mellitus: coexisting phenomenon. J Res Med Sci. 2011 Mar;16 Suppl 1:S401-6. PMID: 22247725.

28. Basso D, Guariso G, Fasolo M, et al. A new indirect chemiluminescent immunoassay to measure anti-tissue transglutaminase antibodies. J Pediatr Gastroenterol Nutr. 2006 Nov;43(5):613-8. PMID: 17130737.

29. Bazzigaluppi E, Parma B, Tronconi GM, et al. IgA anti-actin antibodies in children with celiac disease: comparison of immunofluorescence with Elisa assay in predicting severe intestinal damage. Ital J Pediatr. 2010;36:25. PMID: 20298549.

30. Bdioui F, Sakly N, Hassine M, et al. Prevalence of celiac disease in Tunisian blood donors. Gastroenterol Clin Biol. 2006 Jan;30(1):33-6. PMID: 16514380.

31. Ben Hariz M, Kallel-Sellami M, Kallel L, et al. Prevalence of celiac disease in Tunisia: mass-screening study in schoolchildren. Eur J Gastroenterol Hepatol. 2007 Aug;19(8):687-94. PMID: 17625439.

32. Benkebil F, Combescure C, Anghel SI, et al. Diagnostic accuracy of a new point-of-care screening assay for celiac disease. World J Gastroenterol. 2013 Aug 21;19(31):5111-7. PMID: 23964145.

33. Benson BC, Mulder CJ, Laczek JT. Anti-gliadin antibodies identify celiac patients overlooked by tissue transglutaminase antibodies. Hawaii J Med Public Health. 2013 Sep;72(9 Suppl 4):14-7. PMID: 24052912.

34. Berti I, Trevisiol C, Tommasini A, et al. Usefulness of screening program for celiac disease in autoimmune thyroiditis. Dig Dis Sci. 2000 Feb;45(2):403-6. PMID: 10711459.

35. Betterle C, Lazzarotto F, Spadaccino AC, et al. Celiac disease in North Italian patients with autoimmune Addison's disease. Eur J Endocrinol. 2006 Feb;154(2):275-9. PMID: 16452541.

36. Bhadada SK, Kochhar R, Bhansali A, et al. Prevalence and clinical profile of celiac disease in type 1 diabetes mellitus in north India. J Gastroenterol Hepatol. 2011 Feb;26(2):378-81. PMID: 21261730.

37. Bhattacharya M, Lomash A, Sakhuja P, et al. Clinical and histopathological correlation of duodenal biopsy with IgA anti-tissue transglutaminase titers in children with celiac disease. Indian J Gastroenterol. 2014 Jul;33(4):350-4. PMID: 24859392.

38. Biagi F, Campanella J, Soriani A, et al. Prevalence of coeliac disease in Italian patients affected by Addison's disease. Scand J Gastroenterol. 2006 Mar;41(3):302-5. PMID: 16497617.

39. Bienvenu F, Besson Duvanel C, Seignovert C, et al. Evaluation of a point-of-care test based on deamidated gliadin peptides for celiac disease screening in a large pediatric population. Eur J Gastroenterol Hepatol. 2012 Dec;24(12):1418-23. PMID: 23032795.

40. Bilbao JR, Vitoria JC, Ortiz L, et al. Immunoglobulin G autoantibodies against tissuetransglutaminase. A sensitive, cost-effective assay for the screening of celiac disease. Autoimmunity. 2002 Jul;35(4):255-9. PMID: 12482193. 41. Bizzaro N, Villalta D, Tonutti E, et al. IgA and IgG tissue transglutaminase antibody prevalence and clinical significance in connective tissue diseases, inflammatory bowel disease, and primary biliary cirrhosis. Dig Dis Sci. 2003 Dec;48(12):2360-5. PMID: 14714625.

42. Bjorck S, Brundin C, Lorinc E, et al. Screening detects a high proportion of celiac disease in young HLA-genotyped children. J Pediatr Gastroenterol Nutr. 2010 Jan;50(1):49-53. PMID: 19915493.

43. Bonamico M, Bottaro G, Pasquino AM, et al. Celiac disease and Turner syndrome. Journal of Pediatric Gastroenterology and Nutrition. 1998 May;26(5):496-9. PMID: WOS:000073358400002.

44. Bonamico M, Ferri M, Mariani P, et al. Serologic and genetic markers of celiac disease: a sequential study in the screening of first degree relatives. J Pediatr Gastroenterol Nutr. 2006 Feb;42(2):150-4. PMID: 16456406.

45. Bonamico M, Mariani P, Danesi HM, et al. Prevalence and clinical picture of celiac disease in italian down syndrome patients: a multicenter study. J Pediatr Gastroenterol Nutr. 2001 Aug;33(2):139-43. PMID: 11568513.

46. Bonamico M, Pasquino AM, Mariani P, et al. Prevalence and clinical picture of celiac disease in Turner syndrome. J Clin Endocrinol Metab. 2002 Dec;87(12):5495-8. PMID: 12466343.

47. Bonamico M, RasoreQuartino A, Mariani P, et al. Down syndrome and coeliac disease: Usefulness of antigliadin and antiendomysium antibodies. Acta Paediatrica. 1996 Dec;85(12):1503-5. PMID: WOS:A1996VX97800023.

48. Book L, Zone JJ, Neuhausen SL. Prevalence of celiac disease among relatives of sib pairs with celiac disease in U.S. families. Am J Gastroenterol. 2003 Feb;98(2):377-81. PMID: 12591058.

49. Bottaro G, Failla P, Rotolo N, et al. Changes in coeliac disease behaviour over the years. Acta Paediatr. 1993 Jun-Jul;82(6-7):566-8. PMID: 8338991.

50. Boudraa G, Hachelaf W, Benbouabdellah M, et al. Prevalence of coeliac disease in diabetic children and their first- degree relatives in west Algeria: screening with serological markers. Acta Paediatr Suppl. 1996 May;412:58-60. PMID: 8783762.

51. Bouguerra R, Ben Salem L, Chaabouni H, et al. Celiac disease in adult patients with type 1 diabetes mellitus in Tunisia. Diabetes Metab. 2005 Feb;31(1):83-6. PMID: 15803118.

52. Brottveit M, Raki M, Bergseng E, et al. Assessing possible celiac disease by an HLA-DQ2-gliadin Tetramer Test. Am J Gastroenterol. 2011 Jul;106(7):1318-24. PMID: 21364548.

53. Burgin-Wolff A, Gaze H, Hadziselimovic F, et al. Antigliadin and antiendomysium antibody determination for coeliac disease. Arch Dis Child. 1991 Aug;66(8):941-7. PMID: 1819255.

54. Buysschaert M, Tomasi JP, Hermans MP. Prospective screening for biopsy proven coeliac disease, autoimmunity and malabsorption markers in Belgian subjects with Type 1 diabetes. Diabet Med. 2005 Jul;22(7):889-92. PMID: 15975104.

55. Cannizzaro R, Da Ponte A, Tabuso M, et al. Improving detection of celiac disease patients: a prospective study in iron-deficient blood donors without anemia in north Italy. Eur J Gastroenterol Hepatol. 2014 Jul;26(7):721-4. PMID: 24841904.

56. Carlsson A, Agardh D, Borulf S, et al. Prevalence of celiac disease: before and after a national change in feeding recommendations. Scand J Gastroenterol. 2006 May;41(5):553-8. PMID: 16638697.

57. Carlsson A, Axelsson I, Borulf S, et al. Prevalence of IgA-antigliadin antibodies and IgAantiendomysium antibodies related to celiac disease in children with Down syndrome. Pediatrics. 1998 Feb;101(2):272-5. PMID: 9445503.

58. Carlsson AK, Axelsson IE, Borulf SK, et al. Serological screening for celiac disease in healthy 2.5-year-old children in Sweden. Pediatrics. 2001 Jan;107(1):42-5. PMID: 11134432.

59. Carlsson AK, Axelsson IE, Borulf SK, et al. Prevalence of IgA-antiendomysium and IgAantigliadin autoantibodies at diagnosis of insulin-dependent diabetes mellitus in Swedish children and adolescents. Pediatrics. 1999 Jun;103(6 Pt 1):1248-52. PMID: 10353937.

60. Carnicer J, Farre C, Varea V, et al. Prevalence of coeliac disease in Down's syndrome. Eur J Gastroenterol Hepatol. 2001 Mar;13(3):263-7. PMID: 11293446.

61. Carroccio A, Iannitto E, Cavataio F, et al. Sideropenic anemia and celiac disease: one study, two points of view. Dig Dis Sci. 1998 Mar;43(3):673-8. PMID: 9539667.

62. Casella G, D'Inca R, Oliva L, et al. Prevalence of celiac disease in inflammatory bowel diseases: An IG-IBD multicentre study. Dig Liver Dis. 2010 Mar;42(3):175-8. PMID: 19786375.

63. Casella S, Zanini B, Lanzarotto F, et al. Celiac disease in elderly adults: clinical, serological, and histological characteristics and the effect of a gluten-free diet. J Am Geriatr Soc. 2012 Jun;60(6):1064-9. PMID: 22690983.

64. Cash BD, Rubenstein JH, Young PE, et al. The prevalence of celiac disease among patients with nonconstipated irritable bowel syndrome is similar to controls. Gastroenterology. 2011 Oct;141(4):1187-93. PMID: 21762658.

65. Castano L, Blarduni E, Ortiz L, et al. Prospective population screening for celiac disease: high prevalence in the first 3 years of life. J Pediatr Gastroenterol Nutr. 2004 Jul;39(1):80-4. PMID: 15187786.

66. Castro M, Crino A, Papadatou B, et al. Down's syndrome and celiac disease: the prevalence of high IgA-antigliadin antibodies and HLA-DR and DQ antigens in trisomy 21. J Pediatr Gastroenterol Nutr. 1993 Apr;16(3):265-8. PMID: 8492253.

67. Castro-Antunes MM, Crovella S, Brandao LA, et al. Frequency distribution of HLA DQ2 and DQ8 in celiac patients and first-degree relatives in Recife, northeastern Brazil. Clinics (Sao Paulo). 2011;66(2):227-31. PMID: 21484038.

68. Castro-Antunes MM, Crovella S, Brandao LAC, et al. Fequency distribution of HLA DQ2 and DQ8 in celiac patients and first-degree relatives in Recife, northeastern Brazil. Clinics. 2011;66(2):227-31. PMID: WOS:000289365200008.

69. Cataldo F, Ventura A, Lazzari R, et al. Antiendomysium antibodies and coeliac disease: solved and unsolved questions. An Italian multicentre study. Acta Paediatr. 1995 Oct;84(10):1125-31. PMID: 8563223.

70. Catassi C, Fabiani E, Ratsch IM, et al. The coeliac iceberg in Italy. A multicentre antigliadin antibodies screening for coeliac disease in school-age subjects. Acta Paediatr Suppl. 1996 May;412:29-35. PMID: 8783752.

71. Catassi C, Fanciulli G, D'Appello AR, et al. Antiendomysium versus antigliadin antibodies in screening the general population for coeliac disease. Scand J Gastroenterol. 2000 Jul;35(7):732-6. PMID: 10972177.

72. Catassi C, Ratsch IM, Fabiani E, et al. Coeliac disease in the year 2000: exploring the iceberg. Lancet. 1994 Jan 22;343(8891):200-3. PMID: 7904667.

73. Cerqueira RM, Rocha CM, Fernandes CD, et al. Celiac disease in Portuguese children and adults with Down syndrome. Eur J Gastroenterol Hepatol. 2010 Jul;22(7):868-71. PMID: 20545028.

74. Chan KN, Phillips AD, Mirakian R, et al. Endomysial Antibody Screening in Children. Journal of Pediatric Gastroenterology and Nutrition. 1994 Apr;18(3):316-20. PMID: WOS:A1994NH43000013.

75. Chatzicostas C, Roussomoustakaki M, Drygiannakis D, et al. Primary biliary cirrhosis and autoimmune cholangitis are not associated with coeliac disease in Crete. BMC Gastroenterol. 2002;2:5. PMID: 11914139.

76. Chicco D, Taddio A, Sinagra G, et al. Speeding up coeliac disease diagnosis in cardiological settings. Arch Med Sci. 2010 Oct;6(5):728-32. PMID: 22419932.

77. Chin MW, Mallon DF, Cullen DJ, et al. Screening for coeliac disease using anti-tissue transglutaminase antibody assays, and prevalence of the disease in an Australian community. Med J Aust. 2009 Apr 20;190(8):429-32. PMID: 19374615.

78. Ch'ng CL, Biswas M, Benton A, et al. Prospective screening for coeliac disease in patients with Graves' hyperthyroidism using anti-gliadin and tissue transglutaminase antibodies. Clin Endocrinol (Oxf). 2005 Mar;62(3):303-6. PMID: 15730411.

79. Chogle A, Saps M. Yield and cost of performing screening tests for constipation in children. Can J Gastroenterol. 2013 Dec;27(12):e35-8. PMID: 24228262.

80. Choi JM, Lebwohl B, Wang J, et al. Increased prevalence of celiac disease in patients with unexplained infertility in the United States. J Reprod Med. 2011 May-Jun;56(5-6):199-203. PMID: 21682114.

81. Coburn JA, Vande Voort JL, Lahr BD, et al. Human leukocyte antigen genetics and clinical features of self-treated patients on a gluten-free diet. J Clin Gastroenterol. 2013 Nov-Dec;47(10):828-33. PMID: 23632357.

82. Cogulu O, Ozkinay F, Gunduz C, et al. Celiac disease in children with Down syndrome: importance of follow-up and serologic screening. Pediatr Int. 2003 Aug;45(4):395-9. PMID: 12911473.

83. Collin P, Syrjanen J, Partanen J, et al. Celiac disease and HLA DQ in patients with IgA nephropathy. Am J Gastroenterol. 2002 Oct;97(10):2572-6. PMID: 12385441.

84. Contreas G, Valletta E, Ulmi D, et al. Screening of coeliac disease in north Italian children with type 1 diabetes: limited usefulness of HLA-DQ typing. Acta Paediatr. 2004 May;93(5):628-32. PMID: 15174785.

85. Cook HB, Burt MJ, Collett JA, et al. Adult coeliac disease: prevalence and clinical significance. J Gastroenterol Hepatol. 2000 Sep;15(9):1032-6. PMID: 11059933.

86. Corazza GR, Valentini RA, Andreani ML, et al. Subclinical coeliac disease is a frequent cause of iron-deficiency anaemia. Scand J Gastroenterol. 1995 Feb;30(2):153-6. PMID: 7732338.

87. Costa S, Astarita L, Ben-Hariz M, et al. A Point-of-Care test for facing the burden of undiagnosed celiac disease in the Mediterranean area: a pragmatic design study. BMC Gastroenterol. 2014 Dec 18;14(1):219. PMID: 25518884.

88. Crone J, Rami B, Huber WD, et al. Prevalence of celiac disease and follow-up of EMA in children and adolescents with type 1 diabetes mellitus. J Pediatr Gastroenterol Nutr. 2003 Jul;37(1):67-71. PMID: 12827008.

89. Crovella S, Brandao L, Guimaraes R, et al. Speeding up coeliac disease diagnosis in the developing countries. Dig Liver Dis. 2007 Oct;39(10):900-2. PMID: 17706474.

90. Csizmadia CG, Mearin ML, Oren A, et al. Accuracy and cost-effectiveness of a new strategy to screen for celiac disease in children with Down syndrome. J Pediatr. 2000 Dec;137(6):756-61. PMID: 11113830.

91. Cuoco L, Certo M, Jorizzo RA, et al. Prevalence and early diagnosis of coeliac disease in autoimmune thyroid disorders. Ital J Gastroenterol Hepatol. 1999 May;31(4):283-7. PMID: 10425571.

92. Curione M, Barbato M, De Biase L, et al. Prevalence of coeliac disease in idiopathic dilated cardiomyopathy. Lancet. 1999 Jul 17;354(9174):222-3. PMID: 10421311.

93. Dai AI, Akcali A, Varan C, et al. Prevalence of resistant occipital lobe epilepsy associated with celiac disease in children. Childs Nerv Syst. 2014 Feb 25PMID: 24566676.

94. Dalgic B, Sari S, Basturk B, et al. Prevalence of celiac disease in healthy Turkish school children. Am J Gastroenterol. 2011 Aug;106(8):1512-7. PMID: 21691340.

95. Day AS, Cook HB, Whitehead M, et al. Anti-endomysial and anti-gliadin antibodies in screening for coeliac disease in children at greater risk of developing coeliac disease. N Z Med J. 2000 Oct 13;113(1119):412-3. PMID: 11127356.

96. De Bem RS, Da Ro Sa Utiyama SR, Nisihara RM, et al. Celiac disease prevalence in Brazilian dilated cardiomyopathy patients. Dig Dis Sci. 2006 May;51(5):1016-9. PMID: 16758314.

97. de Lecea A, Ribes-Koninckx C, Polanco I, et al. Serological screening (antigliadin and antiendomysium antibodies) for non-overt coeliac disease in children of short stature. Acta Paediatr Suppl. 1996 May;412:54-5. PMID: 8783760.

98. De Vitis I, Ghirlanda G, Gasbarrini G. Prevalence of coeliac disease in type I diabetes: a multicentre study. Acta Paediatr Suppl. 1996 May;412:56-7. PMID: 8783761.

99. Dehghani SM, Asadi-Pooya AA. Celiac disease in children with short stature. Indian J Pediatr. 2008 Feb;75(2):131-3. PMID: 18334792.

100. Demirceken FG, Kansu A, Kuloglu Z, et al. Human tissue transglutaminase antibody screening by immunochromatographic line immunoassay for early diagnosis of celiac disease in Turkish children. Turk J Gastroenterol. 2008 Mar;19(1):14-21. PMID: 18386235.

101. Diamanti A, Colistro F, Calce A, et al. Clinical value of immunoglobulin A antitransglutaminase assay in the diagnosis of celiac disease. Pediatrics. 2006 Dec;118(6):e1696-700. PMID: 17074840.

102. Dogan Y, Yildirmaz S, Ozercan IH. Prevalence of celiac disease among first-degree relatives of patients with celiac disease. J Pediatr Gastroenterol Nutr. 2012 Aug;55(2):205-8. PMID: 22241509.

103. Donaldson MR, Book LS, Leiferman KM, et al. Strongly positive tissue transglutaminase antibodies are associated with Marsh 3 histopathology in adult and pediatric celiac disease. J Clin Gastroenterol. 2008 Mar;42(3):256-60. PMID: 18223500.

104. Doolan A, Donaghue K, Fairchild J, et al. Use of HLA typing in diagnosing celiac disease in patients with type 1 diabetes. Diabetes Care. 2005 Apr;28(4):806-9. PMID: 15793177.

105. Drastich P, Honsova E, Lodererova A, et al. Celiac disease markers in patients with liver diseases: a single center large scale screening study. World J Gastroenterol. 2012 Nov 21;18(43):6255-62. PMID: 23180946.

106. Eapen CE, Nightingale P, Hubscher SG, et al. Non-cirrhotic intrahepatic portal hypertension: associated gut diseases and prognostic factors. Dig Dis Sci. 2011 Jan;56(1):227-35. PMID: 20499175.

107. El-Salhy M, Lomholt-Beck B, Gundersen D. The prevalence of celiac disease in patients with irritable bowel syndrome. Mol Med Rep. 2011 May-Jun;4(3):403-5. PMID: 21468583.

108. Elsurer R, Tatar G, Simsek H, et al. Celiac disease in the Turkish population. Dig Dis Sci. 2005 Jan;50(1):136-42. PMID: 15712651.

109. Emami MH, Hashemi M, Kouhestani S, et al. Should We Look for Celiac Disease among all Patients with Liver Function Test Abnormalities? Int J Prev Med. 2012 Mar;3(3):167-72. PMID: 22448309.

110. Emami MH, Kouhestani S, Karimi S, et al. Frequency of celiac disease in adult patients with typical or atypical malabsorption symptoms in isfahan, iran. Gastroenterol Res Pract. 2012;2012:106965. PMID: 22545042.

111. Ergur AT, Ocal G, Berberoglu M, et al. Celiac disease and autoimmune thyroid disease in children with type 1 diabetes mellitus: clinical and HLA-genotyping results. J Clin Res Pediatr Endocrinol. 2010;2(4):151-4. PMID: 21274314.

112. Ersoy O, Akin E, Ugras S, et al. Capsule endoscopy findings in celiac disease. Dig Dis Sci. 2009 Apr;54(4):825-9. PMID: 18649134.

113. Ertekin V, Tozun MS, Kucuk N. The prevalence of celiac disease in children with irondeficiency anemia. Turk J Gastroenterol. 2013 Aug;24(4):334-8. PMID: 24254265.

114. Evans KE, Malloy AR, Gorard DA. Changing patterns of coeliac serology requests. Aliment Pharmacol Ther. 2009 May 15;29(10):1137-42. PMID: 19243355.

115. Fabiani E, Catassi C. The serum IgA class anti-tissue transglutaminase antibodies in the diagnosis and follow up of coeliac disease. Results of an international multi-centre study. International Working Group on Eu-tTG. Eur J Gastroenterol Hepatol. 2001 Jun;13(6):659-65. PMID: 11434591.

116. Farre C, Humbert P, Vilar P, et al. Serological markers and HLA-DQ2 haplotype among first-degree relatives of celiac patients. Catalonian Coeliac Disease Study Group. Dig Dis Sci. 1999 Nov;44(11):2344-9. PMID: 10573385.

117. Ferfoglia G, Pulitano R, Sategna-Guidetti C. Do dietary antibodies still play a role in the diagnosis and follow-up of coeliac disease? A comparison among different serological tests. Panminerva Med. 1995 Jun;37(2):55-9. PMID: 8637769.

118. Fernandez-Banares F, Esteve M, Farre C, et al. Predisposing HLA-DQ2 and HLA-DQ8 haplotypes of coelialc disease and associated enteropathy in microscopic colitis. European Journal of Gastroenterology & Hepatology. 2005 Dec;17(12):1333-8. PMID: WOS:000233798100010.

119. Fine KD, Do K, Schulte K, et al. High prevalence of celiac sprue-like HLA-DQ genes and enteropathy in patients with the microscopic colitis syndrome. Am J Gastroenterol. 2000 Aug;95(8):1974-82. PMID: 10950045.

120. Fine KD, Ogunji F, Saloum Y, et al. Celiac sprue: another autoimmune syndrome associated with hepatitis C. Am J Gastroenterol. 2001 Jan;96(1):138-45. PMID: 11197243.

121. Floreani A, Betterle C, Baragiotta A, et al. Prevalence of coeliac disease in primary biliary cirrhosis and of antimitochondrial antibodies in adult coeliac disease patients in Italy. Dig Liver Dis. 2002 Apr;34(4):258-61. PMID: 12038809.

122. Francis J, Carty JE, Scott BB. The prevalence of coeliac disease in rheumatoid arthritis. Eur J Gastroenterol Hepatol. 2002 Dec;14(12):1355-6. PMID: 12468957.

123. Fraser JS, King AL, Ellis HJ, et al. An algorithm for family screening for coeliac disease. World J Gastroenterol. 2006 Dec 28;12(48):7805-9. PMID: 17203524.

124. Fraser-Reynolds KA, Butzner JD, Stephure DK, et al. Use of immunoglobulin Aantiendomysial antibody to screen for celiac disease in North American children with type 1 diabetes. Diabetes Care. 1998 Nov;21(11):1985-9. PMID: 9802755.

125. Freeman HJ. Strongly positive tissue transglutaminase antibody assays without celiac disease. Can J Gastroenterol. 2004 Jan;18(1):25-8. PMID: 14760428.

126. Frost AR, Band MM, Conway GS. Serological screening for coeliac disease in adults with Turner's syndrome: prevalence and clinical significance of endomysium antibody positivity. Eur J Endocrinol. 2009 Apr;160(4):675-9. PMID: 19208776.

127. Frustaci A, Cuoco L, Chimenti C, et al. Celiac disease associated with autoimmune myocarditis. Circulation. 2002 Jun 4;105(22):2611-8. PMID: 12045166.

128. Gabriel S, Mihaela I, Angela B, et al. New Para-Clinical Investigations in the Celiac Disease. Revista Romana De Medicina De Laborator. 2010 Jun;18(2):43-51. PMID: WOS:000279287500005.

129. Gabrielli M, Candelli M, Cremonini F, et al. Idiopathic chronic urticaria and celiac disease. Dig Dis Sci. 2005 Sep;50(9):1702-4. PMID: 16133973.

130. Gale L, Wimalaratna H, Brotodiharjo A, et al. Down's syndrome is strongly associated with coeliac disease. Gut. 1997 Apr;40(4):492-6. PMID: 9176077.

131. Gandolfi L, Catassi C, Garcia S, et al. Antiendomysial antibody test reliability in children with frequent diarrhea and malnutrition: is it celiac disease? J Pediatr Gastroenterol Nutr. 2001 Oct;33(4):483-7. PMID: 11698768.

132. Gatselis NK, Zachou K, Norman GL, et al. IgA antibodies against deamidated gliadin peptides in patients with chronic liver diseases. Clin Chim Acta. 2012 Oct 9;413(19-20):1683-8. PMID: 22643316.

133. Gautam A, Jain BK, Midha V, et al. Prevalence of celiac disease among siblings of celiac disease patients. Indian J Gastroenterol. 2006 Sep-Oct;25(5):233-5. PMID: 17090839.

134. Ghozzi M, Sakly W, Mankai A, et al. Screening for celiac disease, by endomysial antibodies, in patients with unexplained articular manifestations. Rheumatol Int. 2014 May;34(5):637-42. PMID: 24292850.

135. Gillett PM, Gillett HR, Israel DM, et al. High prevalence of celiac disease in patients with type 1 diabetes detected by antibodies to endomysium and tissue transglutaminase. Can J Gastroenterol. 2001 May;15(5):297-301. PMID: 11381296.

136. Giovenale D, Meazza C, Cardinale GM, et al. The prevalence of growth hormone deficiency and celiac disease in short children. Clin Med Res. 2006 Sep;4(3):180-3. PMID: 16988097.

137. Glastras SJ, Craig ME, Verge CF, et al. The role of autoimmunity at diagnosis of type 1 diabetes in the development of thyroid and celiac disease and microvascular complications. Diabetes Care. 2005 Sep;28(9):2170-5. PMID: 16123485.

138. Goh C, Banerjee K. Prevalence of coeliac disease in children and adolescents with type 1 diabetes mellitus in a clinic based population. Postgrad Med J. 2007 Feb;83(976):132-6. PMID: 17308219.

139. Gokhale YA, Sawant PD, Chodankar CM, et al. Celiac disease in osteoporotic Indians. J Assoc Physicians India. 2003 Jun;51:579-83. PMID: 15266924.

140. Gomez JC, Selvaggio G, Pizarro B, et al. Value of a screening algorithm for celiac disease using tissue transglutaminase antibodies as first level in a population-based study. Am J Gastroenterol. 2002 Nov;97(11):2785-90. PMID: 12425549.

141. Gudjonsdottir AH, Nilsson S, Ek J, et al. The risk of celiac disease in 107 families with at least two affected siblings. J Pediatr Gastroenterol Nutr. 2004 Mar;38(3):338-42. PMID: 15076637.

142. Guliter S, Yakaryilmaz F, Ozkurt Z, et al. Prevalence of coeliac disease in patients with autoimmune thyroiditis in a Turkish population. World J Gastroenterol. 2007 Mar 14;13(10):1599-601. PMID: 17461455.

143. Gungor S, Celiloglu OS, Ozcan OO, et al. Frequency of celiac disease in attentiondeficit/hyperactivity disorder. J Pediatr Gastroenterol Nutr. 2013 Feb;56(2):211-4. PMID: 22983377.

144. Gursoy S, Guven K, Simsek T, et al. The prevalence of unrecognized adult celiac disease in Central Anatolia. J Clin Gastroenterol. 2005 Jul;39(6):508-11. PMID: 15942437.

145. Guvenc S, Kaymakoglu S, Gurel N, et al. The prevalence of manifest and latent celiac disease in type 1 diabetes mellitus. Turk J Gastroenterol. 2002 Jun;13(2):103-7. PMID: 16378286.

146. Hadjivassiliou M, Aeschlimann P, Sanders DS, et al. Transglutaminase 6 antibodies in the diagnosis of gluten ataxia. Neurology. 2013 May 7;80(19):1740-5. PMID: 23576621.

147. Hadjivassiliou M, Grunewald RA, Chattopadhyay AK, et al. Clinical, radiological, neurophysiological, and neuropathological characteristics of gluten ataxia. Lancet. 1998 Nov 14;352(9140):1582-5. PMID: 9843103.

148. Hamidian Y, Togha M, Nafisi S, et al. Antigliadin antibody in sporadic adult ataxia. Iran J Neurol. 2012;11(1):16-20. PMID: 24250853.

149. Hansson T, Anneren G, Sjoberg O, et al. Celiac disease in relation to immunologic serum markers, trace elements, and HLA-DR and DQ antigens in Swedish children with Down syndrome. Journal of Pediatric Gastroenterology and Nutrition. 1999 Sep;29(3):286-92. PMID: WOS:000082043300009.

150. Hansson T, Dahlbom I, Rogberg S, et al. Antitissue transglutaminase and antithyroid autoantibodies in children with Down syndrome and celiac disease. J Pediatr Gastroenterol Nutr. 2005 Feb;40(2):170-4; discussion 25-7. PMID: 15699691.

151. Hernandez L, Johnson TC, Naiyer AJ, et al. Chronic hepatitis C virus and celiac disease, is there an association? Dig Dis Sci. 2008 Jan;53(1):256-61. PMID: 17549632.

152. Hin H, Bird G, Fisher P, et al. Coeliac disease in primary care: case finding study. BMJ. 1999 Jan 16;318(7177):164-7. PMID: 9888912.

153. Hoffenberg EJ, Bao F, Eisenbarth GS, et al. Transglutaminase antibodies in children with a genetic risk for celiac disease. J Pediatr. 2000 Sep;137(3):356-60. PMID: 10969260.

154. Hoffenberg EJ, Emery LM, Barriga KJ, et al. Clinical features of children with screeningidentified evidence of celiac disease. Pediatrics. 2004 May;113(5):1254-9. PMID: 15121938.

155. Hoffenberg EJ, MacKenzie T, Barriga KJ, et al. A prospective study of the incidence of childhood celiac disease. J Pediatr. 2003 Sep;143(3):308-14. PMID: 14517510.

156. Hogberg L, Falth-Magnusson K, Grodzinsky E, et al. Familial prevalence of coeliac disease: a twenty-year follow-up study. Scand J Gastroenterol. 2003 Jan;38(1):61-5. PMID: 12608466.

157. Hogen Esch CE, Csizmadia GD, van Hoogstraten IM, et al. Childhood coeliac disease: towards an improved serological mass screening strategy. Aliment Pharmacol Ther. 2010 Apr;31(7):760-6. PMID: 20047580.

158. Hojsak I, Mozer-Glassberg Y, Segal Gilboa N, et al. Celiac disease screening assays for children younger than 3 years of age: the performance of three serological tests. Dig Dis Sci. 2012 Jan;57(1):127-32. PMID: 21847565.

159. Hojsak I, Zevit N, Waisbourd-Zinman O, et al. Concomitant autoantibodies in newly diagnosed diabetic children with transient celiac serology or proven celiac disease. J Pediatr Endocrinol Metab. 2013;26(11-12):1099-104. PMID: 23817597.

160. Howard MR, Turnbull AJ, Morley P, et al. A prospective study of the prevalence of undiagnosed coeliac disease in laboratory defined iron and folate deficiency. J Clin Pathol. 2002 Oct;55(10):754-7. PMID: 12354801.

161. Hummel M, Bonifacio E, Stern M, et al. Development of celiac disease-associated antibodies in offspring of parents with type I diabetes. Diabetologia. 2000 Aug;43(8):1005-11. PMID: 10990078.

162. Imanzadeh F, Sayyari AA, Yaghoobi M, et al. Celiac disease in children with diarrhea is more frequent than previously suspected. J Pediatr Gastroenterol Nutr. 2005 Mar;40(3):309-11. PMID: 15735484.

163. Iuorio R, Mercuri V, Barbarulo F, et al. Prevalence of celiac disease in patients with autoimmune thyroiditis. Minerva Endocrinol. 2007 Dec;32(4):239-43. PMID: 18091661.

164. Ivarsson A, Myleus A, Norstrom F, et al. Prevalence of childhood celiac disease and changes in infant feeding. Pediatrics. 2013 Mar;131(3):e687-94. PMID: 23420914.

165. Ivarsson SA, Carlsson A, Bredberg A, et al. Prevalence of coeliac disease in Turner syndrome. Acta Paediatr. 1999 Sep;88(9):933-6. PMID: 10519331.

166. Jadallah KA, Khader YS. Celiac disease in patients with presumed irritable bowel syndrome: a case-finding study. World J Gastroenterol. 2009 Nov 14;15(42):5321-5. PMID: 19908341.

167. Jansson U, Johansson C. Down syndrome and celiac disease. J Pediatr Gastroenterol Nutr. 1995 Nov;21(4):443-5. PMID: 8583297.

168. Jokinen J, Peters U, Maki M, et al. Celiac sprue in patients with chronic oral mucosal symptoms. J Clin Gastroenterol. 1998 Jan;26(1):23-6. PMID: 9492858.

169. Jores RD, Frau F, Cucca F, et al. HLA-DQB1*0201 homozygosis predisposes to severe intestinal damage in celiac disease. Scandinavian Journal of Gastroenterology. 2007 Jan;42(1):48-53. PMID: WOS:000243724800009.

170. Kabbani TA, Vanga RR, Leffler DA, et al. Celiac Disease or Non-Celiac Gluten Sensitivity? An Approach to Clinical Differential Diagnosis. Am J Gastroenterol. 2014 Mar 11PMID: 24619056.

171. Kakleas K, Karayianni C, Critselis E, et al. The prevalence and risk factors for coeliac disease among children and adolescents with type 1 diabetes mellitus. Diabetes Res Clin Pract. 2010 Nov;90(2):202-8. PMID: 20832887.

172. Kalayci AG, Kanber Y, Birinci A, et al. The prevalence of coeliac disease as detected by screening in children with iron deficiency anaemia. Acta Paediatr. 2005 Jun;94(6):678-81. PMID: 16188768.

173. Karagiozoglou-Lampoudi T, Zellos A, Vlahavas G, et al. Screening for coeliac disease in preschool Greek children: the feasibility study of a community-based project. Acta Paediatr. 2013 Jul;102(7):749-54. PMID: 23600795.

174. Karakan T, Ozyemisci-Taskiran O, Gunendi Z, et al. Prevalence of IgA-antiendomysial antibody in a patient cohort with idiopathic low bone mineral density. World J Gastroenterol. 2007 Jun 7;13(21):2978-82. PMID: 17589950.

175. Karinen H, Karkkainen P, Pihlajamaki J, et al. HLA genotyping is useful in the evaluation of the risk for coeliac disease in the 1st-degree relatives of patients with coeliac disease. Scand J Gastroenterol. 2006 Nov;41(11):1299-304. PMID: 17060123.

176. Karnam US, Felder LR, Raskin JB. Prevalence of occult celiac disease in patients with iron-deficiency anemia: a prospective study. South Med J. 2004 Jan;97(1):30-4. PMID: 14746419.

177. Karnsakul W, Skitarelic K, Gillespie S, et al. Isolated positive anti-gliadin immunoglobin-A antibody in children with gastrointestinal symptoms. Turk J Gastroenterol. 2012;23(5):485-9. PMID: 23161323.

178. Kasperkiewicz M, Dahnrich C, Probst C, et al. Novel assay for detecting celiac diseaseassociated autoantibodies in dermatitis herpetiformis using deamidated gliadin-analogous fusion peptides. J Am Acad Dermatol. 2012 Apr;66(4):583-8. PMID: 21840083.

179. Katsinelos P, Fasoylas K, Chatzimavroudis G, et al. Diagnostic yield and clinical management after capsule endoscopy in daily clinical practice: A single-center experience. Hippokratia. 2010 Oct;14(4):271-6. PMID: 21311636.

180. Katz KD, Rashtak S, Lahr BD, et al. Screening for celiac disease in a North American population: sequential serology and gastrointestinal symptoms. Am J Gastroenterol. 2011 Jul;106(7):1333-9. PMID: 21364545.

181. Kaukinen K, Partanen J, Maki M, et al. HLA-DQ typing in the diagnosis of celiac disease. Am J Gastroenterol. 2002 Mar;97(3):695-9. PMID: 11922565.

182. Kavimandan A, Sharma M, Verma AK, et al. Prevalence of celiac disease in nutritional anemia at a tertiary care center. Indian J Gastroenterol. 2014 Mar;33(2):114-8. PMID: 23996798.

183. Keshavarz AA, Bashiri H, Ahmadi A, et al. The Prevalence of Occult Celiac Disease among Patients with Functional Dyspepsia: A Study from the Western Region of Iran. Gastroenterol Res Pract. 2010;2010:170702. PMID: 21151702.

184. Keswani RN, Neven K, Semrad CE. Screening for celiac disease in short bowel syndrome. Nutr Clin Pract. 2008 Feb;23(1):72-5. PMID: 18203966.

185. Kocsis D, Miheller P, Lorinczy K, et al. Coeliac disease in a 15-year period of observation (1997 and 2011) in a Hungarian referral centre. Eur J Intern Med. 2013 Jul;24(5):461-7. PMID: 23535227.

186. Koehne Vde B, Bahia M, Lanna CC, et al. Prevalence of serological markers for celiac disease (IgA and IgG class antigliadin antibodies and IgA class antiendomysium antibodies) in patients with autoimmune rheumatologic diseases in Belo Horizonte, MG, Brazil. Arq Gastroenterol. 2010 Jul-Sep;47(3):250-6. PMID: 21140085.

187. Kordonouri O, Dieterich W, Schuppan D, et al. Autoantibodies to tissue transglutaminase are sensitive serological parameters for detecting silent coeliac disease in patients with Type 1 diabetes mellitus. Diabet Med. 2000 Jun;17(6):441-4. PMID: 10975212.

188. Korkut E, Bektas M, Oztas E, et al. The prevalence of celiac disease in patients fulfilling Rome III criteria for irritable bowel syndrome. Eur J Intern Med. 2010 Oct;21(5):389-92. PMID: 20816591.

189. Korponay-Szabo IR, Szabados K, Pusztai J, et al. Population screening for coeliac disease in primary care by district nurses using a rapid antibody test: diagnostic accuracy and feasibility study. BMJ. 2007 Dec 15;335(7632):1244-7. PMID: 18063612.

190. Kotze LM, Brambila Rodrigues AP, Kotze LR, et al. A Brazilian experience of the self transglutaminase-based test for celiac disease case finding and diet monitoring. World J Gastroenterol. 2009 Sep 21;15(35):4423-8. PMID: 19764094.

191. Kratzer W, Kibele M, Akinli A, et al. Prevalence of celiac disease in Germany: a prospective follow-up study. World J Gastroenterol. 2013 May 7;19(17):2612-20. PMID: 23674868.

192. Kumar V, Jarzabek-Chorzelska M, Sulej J, et al. Celiac disease and immunoglobulin a deficiency: how effective are the serological methods of diagnosis? Clin Diagn Lab Immunol. 2002 Nov;9(6):1295-300. PMID: 12414763.

193. Kumar V, Jarzabek-Chorzelska M, Sulej J, et al. Tissue transglutaminase and endomysial antibodies-diagnostic markers of gluten-sensitive enteropathy in dermatitis herpetiformis. Clin Immunol. 2001 Mar;98(3):378-82. PMID: 11237562.

194. Kurien M, Evans KE, Aziz I, et al. Capsule endoscopy in adult celiac disease: a potential role in equivocal cases of celiac disease? Gastrointest Endosc. 2013 Feb;77(2):227-32. PMID: 23200728.

195. Kurien M, Leeds JS, Hopper AD, et al. Serological testing for coeliac disease in Type 1 diabetes mellitus: is immunoglobulin A level measurement necessary? Diabet Med. 2013 Jul;30(7):840-5. PMID: 23461783.

196. Kurppa K, Collin P, Sievanen H, et al. Gastrointestinal symptoms, quality of life and bone mineral density in mild enteropathic coeliac disease: a prospective clinical trial. Scand J Gastroenterol. 2010 Mar;45(3):305-14. PMID: 20059405.

197. Kurppa K, Salminiemi J, Ukkola A, et al. Utility of the new ESPGHAN criteria for the diagnosis of celiac disease in at-risk groups. J Pediatr Gastroenterol Nutr. 2012 Mar;54(3):387-91. PMID: 22094901.

198. Labate A, Gambardella A, Messina D, et al. Silent celiac disease in patients with childhood localization-related epilepsies. Epilepsia. 2001 Sep;42(9):1153-5. PMID: 11580763.

199. Lagerqvist C, Ivarsson A, Juto P, et al. Screening for adult coeliac disease - which serological marker(s) to use? J Intern Med. 2001 Sep;250(3):241-8. PMID: 11555129.

200. Lahteenoja H, Toivanen A, Raiha I, et al. Salivary antigliadin and antiendomysium antibodies in coeliac disease. Scand J Immunol. 1999 Nov;50(5):528-35. PMID: 10564556.
201. Lanzini A, Villanacci V, Apillan N, et al. Epidemiological, clinical and histopathologic characteristics of celiac disease: results of a case-finding population-based program in an Italian community. Scand J Gastroenterol. 2005 Aug;40(8):950-7. PMID: 16165709.

202. Lecleire S, Di Fiore F, Antonietti M, et al. Endoscopic markers of villous atrophy are not useful for the detection of celiac disease in patients with dyspeptic symptoms. Endoscopy. 2006 Jul;38(7):696-701. PMID: 16761210.

203. Leeds JS, Hopper AD, Hadjivassiliou M, et al. High prevalence of microvascular complications in adults with type 1 diabetes and newly diagnosed celiac disease. Diabetes Care. 2011 Oct;34(10):2158-63. PMID: 21911773.

204. Lenhardt A, Plebani A, Marchetti F, et al. Role of human-tissue transglutaminase IgG and anti-gliadin IgG antibodies in the diagnosis of coeliac disease in patients with selective immunoglobulin A deficiency. Dig Liver Dis. 2004 Nov;36(11):730-4. PMID: 15571003.

205. Lepore L, Martelossi S, Pennesi M, et al. Prevalence of celiac disease in patients with juvenile chronic arthritis. J Pediatr. 1996 Aug;129(2):311-3. PMID: 8765635.

206. Lewis C, Book L, Black J, et al. Celiac disease and human leukocyte antigen genotype: accuracy of diagnosis in self-diagnosed individuals, dosage effect, and sibling risk. J Pediatr Gastroenterol Nutr. 2000 Jul;31(1):22-7. PMID: 10896066.

207. Lima VM, Gandolfi L, Pires JA, et al. Prevalence of celiac disease in dyspeptic patients. Arq Gastroenterol. 2005 Jul-Sep;42(3):153-6. PMID: 16200250.

208. Lindgren S, Sjoberg K, Eriksson S. Unsuspected coeliac disease in chronic 'cryptogenic' liver disease. Scand J Gastroenterol. 1994 Jul;29(7):661-4. PMID: 7939405.

209. Liu E, Bao F, Barriga K, et al. Fluctuating transglutaminase autoantibodies are related to histologic features of celiac disease. Clin Gastroenterol Hepatol. 2003 Sep;1(5):356-62. PMID: 15017653.

210. Lo Iacono O, Petta S, Venezia G, et al. Anti-tissue transglutaminase antibodies in patients with abnormal liver tests: is it always coeliac disease? Am J Gastroenterol. 2005 Nov;100(11):2472-7. PMID: 16279902.

211. Lock RJ, Unsworth DJ. Identifying immunoglobulin-A--deficient children and adults does not necessarily help the serologic diagnosis of coeliac disease. J Pediatr Gastroenterol Nutr. 1999 Jan;28(1):81-3. PMID: 9890474.

212. Lorini R, Scotta MS, Cortona L, et al. Celiac disease and type I (insulin-dependent) diabetes mellitus in childhood: follow-up study. J Diabetes Complications. 1996 May-Jun;10(3):154-9. PMID: 8807465.

213. Ludvigsson JF, Ansved P, Falth-Magnusson K, et al. Symptoms and signs have changed in Swedish children with coeliac disease. J Pediatr Gastroenterol Nutr. 2004 Feb;38(2):181-6. PMID: 14734881.

214. Luzza F, Mancuso M, Imeneo M, et al. Helicobacter pylori infection in children with celiac disease: prevalence and clinicopathologic features. J Pediatr Gastroenterol Nutr. 1999 Feb;28(2):143-6. PMID: 9932844.

215. Lytton SD, Antiga E, Pfeiffer S, et al. Neo-epitope tissue transglutaminase autoantibodies as a biomarker of the gluten sensitive skin disease--dermatitis herpetiformis. Clin Chim Acta. 2013 Jan 16;415:346-9. PMID: 23142793.

216. Machado AP, Silva LR, Zausner B, et al. Undiagnosed celiac disease in women with infertility. J Reprod Med. 2013 Jan-Feb;58(1-2):61-6. PMID: 23447921.

217. Magaudda A, Dalla Bernardina B, De Marco P, et al. Bilateral occipital calcification, epilepsy and coeliac disease: clinical and neuroimaging features of a new syndrome. J Neurol Neurosurg Psychiatry. 1993 Aug;56(8):885-9. PMID: 8350105.

218. Maguire AA, Greenson JK, Lauwers GY, et al. Collagenous sprue: a clinicopathologic study of 12 cases. Am J Surg Pathol. 2009 Oct;33(10):1440-9. PMID: 19641452.

219. Mahmud FH, Murray JA, Kudva YC, et al. Celiac disease in type 1 diabetes mellitus in a North American community: prevalence, serologic screening, and clinical features. Mayo Clin Proc. 2005 Nov;80(11):1429-34. PMID: 16295022.

220. Marai I, Shoenfeld Y, Bizzaro N, et al. IgA and IgG tissue transglutaminase antibodies in systemic lupus erythematosus. Lupus. 2004;13(4):241-4. PMID: 15176659.

221. Marine M, Farre C, Alsina M, et al. The prevalence of coeliac disease is significantly higher in children compared with adults. Aliment Pharmacol Ther. 2011 Feb;33(4):477-86. PMID: 21166832.

222. Martinelli P, Troncone R, Paparo F, et al. Coeliac disease and unfavourable outcome of pregnancy. Gut. 2000 Mar;46(3):332-5. PMID: 10673293.

223. Martins Rde C, Gandolfi L, Modelli IC, et al. Serologic screening and genetic testing among brazilian patients with celiac disease and their first degree relatives. Arq Gastroenterol. 2010 Jul-Sep;47(3):257-62. PMID: 21140086.

224. Masjedizadeh R, Hajiani E, Hashemi J, et al. Celiac disease in South-West of Iran. World J Gastroenterol. 2006 Jul 21;12(27):4416-9. PMID: 16865789.

225. Mastrandrea F, Semeraro FP, Coradduzza G, et al. CD34+ hemopoietic precursor and stem cells traffic in peripheral blood of celiac patients is significantly increased but not directly related

to epithelial damage severity. Eur Ann Allergy Clin Immunol. 2008 Nov;40(3):90-103. PMID: 19334373.

226. Mather KJ, Meddings JB, Beck PL, et al. Prevalence of IgA-antiendomysial antibody in asymptomatic low bone mineral density. Am J Gastroenterol. 2001 Jan;96(1):120-5. PMID: 11197240.

227. Matteoni CA, Goldblum JR, Wang N, et al. Celiac disease is highly prevalent in lymphocytic colitis. J Clin Gastroenterol. 2001 Mar;32(3):225-7. PMID: 11246349.

228. Mazzilli MC, Ferrante P, Mariani P, et al. A study of Italian pediatric celiac disease patients confirms that the primary HLA association is to the DQ(alpha 1*0501, beta 1*0201) heterodimer. Hum Immunol. 1992 Feb;33(2):133-9. PMID: 1563982.

229. McGowan KE, Lyon ME, Butzner JD. Celiac disease and IgA deficiency: complications of serological testing approaches encountered in the clinic. Clin Chem. 2008 Jul;54(7):1203-9. PMID: 18487281.

230. McGowan KE, Lyon ME, Loken SD, et al. Celiac disease: are endomysial antibody test results being used appropriately? Clin Chem. 2007 Oct;53(10):1775-81. PMID: 17693523.

231. Mein SM, Ladabaum U. Serological testing for coeliac disease in patients with symptoms of irritable bowel syndrome: a cost-effectiveness analysis. Aliment Pharmacol Ther. 2004 Jun 1;19(11):1199-210. PMID: 15153173.

232. Meloni GF, Dessole S, Vargiu N, et al. The prevalence of coeliac disease in infertility. Hum Reprod. 1999 Nov;14(11):2759-61. PMID: 10548618.

233. Meloni GF, Tomasi PA, Bertoncelli A, et al. Prevalence of silent celiac disease in patients with autoimmune thyroiditis from Northern Sardinia. J Endocrinol Invest. 2001 May;24(5):298-302. PMID: 11407647.

234. Menekse E, Selimoglu MA, Temel I, et al. Celiac disease in children with urolithiasis. Turk J Pediatr. 2012 Jul-Aug;54(4):382-6. PMID: 23692719.

235. Menezes TM, Motta ME. Celiac disease prevalence in children and adolescents with myocarditis and dilated cardiomiopathy. J Pediatr (Rio J). 2012 Sep-Oct;88(5):439-42. PMID: 23093320.

236. Modelli IC, Gandolfi L, Almeida RC, et al. Serological screening for celiac disease in symptomatic 12 to 36 month-old children. Arq Gastroenterol. 2010 Jan-Mar;47(1):61-5. PMID: 20520977.

237. Mohammed IM, Karrar ZE, El-Safi SH. Coeliac disease in Sudanese children with clinical features suggestive of the disease. East Mediterr Health J. 2006 Sep;12(5):582-9. PMID: 17333797.

238. Monsuur AJ, de Bakker PI, Zhernakova A, et al. Effective detection of human leukocyte antigen risk alleles in celiac disease using tag single nucleotide polymorphisms. PLoS One. 2008;3(5):e2270. PMID: 18509540.

239. Mora M, Litwin N, Toca MD, et al. Prevalence of celiac disease: a multicenter trial in pediatric population from five urban districts of Argentina. Archivos Argentinos De Pediatria. 2012 Dec;110(6):490-5. PMID: WOS:000326563200013.

240. Mubarak A, Wolters VM, Gmelig-Meyling FH, et al. Tissue transglutaminase levels above 100 U/mL and celiac disease: a prospective study. World J Gastroenterol. 2012 Aug 28;18(32):4399-403. PMID: 22969205.

241. Mugica F, Castiella A, Otazua P, et al. Prevalence of coeliac disease in unexplained chronic hypertransaminasemia. Rev Esp Enferm Dig. 2001 Nov;93(11):707-14. PMID: 11995370.

242. Muhammad A, Pitchumoni CS. Newly detected celiac disease by wireless capsule endoscopy in older adults with iron deficiency anemia. J Clin Gastroenterol. 2008 Oct;42(9):980-3. PMID: 18596537.

243. Murray IA, Smith JA, Coupland K, et al. Intestinal disaccharidase deficiency without villous atrophy may represent early celiac disease. Scandinavian Journal of Gastroenterology. 2001 Feb;36(2):163-8. PMID: WOS:000166888500009.

244. Mustalahti K, Catassi C, Reunanen A, et al. The prevalence of celiac disease in Europe: results of a centralized, international mass screening project. Ann Med. 2010 Dec;42(8):587-95. PMID: 21070098.

245. Mustalahti K, Sulkanen S, Holopainen P, et al. Coeliac disease among healthy members of multiple case coeliac disease families. Scand J Gastroenterol. 2002 Feb;37(2):161-5. PMID: 11843051.

246. Myleus A, Ivarsson A, Webb C, et al. Celiac disease revealed in 3% of Swedish 12-yearolds born during an epidemic. J Pediatr Gastroenterol Nutr. 2009 Aug;49(2):170-6. PMID: 19516192.

247. Nachman F, Sugai E, Vazquez H, et al. Serological tests for celiac disease as indicators of long-term compliance with the gluten-free diet. Eur J Gastroenterol Hepatol. 2011 Jun;23(6):473-80. PMID: 21537123.

248. Nakazawa H, Makishima H, Ito T, et al. Screening tests using serum tissue transglutaminase IgA may facilitate the identification of undiagnosed celiac disease among Japanese population. Int J Med Sci. 2014;11(8):819-23. PMID: 24936145.

249. Narula P, Porter L, Langton J, et al. Gastrointestinal Symptoms in Children With Type 1 Diabetes Screened for Celiac Disease. Pediatrics. 2009 Sep;124(3):E489-E95. PMID: WOS:000269383100039.

250. Nastasio S, Sciveres M, Riva S, et al. Celiac disease-associated autoimmune hepatitis in childhood: long-term response to treatment. J Pediatr Gastroenterol Nutr. 2013 Jun;56(6):671-4. PMID: 23403438.

251. Nau AL, Fayad L, Lazzarotto C, et al. Prevalence and clinical features of celiac disease in patients with hepatitis B virus infection in Southern Brazil. Rev Soc Bras Med Trop. 2013 Jul-Aug;46(4):397-402. PMID: 23982094.

252. Nemec G, Ventura A, Stefano M, et al. Looking for celiac disease: diagnostic accuracy of two rapid commercial assays. Am J Gastroenterol. 2006 Jul;101(7):1597-600. PMID: 16863566.

253. Nenna R, Tiberti C, Petrarca L, et al. The celiac iceberg: characterization of the disease in primary schoolchildren. J Pediatr Gastroenterol Nutr. 2013 Apr;56(4):416-21. PMID: 23149808.

254. Nieto A, Blanco Quiros A, Arranz E, et al. Study of HLA-DQA1 alleles in celiac children. J Investig Allergol Clin Immunol. 1995 Jul-Aug;5(4):209-15. PMID: 8705011.

255. Norris JM, Barriga K, Hoffenberg EJ, et al. Risk of celiac disease autoimmunity and timing of gluten introduction in the diet of infants at increased risk of disease. JAMA. 2005 May 18;293(19):2343-51. PMID: 15900004.

256. Not T, Faleschini E, Tommasini A, et al. Celiac disease in patients with sporadic and inherited cardiomyopathies and in their relatives. Eur Heart J. 2003 Aug;24(15):1455-61. PMID: 12909075.

257. Not T, Tommasini A, Tonini G, et al. Undiagnosed coeliac disease and risk of autoimmune disorders in subjects with Type I diabetes mellitus. Diabetologia. 2001 Feb;44(2):151-5. PMID: 11270670.

258. O'Leary C, Feighery C, Feighery A, et al. The prevalence of coeliac disease among female subjects having bone densitometry. Ir J Med Sci. 2002 Jul-Sep;171(3):145-7. PMID: 15736353.

259. O'Leary C, Walsh CH, Wieneke P, et al. Coeliac disease and autoimmune Addison's disease: a clinical pitfall. QJM. 2002 Feb;95(2):79-82. PMID: 11861954.

260. Oliveira A, Trindade E, Tavares M, et al. Celiac disease in first degree relatives of celiac children. Arq Gastroenterol. 2012 Jul-Sep;49(3):204-7. PMID: 23011243.

261. Olszewska M, Sulej J, Kotowski B. Frequency and prognostic value of IgA and IgG endomysial antibodies in recurrent aphthous stomatitis. Acta Dermato-Venereologica. 2006;86(4):332-4. PMID: WOS:000239602200008.

262. Osmancevic L, Terzic S. Frequency of serological tests positive findings for celiac disease at the first relative of children with celiac disease. Med Arh. 2011;65(6):354-6. PMID: 22299298.

263. Ozaslan E, Akkorlu S, Eskioglu E, et al. Prevalence of silent celiac disease in patients with dyspepsia. Dig Dis Sci. 2007 Mar;52(3):692-7. PMID: 17235704.

264. Ozgor B, Selimoglu MA, Temel I, et al. Prevalence of celiac disease in parents of preterm or low birthweight newborns. J Obstet Gynaecol Res. 2011 Nov;37(11):1615-9. PMID: 21733039.

265. Paavola A, Kurppa K, Ukkola A, et al. Gastrointestinal symptoms and quality of life in screen-detected celiac disease. Dig Liver Dis. 2012 Oct;44(10):814-8. PMID: 22673312.

266. Pallav K, Kabbani T, Tariq S, et al. Clinical Utility of Celiac Disease-Associated HLA Testing. Dig Dis Sci. 2014 Apr 6PMID: 24705698.

267. Pallav K, Leffler DA, Tariq S, et al. Noncoeliac enteropathy: the differential diagnosis of villous atrophy in contemporary clinical practice. Aliment Pharmacol Ther. 2012 Feb;35(3):380-90. PMID: 22145590.

268. Parizade M, Shainberg B. Positive deamidated gliadin peptide antibodies and negative tissue transglutaminase IgA antibodies in a pediatric population: to biopsy or not to biopsy. Clin Vaccine Immunol. 2010 May;17(5):884-6. PMID: 20357057.

269. Pavlovic M, Radlovic N, Lekovic Z, et al. When to screen children with Down syndrome for celiac disease? J Trop Pediatr. 2010 Dec;56(6):443-5. PMID: 20388656.

270. Pellegrino S, Furfaro F, Tortora A, et al. The importance of disease prevalence in assessing the diagnostic value of a test: endoscopic markers in celiac disease. Digestion. 2013;87(4):254-61. PMID: 23751460.

271. Pena-Quintana L, Torres-Galvan MJ, Deniz-Naranjo MC, et al. Assessment of the DQ heterodimer test in the diagnosis of celiac disease in the Canary Islands (Spain). J Pediatr Gastroenterol Nutr. 2003 Nov;37(5):604-8. PMID: 14581805.

272. Pham-Short A, Donaghue KC, Ambler G, et al. Coeliac disease in Type 1 diabetes from 1990 to 2009: higher incidence in young children after longer diabetes duration. Diabet Med. 2012 Sep;29(9):e286-9. PMID: 22672045.

273. Picarelli A, Sabbatella L, Di Tola M, et al. Celiac disease diagnosis in misdiagnosed children. Pediatr Res. 2000 Nov;48(5):590-2. PMID: 11044476.

274. Picarelli A, Sabbatella L, Di Tola M, et al. Anti-endomysial antibody of IgG1 isotype detection strongly increases the prevalence of coeliac disease in patients affected by type I diabetes mellitus. Clin Exp Immunol. 2005 Oct;142(1):111-5. PMID: 16178863.

275. Piccini B, Vascotto M, Serracca L, et al. HLA-DQ typing in the diagnostic algorithm of celiac disease. Rev Esp Enferm Dig. 2012 May;104(5):248-54. PMID: 22662777.

276. Pittschieler K, Ladinser B. Coeliac disease: screened by a new strategy. Acta Paediatr Suppl. 1996 May;412:42-5. PMID: 8783755.

277. Poland DC, Ceelie H, Dinkelaar RB, et al. Determination of anti-endomysium IgA antibodies in the diagnosis of celiac disease: comparison of a novel ELISA-based assay with conventional immunofluorescence. World J Gastroenterol. 2006 May 7;12(17):2779-80. PMID: 16718769.

278. Popp A, Mihu M, Munteanu M, et al. Prospective antibody case finding of coeliac disease in type-1 diabetes children: need of biopsy revisited. Acta Paediatr. 2013 Mar;102(3):e102-6. PMID: 23211000.

279. Porcelli B, Ferretti F, Vindigni C, et al. Assessment of a Test for the Screening and Diagnosis of Celiac Disease. J Clin Lab Anal. 2014 Nov 10PMID: 25385391.

280. Poulain C, Johanet C, Delcroix C, et al. Prevalence and clinical features of celiac disease in 950 children with type 1 diabetes in France. Diabetes Metab. 2007 Dec;33(6):453-8. PMID: 17964843.

281. Prasad S, Thomas P, Nicholas DS, et al. Adult endomysial antibody-negative coeliac disease and cigarette smoking. Eur J Gastroenterol Hepatol. 2001 Jun;13(6):667-71. PMID: 11434592.

282. Pratesi R, Gandolfi L, Garcia SG, et al. Prevalence of coeliac disease: unexplained agerelated variation in the same population. Scand J Gastroenterol. 2003 Jul;38(7):747-50. PMID: 12889561.

283. Prince HE, Norman GL, Binder WL. Immunoglobulin A (IgA) deficiency and alternative celiac disease-associated antibodies in sera submitted to a reference laboratory for endomysial IgA testing. Clin Diagn Lab Immunol. 2000 Mar;7(2):192-6. PMID: 10702491.

284. Pueschel SM, Romano C, Failla P, et al. A prevalence study of celiac disease in persons with Down syndrome residing in the United States of America. Acta Paediatr. 1999 Sep;88(9):953-6. PMID: 10519335.

285. Queiroz MS, Nery M, Cancado EL, et al. Prevalence of celiac disease in Brazilian children of short stature. Braz J Med Biol Res. 2004 Jan;37(1):55-60. PMID: 14689044.

286. Rahmati A, Shakeri R, Sohrabi M, et al. Correlation of tissue transglutaminase antibody with duodenal histologic marsh grading. Middle East J Dig Dis. 2014 Jul;6(3):131-6. PMID: 25093060.

287. Ransford RA, Hayes M, Palmer M, et al. A controlled, prospective screening study of celiac disease presenting as iron deficiency anemia. J Clin Gastroenterol. 2002 Sep;35(3):228-33. PMID: 12192198.

288. Rashid M, MacDonald A. Importance of duodenal bulb biopsies in children for diagnosis of celiac disease in clinical practice. BMC Gastroenterol. 2009;9:78. PMID: 19835611.

289. Ravelli A, Villanacci V, Monfredini C, et al. How patchy is patchy villous atrophy?: distribution pattern of histological lesions in the duodenum of children with celiac disease. Am J Gastroenterol. 2010 Sep;105(9):2103-10. PMID: 20372112.

290. Ravikumara M, Nootigattu VK, Sandhu BK. Ninety percent of celiac disease is being missed. J Pediatr Gastroenterol Nutr. 2007 Oct;45(4):497-9. PMID: 18030224.

291. Rensch MJ, Szyjkowski R, Shaffer RT, et al. The prevalence of celiac disease autoantibodies in patients with systemic lupus erythematosus. Am J Gastroenterol. 2001 Apr;96(4):1113-5. PMID: 11316156.

292. Ress K, Harro J, Uibo O, et al. Use of a fully automated immunoassay for celiac disease screening in a pediatric population. Clin Chem Lab Med. 2011 Jun;49(6):983-7. PMID: 21428860.

293. Ribeiro-Cabral VL, da-Silva-Patricio FR, Ambrogini-Junior O, et al. Anti-tissue transglutaminase antibodies (IgA and IgG) in both Crohn s disease and autoimmune diabetes. Rev Esp Enferm Dig. 2011 Sep;103(9):453-7. PMID: 21951113.

294. Richter JC, Netzer P, Cottagnoud P, et al. Testing strategies and follow-up for coeliac disease in a general internal medicine outpatient department from 2000 to 2005 - A retrospective analysis and proposal for clinical practice. Swiss Medical Weekly. 2006 Nov;136(45-46):732-8. PMID: WOS:000243057600004.

295. Riestra S, Fernandez E, Rodrigo L, et al. Prevalence of Coeliac disease in the general population of northern Spain. Strategies of serologic screening. Scand J Gastroenterol. 2000 Apr;35(4):398-402. PMID: 10831263.

296. Robazzi TC, Adan LF, Pimentel K, et al. Autoimmune endocrine disorders and coeliac disease in children and adolescents with juvenile idiopathic arthritis and rheumatic fever. Clin Exp Rheumatol. 2013 Mar-Apr;31(2):310-7. PMID: 23406715.

297. Robson K, Alizart M, Martin J, et al. Coeliac patients are undiagnosed at routine upper endoscopy. PLoS One. 2014;9(3):e90552. PMID: 24595045.

298. Rodrigo L, Hernandez-Lahoz C, Fuentes D, et al. Prevalence of celiac disease in multiple sclerosis. BMC Neurol. 2011;11:31. PMID: 21385364.

299. Roka V, Potamianos SP, Kapsoritakis AN, et al. Prevalence of coeliac disease in the adult population of central Greece. Eur J Gastroenterol Hepatol. 2007 Nov;19(11):982-7. PMID: 18049168.

300. Roldan MB, Barrio R, Roy G, et al. Diagnostic value of serological markers for celiac disease in diabetic children and adolescents. J Pediatr Endocrinol Metab. 1998 Nov-Dec;11(6):751-6. PMID: 9829231.

301. Rosenberg NR, Vermeulen M. Should coeliac disease be considered in the work up of patients with chronic peripheral neuropathy? J Neurol Neurosurg Psychiatry. 2005 Oct;76(10):1415-9. PMID: 16170088.

302. Rostami K, Mulder CJ, van Overbeek FM, et al. Should relatives of coeliacs with mild clinical complaints undergo a small-bowel biopsy despite negative serology? Eur J Gastroenterol Hepatol. 2000 Jan;12(1):51-5. PMID: 10656210.

303. Rumbo M, Chirdo FG, Ben R, et al. Evaluation of coeliac disease serological markers in Down syndrome patients. Dig Liver Dis. 2002 Feb;34(2):116-21. PMID: 11926554.

304. Rutherford RM, Brutsche MH, Kearns M, et al. Prevalence of coeliac disease in patients with sarcoidosis. Eur J Gastroenterol Hepatol. 2004 Sep;16(9):911-5. PMID: 15316417.

305. Ruuskanen A, Kaukinen K, Collin P, et al. Positive serum antigliadin antibodies without celiac disease in the elderly population: does it matter? Scand J Gastroenterol. 2010 Oct;45(10):1197-202. PMID: 20545470.

306. Ruuskanen A, Luostarinen L, Collin P, et al. Persistently positive gliadin antibodies without transglutaminase antibodies in the elderly: gluten intolerance beyond coeliac disease. Dig Liver Dis. 2011 Oct;43(10):772-8. PMID: 21641886.

307. Saadah OI. Celiac disease in children and adolescents at a singe center in Saudi Arabia. Ann Saudi Med. 2011 Jan-Feb;31(1):51-7. PMID: 21245600.

308. Saberi-Firouzi M, Omrani GR, Nejabat M, et al. Prevalence of celiac disease in Shiraz, southern Iran. Saudi J Gastroenterol. 2008 Jul;14(3):135-8. PMID: 19568522.

309. Sachdev A, Srinivasan V, Maheswary S, et al. Adult onset celiac disease in north India. Trop Gastroenterol. 2002 Jul-Sep;23(3):117-9. PMID: 12693151.

310. Sahin I, Eminbeyli L, Andic S, et al. Screening for celiac disease among patients with chronic kidney disease. Ren Fail. 2012;34(5):545-9. PMID: 22563918.

311. Sakly W, Bienvenu F, Peretti N, et al. IgA anti-transglutaminase antibodies as a tool for screening atypical forms of coeliac disease in a French at-risk paediatric population. Eur J Gastroenterol Hepatol. 2005 Feb;17(2):235-9. PMID: 15674103.

312. Salardi S, Volta U, Zucchini S, et al. Prevalence of celiac disease in children with type 1 diabetes mellitus increased in the mid-1990 s: an 18-year longitudinal study based on antiendomysial antibodies. J Pediatr Gastroenterol Nutr. 2008 May;46(5):612-4. PMID: 18493223.

313. Salmi TT, Collin P, Korponay-Szabo IR, et al. Endomysial antibody-negative coeliac disease: clinical characteristics and intestinal autoantibody deposits. Gut. 2006 Dec;55(12):1746-53. PMID: 16571636.

314. Salmi TT, Collin P, Reunala T, et al. Diagnostic methods beyond conventional histology in coeliac disease diagnosis. Dig Liver Dis. 2010 Jan;42(1):28-32. PMID: 19473894.

315. Salvatore S, Finazzi S, Radaelli G, et al. Prevalence of undiagnosed celiac disease in the parents of preterm and/or small for gestational age infants. Am J Gastroenterol. 2007 Jan;102(1):168-73. PMID: 17100963.

316. Sanchez-Albisua I, Wolf J, Neu A, et al. Coeliac disease in children with Type 1 diabetes mellitus: the effect of the gluten-free diet. Diabet Med. 2005 Aug;22(8):1079-82. PMID: 16026376.

317. Sanders DS, Hopper AD, Azmy IA, et al. Association of adult celiac disease with surgical abdominal pain: a case-control study in patients referred to secondary care. Ann Surg. 2005 Aug;242(2):201-7. PMID: 16041210.

318. Sanders DS, Patel D, Khan FB, et al. Case-finding for adult celiac disease in patients with reduced bone mineral density. Dig Dis Sci. 2005 Mar;50(3):587-92. PMID: 15810647.

319. Sanders DS, Patel D, Stephenson TJ, et al. A primary care cross-sectional study of undiagnosed adult coeliac disease. Eur J Gastroenterol Hepatol. 2003 Apr;15(4):407-13. PMID: 12655262.

320. Sandstrom O, Rosen A, Lagerqvist C, et al. Transglutaminase IgA antibodies in a celiac disease mass screening and the role of HLA-DQ genotyping and endomysial antibodies in sequential testing. J Pediatr Gastroenterol Nutr. 2013 Oct;57(4):472-6. PMID: 23783015.

321. Santolaria S, Alcedo J, Cuartero B, et al. Spectrum of gluten-sensitive enteropathy in patients with dysmotility-like dyspepsia. Gastroenterol Hepatol. 2013 Jan;36(1):11-20. PMID: 23103052.

322. Sari S, Yesilkaya E, Egritas O, et al. Prevalence of Celiac disease in Turkish children with type 1 diabetes mellitus and their non-diabetic first-degree relatives. Turk J Gastroenterol. 2010 Mar;21(1):34-8. PMID: 20533110.

323. Sari S, Yesilkaya E, Egritas O, et al. Prevalence of celiac disease in Turkish children with autoimmune thyroiditis. Dig Dis Sci. 2009 Apr;54(4):830-2. PMID: 18716873.

324. Sategnaguidetti C, Grosso S, Pulitano R, et al. Celiac-Disease and Insulin-Dependent Diabetes-Mellitus - Screening in an Adult-Population. Digestive Diseases and Sciences. 1994 Aug;39(8):1633-7. PMID: WOS:A1994PA97900007.

325. Sattar N, Lazare F, Kacer M, et al. Celiac disease in children, adolescents, and young adults with autoimmune thyroid disease. J Pediatr. 2011 Feb;158(2):272-5 e1. PMID: 20961564.

326. Saukkonen T, Ilonen J, Akerblom HK, et al. Prevalence of coeliac disease in siblings of patients with Type I diabetes is related to the prevalence of DQB1*02 allele. Diabetologia. 2001 Aug;44(8):1051-3. PMID: 11484084.

327. Schirru E, Jores RD, Cicotto L, et al. High frequency of low-risk human leukocyte antigen class II genotypes in latent celiac disease. Hum Immunol. 2011 Feb;72(2):179-82. PMID: 21075156.

328. Sciberras C, Vella C, Grech V. The prevalence of coeliac disease in Down's syndrome in Malta. Ann Trop Paediatr. 2004 Mar;24(1):81-3. PMID: 15005971.

329. Seissler J, Boms S, Wohlrab U, et al. Antibodies to human recombinant tissue transglutaminase measured by radioligand assay: evidence for high diagnostic sensitivity for celiac disease. Horm Metab Res. 1999 Jun;31(6):375-9. PMID: 10437627.

330. Shah VH, Rotterdam H, Kotler DP, et al. All that scallops is not celiac disease. Gastrointest Endosc. 2000 Jun;51(6):717-20. PMID: 10840307.

331. Shahbazkhani B, Faezi T, Akbari MR, et al. Coeliac disease in Iranian type I diabetic patients. Dig Liver Dis. 2004 Mar;36(3):191-4. PMID: 15046188.

332. Shahbazkhani B, Mohamadnejad M, Malekzadeh R, et al. Coeliac disease is the most common cause of chronic diarrhoea in Iran. Eur J Gastroenterol Hepatol. 2004 Jul;16(7):665-8. PMID: 15201579.

333. Shamaly H, Hartman C, Pollack S, et al. Tissue transglutaminase antibodies are a useful serological marker for the diagnosis of celiac disease in patients with Down syndrome. J Pediatr Gastroenterol Nutr. 2007 May;44(5):583-6. PMID: 17460490.

334. Shamir R, Lerner A, Shinar E, et al. The use of a single serological marker underestimates the prevalence of celiac disease in Israel: a study of blood donors. Am J Gastroenterol. 2002 Oct;97(10):2589-94. PMID: 12385444.

335. Sharifi N, Khoshbaten M, Aliasgarzade A, et al. Celiac disease in patients with type-1 diabetes mellitus screened by tissue transglutaminase antibodies in northwest of Iran. Int J Diabetes Dev Ctries. 2008 Jul;28(3):95-9. PMID: 19902043.

336. Sherwani RK, Alam S, Akhtar K, et al. Prevalence of iron deficiency anemia in chronic diarrhoea and celiac disease - a western UP experience. Indian J Hematol Blood Transfus. 2008 Mar;24(1):12-5. PMID: 23100934.

337. Simell S, Hoppu S, Hekkala A, et al. Fate of five celiac disease-associated antibodies during normal diet in genetically at-risk children observed from birth in a natural history study. Am J Gastroenterol. 2007 Sep;102(9):2026-35. PMID: 17573785.

338. Simell S, Hoppu S, Simell T, et al. Age at development of type 1 diabetes- and celiac disease-associated antibodies and clinical disease in genetically susceptible children observed from birth. Diabetes Care. 2010 Apr;33(4):774-9. PMID: 20056952.

339. Simell S, Kupila A, Hoppu S, et al. Natural history of transglutaminase autoantibodies and mucosal changes in children carrying HLA-conferred celiac disease susceptibility. Scand J Gastroenterol. 2005 Oct;40(10):1182-91. PMID: 16265775.

340. Simmons JH, Klingensmith GJ, McFann K, et al. Celiac autoimmunity in children with type 1 diabetes: a two-year follow-up. J Pediatr. 2011 Feb;158(2):276-81 e1. PMID: 20817171.

341. Singh P, Wadhwa N, Chaturvedi MK, et al. Validation of point-of-care testing for coeliac disease in children in a tertiary hospital in north India. Arch Dis Child. 2014 Nov;99(11):1004-8. PMID: 24942708.

342. Sinha SK, Nain CK, Udawat HP, et al. Cervical esophageal web and celiac disease. J Gastroenterol Hepatol. 2008 Jul;23(7 Pt 1):1149-52. PMID: 18554241.

343. Sjoberg K, Eriksson KF, Bredberg A, et al. Screening for coeliac disease in adult insulindependent diabetes mellitus. J Intern Med. 1998 Feb;243(2):133-40. PMID: 9566642.

344. Sjoberg K, Eriksson S. Regional differences in coeliac disease prevalence in Scandinavia? Scand J Gastroenterol. 1999 Jan;34(1):41-5. PMID: 10048731.

345. Smecuol E, Vazquez H, Sugai E, et al. Sugar tests detect celiac disease among first-degree relatives. Am J Gastroenterol. 1999 Dec;94(12):3547-52. PMID: 10606317.

346. Smith CM, Clarke CF, Porteous LE, et al. Prevalence of coeliac disease and longitudinal follow-up of antigliadin antibody status in children and adolescents with type 1 diabetes mellitus. Pediatr Diabetes. 2000 Dec;1(4):199-203. PMID: 15016216.

347. Soresi M, Amplo M, Agliastro R, et al. Screening for autoantibodies to tissue transglutaminase reveals a low prevalence of celiac disease in blood donors with cryptogenic hypertransaminasemia. Digestion. 2001;64(2):87-91. PMID: 11684821.

348. Spadaccino AC, Basso D, Chiarelli S, et al. Celiac disease in North Italian patients with autoimmune thyroid diseases. Autoimmunity. 2008 Feb;41(1):116-21. PMID: 18176874.

349. Spiekerkoetter U, Seissler J, Wendel U. General screening for celiac disease is advisable in children with type 1 diabetes. Horm Metab Res. 2002 Apr;34(4):192-5. PMID: 11987028.

350. Srivastava A, Yachha SK, Mathias A, et al. Prevalence, human leukocyte antigen typing and strategy for screening among Asian first-degree relatives of children with celiac disease. J Gastroenterol Hepatol. 2010 Feb;25(2):319-24. PMID: 19929927.

351. Stagi S, Manoni C, Cecchi C, et al. Increased risk of coeliac disease in patients with congenital hypothyroidism. Horm Res Paediatr. 2011;76(3):186-92. PMID: 21757873.

352. Stene LC, Honeyman MC, Hoffenberg EJ, et al. Rotavirus infection frequency and risk of celiac disease autoimmunity in early childhood: a longitudinal study. Am J Gastroenterol. 2006 Oct;101(10):2333-40. PMID: 17032199.

353. Sumnik Z, Cinek O, Bratanic N, et al. Thyroid autoimmunity in children with coexisting type 1 diabetes mellitus and celiac disease: a multicenter study. J Pediatr Endocrinol Metab. 2006 Apr;19(4):517-22. PMID: 16759037.

354. Sun S, Puttha R, Ghezaiel S, et al. The effect of biopsy-positive silent coeliac disease and treatment with a gluten-free diet on growth and glycaemic control in children with Type 1 diabetes. Diabetic Medicine. 2009 Dec;26(12):1250-4. PMID: WOS:000272161900010.

355. Sundar N, Crimmins R, Swift G. Clinical presentation and incidence of complications in patients with coeliac disease diagnosed by relative screening. Postgrad Med J. 2007 Apr;83(978):273-6. PMID: 17403956.

356. Sweis R, Pee L, Smith-Laing G. Discrepancies between histology and serology for the diagnosis of coeliac disease in a district general hospital: is this an unrecognised problem in other hospitals? Clinical Medicine. 2009 Aug;9(4):346-8. PMID: WOS:000269025000013.

357. Szalowska-Wozniak DA, Bak-Romaniszyn L, Cywinska-Bernas A, et al. Evaluation of HLA-DQ2/DQ8 genotype in patients with celiac disease hospitalised in 2012 at the Department of Paediatrics. Prz Gastroenterol. 2014;9(1):32-7. PMID: 24868296.

358. Talal AH, Murray JA, Goeken JA, et al. Celiac disease in an adult population with insulindependent diabetes mellitus: use of endomysial antibody testing. Am J Gastroenterol. 1997 Aug;92(8):1280-4. PMID: 9260789.

359. Tarmure S, Grigorescu M, Cristea A, et al. Antiendomysial and antitissue transglutaminase antibodies in gluten-induced enteropathy. Rom J Gastroenterol. 2002 Jun;11(2):91-5. PMID: 12145663.

360. Tatar G, Elsurer R, Simsek H, et al. Screening of tissue transglutaminase antibody in healthy blood donors for celiac disease screening in the Turkish population. Dig Dis Sci. 2004 Sep;49(9):1479-84. PMID: 15481323.

361. Tesija-Kuna A, Topic E, Zizic V, et al. Antiendomysial and antigliadin antibodies in the diagnosis of celiac disease in children of short stature. Periodicum Biologorum. 2005 Jun;107(2):235-8. PMID: WOS:000230675700017.

362. Thapa BR, Rawal P, Sapra B, et al. Familial prevalence of celiac disease. J Trop Pediatr. 2011 Feb;57(1):45-50. PMID: 20554512.

363. Tiboni GM, de Vita MG, Faricelli R, et al. Serological testing for celiac disease in women undergoing assisted reproduction techniques. Hum Reprod. 2006 Feb;21(2):376-9. PMID: 16172142.

364. Tikkakoski S, Savilahti E, Kolho KL. Undiagnosed coeliac disease and nutritional deficiencies in adults screened in primary health care. Scand J Gastroenterol. 2007 Jan;42(1):60-5. PMID: 17190764.

365. Tjellstrom B, Stenhammar L, Hogberg L, et al. Screening-detected and symptomatic untreated celiac children show similar gut microflora-associated characteristics. Scand J Gastroenterol. 2010 Sep;45(9):1059-62. PMID: 20509753.

366. Toftedal P, Nielsen C, Madsen JT, et al. Positive predictive value of serological diagnostic measures in celiac disease. Clin Chem Lab Med. 2010 May;48(5):685-91. PMID: 20201743.

367. Tonutti E, Visentini D, Picierno A, et al. Diagnostic efficacy of the ELISA test for the detection of deamidated anti-gliadin peptide antibodies in the diagnosis and monitoring of celiac disease. J Clin Lab Anal. 2009;23(3):165-71. PMID: 19455636.

368. Tortora R, Imperatore N, Capone P, et al. The presence of anti-endomysial antibodies and the level of anti-tissue transglutaminases can be used to diagnose adult coeliac disease without duodenal biopsy. Aliment Pharmacol Ther. 2014 Nov;40(10):1223-9. PMID: 25263177.

369. Tosco A, Auricchio R, Aitoro R, et al. Intestinal titres of anti-tissue transglutaminase 2 antibodies correlate positively with mucosal damage degree and inversely with gluten-free diet duration in coeliac disease. Clin Exp Immunol. 2014 Sep;177(3):611-7. PMID: 24773630.

370. Trevisiol C, Not T, Berti I, et al. Screening for coeliac disease in healthy blood donors at two immuno-transfusion centres in north-east Italy. Ital J Gastroenterol Hepatol. 1999 Oct;31(7):584-6. PMID: 10604097.

371. Trevisiol C, Ventura A, Baldas V, et al. A reliable screening procedure for coeliac disease in clinical practice. Scand J Gastroenterol. 2002 Jun;37(6):679-84. PMID: 12126246.

372. Trigoni E, Tsirogianni A, Pipi E, et al. Celiac disease in adult patients: specific autoantibodies in the diagnosis, monitoring, and screening. Autoimmune Dis. 2014;2014:623514. PMID: 24804083.

373. Tumer L, Hasanoglu A, Aybay C. Endomysium antibodies in the diagnosis of celiac disease in short-statured children with no gastrointestinal symptoms. Pediatr Int. 2001 Feb;43(1):71-3. PMID: 11208004.

374. Tursi A, Giorgetti GM, Brandimarte G, et al. High prevalence of celiac disease among patients affected by Crohn's disease. Inflamm Bowel Dis. 2005 Jul;11(7):662-6. PMID: 15973121.

375. Uenishi RH, Gandolfi L, Almeida LM, et al. Screening for celiac disease in 1st degree relatives: a 10-year follow-up study. BMC Gastroenterol. 2014;14:36. PMID: 24552206.

376. Uibo O, Heilman K, Rago T, et al. Symptomless celiac disease in type 1 diabetes: 12-year experience in Estonia. Pediatr Int. 2010 Apr;52(2):230-3. PMID: 19744227.

377. Unsworth DJ, Brown DL. Serological screening suggests that adult coeliac disease is underdiagnosed in the UK and increases the incidence by up to 12%. Gut. 1994 Jan;35(1):61-4. PMID: 8307451.

378. Valentino R, Savastano S, Tommaselli AP, et al. Prevalence of coeliac disease in patients with thyroid autoimmunity. Horm Res. 1999;51(3):124-7. PMID: 10461017.

379. Valerio G, Maiuri L, Troncone R, et al. Severe clinical onset of diabetes and increased prevalence of other autoimmune diseases in children with coeliac disease diagnosed before diabetes mellitus. Diabetologia. 2002 Dec;45(12):1719-22. PMID: 12488963.

380. Vallejo-Diez S, Bernardo D, Moreno Mde L, et al. Detection of specific IgA antibodies against a novel deamidated 8-Mer gliadin peptide in blood plasma samples from celiac patients. PLoS One. 2013;8(11):e80982. PMID: 24278359.

381. Valletta E, Ulmi D, Mabboni I, et al. Early diagnosis and treatment of celiac disease in type 1 diabetes. A longitudinal, case-control study. Pediatr Med Chir. 2007 Mar-Apr;29(2):99-104. PMID: 17461097.

382. van Koppen EJ, Schweizer JJ, Csizmadia CG, et al. Long-term health and quality-of-life consequences of mass screening for childhood celiac disease: a 10-year follow-up study. Pediatrics. 2009 Apr;123(4):e582-8. PMID: 19336349.

383. Vannella L, Gianni D, Lahner E, et al. Pre-endoscopic screening for Helicobacter pylori and celiac disease in young anemic women. World J Gastroenterol. 2009 Jun 14;15(22):2748-53. PMID: 19522025.

384. Vazquez H, Cabanne A, Sugai E, et al. Serological markers identify histologically latent coeliac disease among first-degree relatives. Eur J Gastroenterol Hepatol. 1996 Jan;8(1):15-21. PMID: 8900904.

385. Vecsei A, Arenz T, Heilig G, et al. Influence of age and genetic risk on anti-tissue transglutaminase IgA titers. J Pediatr Gastroenterol Nutr. 2009 May;48(5):544-9. PMID: 19367177.

386. Venkatasubramani N, Telega G, Werlin SL. Obesity in pediatric celiac disease. J Pediatr Gastroenterol Nutr. 2010 Sep;51(3):295-7. PMID: 20479683.

387. Vermeersch P, Geboes K, Marien G, et al. Defining thresholds of antibody levels improves diagnosis of celiac disease. Clin Gastroenterol Hepatol. 2013 Apr;11(4):398-403; quiz e32. PMID: 23103822.

388. Villalta D, Alessio MG, Tampoia M, et al. Testing for IgG class antibodies in celiac disease patients with selective IgA deficiency. A comparison of the diagnostic accuracy of 9 IgG anti-tissue transglutaminase, 1 IgG anti-gliadin and 1 IgG anti-deaminated gliadin peptide antibody assays. Clin Chim Acta. 2007 Jul;382(1-2):95-9. PMID: 17490629.

389. Vilppula A, Collin P, Maki M, et al. Undetected coeliac disease in the elderly: a biopsyproven population-based study. Dig Liver Dis. 2008 Oct;40(10):809-13. PMID: 18467196.

390. Vivas S, Ruiz de Morales JM, Martinez J, et al. Human recombinant anti-transglutaminase antibody testing is useful in the diagnosis of silent coeliac disease in a selected group of at-risk patients. Eur J Gastroenterol Hepatol. 2003 May;15(5):479-83. PMID: 12702903.

391. Vives MJ, Esteve M, Marine M, et al. Prevalence and clinical relevance of enteropathy associated with systemic autoimmune diseases. Dig Liver Dis. 2012 Aug;44(8):636-42. PMID: 22465228.

392. Volta U, Granito A, De Franceschi L, et al. Anti tissue transglutaminase antibodies as predictors of silent coeliac disease in patients with hypertransaminasaemia of unknown origin. Dig Liver Dis. 2001 Jun-Jul;33(5):420-5. PMID: 11529654.

393. Volta U, Ravaglia G, Granito A, et al. Coeliac disease in patients with autoimmune thyroiditis. Digestion. 2001;64(1):61-5. PMID: 11549838.

394. Volta U, Rodrigo L, Granito A, et al. Celiac disease in autoimmune cholestatic liver disorders. American Journal of Gastroenterology. 2002 Oct;97(10):2609-13. PMID: WOS:000178504800022.

395. Waisbourd-Zinman O, Hojsak I, Rosenbach Y, et al. Spontaneous normalization of antitissue transglutaminase antibody levels is common in children with type 1 diabetes mellitus. Dig Dis Sci. 2012 May;57(5):1314-20. PMID: 22173747.

396. Wakim-Fleming J, Pagadala MR, Lemyre MS, et al. Diagnosis of celiac disease in adults based on serology test results, without small-bowel biopsy. Clin Gastroenterol Hepatol. 2013 May;11(5):511-6. PMID: 23305824.

397. Walker MM, Murray JA, Ronkainen J, et al. Detection of celiac disease and lymphocytic enteropathy by parallel serology and histopathology in a population-based study. Gastroenterology. 2010 Jul;139(1):112-9. PMID: 20398668.

398. Walkowiak J, Blask-Osipa A, Lisowska A, et al. Cystic fibrosis is a risk factor for celiac disease. Acta Biochim Pol. 2010;57(1):115-8. PMID: 20300660.

399. Wang N, Truedsson L, Elvin K, et al. Serological assessment for celiac disease in IgA deficient adults. PLoS One. 2014;9(4):e93180. PMID: 24709954.

400. Webb C, Halvarsson B, Norstrom F, et al. Accuracy in celiac disease diagnostics by controlling the small-bowel biopsy process. J Pediatr Gastroenterol Nutr. 2011 May;52(5):549-53. PMID: 21502825.

401. Weiss B, Bujanover Y, Avidan B, et al. Positive tissue transglutaminase antibodies with negative endomysial antibodies: low rate of celiac disease. Isr Med Assoc J. 2004 Jan;6(1):9-12. PMID: 14740501.

402. Wengrower D, Doron D, Goldin E, et al. Should stored serum of patients previously tested for celiac disease serology be retested for transglutaminase antibodies? J Clin Gastroenterol. 2006 Oct;40(9):806-8. PMID: 17016136.

403. Wiland HOt, Henricks WH, Daly TM. Limited utilization of serologic testing in patients undergoing duodenal biopsy for celiac disease. BMC Gastroenterol. 2013;13(1):156. PMID: 24209459.

404. Wolters VM, van de Nadort C, Gerritsen SA, et al. Is gluten challenge really necessary for the diagnosis of coeliac disease in children younger than age 2 years? J Pediatr Gastroenterol Nutr. 2009 May;48(5):566-70. PMID: 19367182.

405. Wouters J, Weijerman ME, van Furth AM, et al. Prospective human leukocyte antigen, endomysium immunoglobulin A antibodies, and transglutaminase antibodies testing for celiac disease in children with Down syndrome. J Pediatr. 2009 Feb;154(2):239-42. PMID: 18822429.

406. Yagil Y, Goldenberg I, Arnon R, et al. Serologic testing for celiac disease in young adults-a cost-effect analysis. Dig Dis Sci. 2005 Apr;50(4):796-805. PMID: 15844721.

407. Zachor DA, Mroczek-Musulman E, Brown P. Prevalence of celiac disease in Down syndrome in the United States. J Pediatr Gastroenterol Nutr. 2000 Sep;31(3):275-9. PMID: 10997372.

408. Zamani M, Modares-Sadegi M, Shirvani F, et al. The involvement of the HLA-DQB1 alleles in the risk and the severity of Iranian coeliac disease patients. Int J Immunogenet. 2014 Aug;41(4):312-7. PMID: 24917237.

409. Zauli D, Grassi A, Granito A, et al. Prevalence of silent coeliac disease in atopics. Dig Liver Dis. 2000 Dec;32(9):775-9. PMID: 11215557.

410. Zhang F, Yang B, Lin Y, et al. Dermatitis herpetiformis in China: a report of 22 cases. Journal of the European Academy of Dermatology and Venereology. 2012 Jul;26(7):903-7. PMID: WOS:000305573600018.

No or Unclear on Consecutive or Random Sample - N=43

1. Barbato M, Maiella G, Di Camillo C, et al. The anti-deamidated gliadin peptide antibodies unmask celiac disease in small children with chronic diarrhoea. Dig Liver Dis. 2011 Jun;43(6):465-9. PMID: 21257356.

2. Basso D, Gallo N, Guariso G, et al. Role of anti-transglutaminase (anti-tTG), anti-gliadin, and anti-endomysium serum antibodies in diagnosing celiac disease: a comparison of four different commercial kits for anti-tTG determination. J Clin Lab Anal. 2001;15(3):112-5. PMID: 11344524.

3. Biagi F, Rondonotti E, Campanella J, et al. Video capsule endoscopy and histology for smallbowel mucosa evaluation: a comparison performed by blinded observers. Clin Gastroenterol Hepatol. 2006 Aug;4(8):998-1003. PMID: 16814612.

4. Biagi F, Schiepatti A, Malamut G, et al. PROgnosticating COeliac patieNts SUrvivaL: the PROCONSUL score. PLoS One. 2014;9(1):e84163. PMID: 24392112.

5. Burgin-Wolff A, Mauro B, Faruk H. Intestinal biopsy is not always required to diagnose celiac disease: a retrospective analysis of combined antibody tests. BMC Gastroenterol. 2013;13:19. PMID: 23343249.

6. Clouzeau-Girard H, Rebouissoux L, Taupin JL, et al. HLA-DQ genotyping combined with serological markers for the diagnosis of celiac disease: is intestinal biopsy still mandatory? J Pediatr Gastroenterol Nutr. 2011 Jun;52(6):729-33. PMID: 21593645.

7. Di Leo M, Weisz G, Ansaldi Balocco N. Serum and salivary antiendomysium antibodies in the screening of coeliac disease. Panminerva Med. 1999 Mar;41(1):68-71. PMID: 10230262.

8. Donaldson MR, Firth SD, Wimpee H, et al. Correlation of duodenal histology with tissue transglutaminase and endomysial antibody levels in pediatric celiac disease. Clin Gastroenterol Hepatol. 2007 May;5(5):567-73. PMID: 17428743.

9. Eremic N, Deric M, Hadnadev L. Diagnostic Accuracy of Iga Anti-Tissue Transglutaminase Antibody Testing in Celiac Disease. Journal of Medical Biochemistry. 2012 Apr-Jun;31(2):100-6. PMID: WOS:000300224400004.

10. Evans KE, Leeds JS, Morley S, et al. Pancreatic insufficiency in adult celiac disease: do patients require long-term enzyme supplementation? Dig Dis Sci. 2010 Oct;55(10):2999-3004. PMID: 20458623.

11. Fayed SB, Aref MI, Fathy HM, et al. Prevalence of celiac disease, Helicobacter pylori and gastroesophageal reflux in patients with refractory iron deficiency anemia. J Trop Pediatr. 2008 Feb;54(1):43-53. PMID: 17908698.

12. Hansson T, Dahlbom I, Rogberg S, et al. Recombinant human tissue transglutaminase for diagnosis and follow-up of childhood coeliac disease. Pediatr Res. 2002 Jun;51(6):700-5. PMID: 12032264.

13. Hope BC, Ameratunga R, Austin PM, et al. Diagnostic utility of modified gliadin peptide antibody assays in New Zealand children. J Pediatr Gastroenterol Nutr. 2013 Jul;57(1):43-8. PMID: 23403444.

14. Hopper AD, Hurlstone DP, Leeds JS, et al. The occurrence of terminal ileal histological abnormalities in patients with coeliac disease. Dig Liver Dis. 2006 Nov;38(11):815-9. PMID: 16787773.

15. Jatla M, Bokhari A, Bierly P, et al. Anthropometric, serologic, and laboratory correlation with villous blunting in pediatric celiac disease: diabetics are different. J Clin Gastroenterol. 2009 Aug;43(7):622-6. PMID: 19238095.

16. Kotze LM, Utiyama SR, Nisihara RM, et al. IgA class anti-endomysial and anti-tissue transglutaminase antibodies in relation to duodenal mucosa changes in coeliac disease. Pathology. 2003 Feb;35(1):56-60. PMID: 12701686.

17. Kurppa K, Lindfors K, Collin P, et al. Antibodies against deamidated gliadin peptides in early-stage celiac disease. J Clin Gastroenterol. 2011 Sep;45(8):673-8. PMID: 21063208.

18. Lammi A, Arikoski P, Simell S, et al. Antibodies to Deamidated Gliadin Peptide in Diagnosis of Celiac Disease in Children. J Pediatr Gastroenterol Nutr. 2014 Dec 16PMID: 25522308.

19. Li M, Yu LP, Tiberti C, et al. A Report on the International Transglutaminase Autoantibody Workshop for Celiac Disease. American Journal of Gastroenterology. 2009 Jan;104(1):154-63. PMID: WOS:000262265800026.

20. Lidums I, Teo E, Field J, et al. Capsule endoscopy: a valuable tool in the follow-up of people with celiac disease on a gluten-free diet. Clin Transl Gastroenterol. 2011;2:e4. PMID: 23237971.

21. Lindfors K, Koskinen O, Kurppa K, et al. Serodiagnostic assays for celiac disease based on the open or closed conformation of the autoantigen, transglutaminase 2. J Clin Immunol. 2011 Jun;31(3):436-42. PMID: 21384250.

22. Maglio M, Tosco A, Paparo F, et al. Serum and intestinal celiac disease-associated antibodies in children with celiac disease younger than 2 years of age. J Pediatr Gastroenterol Nutr. 2010 Jan;50(1):43-8. PMID: 19934769.

23. Maiden L, Elliott T, McLaughlin SD, et al. A blinded pilot comparison of capsule endoscopy and small bowel histology in unresponsive celiac disease. Dig Dis Sci. 2009 Jun;54(6):1280-3. PMID: 18975089.

24. Mankai A, Sakly W, Landolsi H, et al. Tissue transglutaminase antibodies in celiac disease, comparison of an enzyme linked immunosorbent assay and a dot blot assay. Pathol Biol (Paris). 2005 May;53(4):204-9. PMID: 15850953.

25. McMillan SA, Dickey W, Douglas JP, et al. Transthyretin values correlate with mucosal recovery in patients with coeliac disease taking a gluten free diet. J Clin Pathol. 2001 Oct;54(10):783-6. PMID: 11577127.

26. Mubarak A, Spierings E, Wolters VM, et al. Children with celiac disease and high tTGA are genetically and phenotypically different. World J Gastroenterol. 2013 Nov 7;19(41):7114-20. PMID: 24222955.

27. Panetta F, Torre G, Colistro F, et al. Clinical accuracy of anti-tissue transglutaminase as screening test for celiac disease under 2 years. Acta Paediatr. 2011 May;100(5):728-31. PMID: 21166861.

28. Raivio T, Korponay-Szabo I, Collin P, et al. Performance of a new rapid whole blood coeliac test in adult patients with low prevalence of endomysial antibodies. Dig Liver Dis. 2007 Dec;39(12):1057-63. PMID: 17983878.

29. Rostami Nejad M, Rostami K, Yamaoka Y, et al. Clinical and histological presentation of Helicobacter pylori and gluten related gastroenteropathy. Arch Iran Med. 2011 Mar;14(2):115-8. PMID: 21361718.

30. Rujner J, Socha J, Barra E, et al. Serum and salivary antigliadin antibodies and serum IgA anti-endomysium antibodies as a screening test for coeliac disease. Acta Paediatr. 1996 Jul;85(7):814-7. PMID: 8819547.

31. Saneian H, Gorgani AM. Diagnostic value of serologic tests in celiac screening. Int J Prev Med. 2012 Mar;3(Suppl 1):S58-63. PMID: 22826771.

32. Sardy M, Odenthal U, Karpati S, et al. Recombinant human tissue transglutaminase ELISA for the diagnosis of gluten-sensitive enteropathy. Clin Chem. 1999 Dec;45(12):2142-9. PMID: 10585346.

33. Sategna-Guidetti C, Grosso S, Bruno M, et al. Reliability of immunologic markers of celiac sprue in the assessment of mucosal recovery after gluten withdrawal. J Clin Gastroenterol. 1996 Sep;23(2):101-4. PMID: 8877634.

34. Spatola BN, Kaukinen K, Collin P, et al. Persistence of elevated deamidated gliadin peptide antibodies on a gluten-free diet indicates nonresponsive coeliac disease. Aliment Pharmacol Ther. 2014 Feb;39(4):407-17. PMID: 24392888.

35. Tanpowpong P, Broder-Fingert S, Katz AJ, et al. Characteristics of children with positive coeliac serology and normal villous morphology: potential coeliac disease. APMIS. 2013 Apr;121(4):266-71. PMID: 23030455.

36. Tarmure S, Cristea A, Sampelean D, et al. Serological and histological correlations in celiac disease. Rom J Intern Med. 2007;45(3):263-8. PMID: 18333359.

37. Tonutti E, Visentini D, Bizzaro N, et al. The role of antitissue transglutaminase assay for the diagnosis and monitoring of coeliac disease: a French-Italian multicentre study. J Clin Pathol. 2003 May;56(5):389-93. PMID: 12719462.

38. Tosco A, Auricchio R, Aitoro R, et al. In celiac disease intestinal titers of anti-tissue transglutaminase2 antibodies positively correlate with the mucosal damage degree and inversely with the gluten-free diet duration. Clin Exp Immunol. 2014 Apr 28PMID: 24773630.

39. Tosco A, Salvati VM, Auricchio R, et al. Natural history of potential celiac disease in children. Clin Gastroenterol Hepatol. 2011 Apr;9(4):320-5; quiz e36. PMID: 20851213.

40. Venhoff N, Emmerich F, Neagu M, et al. The role of HLA DQ2 and DQ8 in dissecting celiac-like disease in common variable immunodeficiency. J Clin Immunol. 2013 Jul;33(5):909-16. PMID: 23609110.

41. Vivas S, Ruiz de Morales JG, Riestra S, et al. Duodenal biopsy may be avoided when high transglutaminase antibody titers are present. World J Gastroenterol. 2009 Oct 14;15(38):4775-80. PMID: 19824110.

42. Zanini B, Caselani F, Magni A, et al. Celiac disease with mild enteropathy is not mild disease. Clin Gastroenterol Hepatol. 2013 Mar;11(3):253-8. PMID: 23022697.

43. Zanini B, Lanzarotto F, Mora A, et al. Five year time course of celiac disease serology during gluten free diet: results of a community based "CD-Watch" program. Dig Liver Dis. 2010 Dec;42(12):865-70. PMID: 20598661.

Sample Size Less Than 300 and Not a Special Population - N=83

1. Abrams JA, Diamond B, Rotterdam H, et al. Seronegative celiac disease: increased prevalence with lesser degrees of villous atrophy. Dig Dis Sci. 2004 Apr;49(4):546-50. PMID: 15185855.

2. Adriaanse MP, Tack GJ, Passos VL, et al. Serum I-FABP as marker for enterocyte damage in coeliac disease and its relation to villous atrophy and circulating autoantibodies. Aliment Pharmacol Ther. 2013 Feb;37(4):482-90. PMID: 23289539.

3. Aleksandra B, Ivana K, Ivica S, et al. Profile of Typical and Atypical Celiac Disease in Serbian Children. Indian Pediatrics. 2013 Nov;50(11):1061-2. PMID: WOS:000327082600018.

4. Alonso-Llamazares J, Gibson LE, Rogers RS, 3rd. Clinical, pathologic, and immunopathologic features of dermatitis herpetiformis: review of the Mayo Clinic experience. Int J Dermatol. 2007 Sep;46(9):910-9. PMID: 17822491.

5. Arevalo F, Roe E, Arias-Stella-Castillo J, et al. Low serological positivity in patients with histology compatible with celiac disease in Peru. Rev Esp Enferm Dig. 2010 Jun;102(6):372-5. PMID: 20575597.

6. Ashorn S, Valineva T, Kaukinen K, et al. Serological responses to microbial antigens in celiac disease patients during a gluten-free diet. J Clin Immunol. 2009 Mar;29(2):190-5. PMID: 18987962.

7. Auricchio R, Tosco A, Piccolo E, et al. Potential celiac children: 9-year follow-up on a gluten-containing diet. Am J Gastroenterol. 2014 Jun;109(6):913-21. PMID: 24777149.

8. Aydemir S, Tekin NS, Aktunc E, et al. Celiac disease in patients having recurrent aphthous stomatitis. Turk J Gastroenterol. 2004 Sep;15(3):192-5. PMID: 15492921.

9. Aziz I, Key T, Goodwin JG, et al. Predictors for Celiac Disease in Adult Cases of Duodenal Intraepithelial Lymphocytosis. J Clin Gastroenterol. 2014 Jul 10PMID: 25014240.

10. Balamtekin N, Uslu N, Baysoy G, et al. The presentation of celiac disease in 220 Turkish children. Turk J Pediatr. 2010 May-Jun;52(3):239-44. PMID: 20718180.

11. Basso D, Guariso G, Fogar P, et al. Antibodies against synthetic deamidated gliadin peptides for celiac disease diagnosis and follow-up in children. Clin Chem. 2009 Jan;55(1):150-7. PMID: 18988751.

12. Baviera LC, Aliaga ED, Ortigosa L, et al. Celiac disease screening by immunochromatographic visual assays: results of a multicenter study. J Pediatr Gastroenterol Nutr. 2007 Nov;45(5):546-50. PMID: 18030231.

13. Beltran L, Koenig M, Egner W, et al. High-titre circulating TTG2 antibodies predict small bowel villous atrophy, but decision cut-off limits must be locally validated. Clin Exp Immunol. 2013 Dec 11PMID: 24325651.

14. Beltran L, Koenig M, Egner W, et al. High-titre circulating tissue transglutaminase-2 antibodies predict small bowel villous atrophy, but decision cut-off limits must be locally validated. Clin Exp Immunol. 2014 May;176(2):190-8. PMID: 24325651.

15. Biagi F, Bianchi PI, Campanella J, et al. The prevalence and the causes of minimal intestinal lesions in patients complaining of symptoms suggestive of enteropathy: a follow-up study. J Clin Pathol. 2008 Oct;61(10):1116-8. PMID: 18708422.

16. Boger CP, Thomas PW, Nicholas DS, et al. Determinants of endomysial antibody status in untreated coeliac disease. Eur J Gastroenterol Hepatol. 2007 Oct;19(10):890-5. PMID: 17873614.

17. Borrelli M, Maglio M, Agnese M, et al. High density of intraepithelial gammadelta lymphocytes and deposits of immunoglobulin (Ig)M anti-tissue transglutaminase antibodies in the jejunum of coeliac patients with IgA deficiency. Clin Exp Immunol. 2010 May;160(2):199-206. PMID: 20030673.

18. Brusca I, Carroccio A, Tonutti E, et al. The old and new tests for celiac disease: which is the best test combination to diagnose celiac disease in pediatric patients? Clin Chem Lab Med. 2012 Jan;50(1):111-7. PMID: 21942854.

19. Butterworth JR, Iqbal TH, Cooper BT. Coeliac disease in South Asians resident in Britain: comparison with white Caucasian coeliac patients. Eur J Gastroenterol Hepatol. 2005 May;17(5):541-5. PMID: 15827445.

20. Carroccio A, Di Prima L, Pirrone G, et al. Anti-transglutaminase antibody assay of the culture medium of intestinal biopsy specimens can improve the accuracy of celiac disease diagnosis. Clin Chem. 2006 Jun;52(6):1175-80. PMID: 16574764.

21. Corrao G, Corazza GR, Andreani ML, et al. Serological screening of coeliac disease: choosing the optimal procedure according to various prevalence values. Gut. 1994 Jun;35(6):771-5. PMID: 8020803.

22. Delgado JF, Amengual MJ, Veraguas A, et al. Paediatric celiac patients carrying the HLA-DR7-DQ2 and HLA-DR3-DQ2 haplotypes display small clinical differences. Acta Paediatr. 2014 Feb 19PMID: 24628273.

23. Demir H, Yuce A, Kocak N, et al. Celiac disease in Turkish children: Presentation of 104 cases. Pediatrics International. 2000 Oct;42(5):483-7. PMID: WOS:000089959700005.

24. Diamanti A, Maino C, Niveloni S, et al. Characterization of gastric mucosal lesions in patients with celiac disease: a prospective controlled study. Am J Gastroenterol. 1999 May;94(5):1313-9. PMID: 10235212.

25. Dickey W, Hughes DF, McMillan SA. Disappearance of endomysial antibodies in treated celiac disease does not indicate histological recovery. Am J Gastroenterol. 2000 Mar;95(3):712-4. PMID: 10710062.

26. Dickey W, Hughes DF, McMillan SA. Reliance on serum endomysial antibody testing underestimates the true prevalence of coeliac disease by one fifth. Scand J Gastroenterol. 2000 Feb;35(2):181-3. PMID: 10720117.

27. Dinler G, Atalay E, Kalayci AG. Celiac disease in 87 children with typical and atypical symptoms in Black Sea region of Turkey. World J Pediatr. 2009 Nov;5(4):282-6. PMID: 19911143.

28. Duerksen DR, Wilhelm-Boyles C, Veitch R, et al. A comparison of antibody testing, permeability testing, and zonulin levels with small-bowel biopsy in celiac disease patients on a gluten-free diet. Dig Dis Sci. 2010 Apr;55(4):1026-31. PMID: 19399613.

29. Fabiani E, Taccari LM, Ratsch IM, et al. Compliance with gluten-free diet in adolescents with screening-detected celiac disease: a 5-year follow-up study. J Pediatr. 2000 Jun;136(6):841-3. PMID: 10839888.

30. Fernandez-Banares F, Alsina M, Modolell I, et al. Are positive serum-IgA-tissuetransglutaminase antibodies enough to diagnose coeliac disease without a small bowel biopsy? Post-test probability of coeliac disease. J Crohns Colitis. 2012 Sep;6(8):861-6. PMID: 22398076.

31. Galli G, Esposito G, Lahner E, et al. Histological recovery and gluten-free diet adherence: a prospective 1-year follow-up study of adult patients with coeliac disease. Aliment Pharmacol Ther. 2014 Sep;40(6):639-47. PMID: 25066096.

32. Hashemi J, Hajiani E, Shahbazin HB, et al. Prevalence of celiac disease in Iranian children with idiopathic short stature. World J Gastroenterol. 2008 Dec 28;14(48):7376-80. PMID: 19109872.

33. Hill PG, Holmes GK. Coeliac disease: a biopsy is not always necessary for diagnosis. Aliment Pharmacol Ther. 2008 Apr 1;27(7):572-7. PMID: 18194500.

34. Hogberg L, Grodzinsky E, Stenhammar L. Better dietary compliance in patients with coeliac disease diagnosed in early childhood. Scandinavian Journal of Gastroenterology. 2003 Jul;38(7):751-4. PMID: WOS:000184291900012.

35. Iwanczak B, Matusiewicz K, Iwanczak F. Clinical picture of classical, atypical and silent celiac disease in children and adolescents. Adv Clin Exp Med. 2013 Sep-Oct;22(5):667-73. PMID: 24285451.

36. Johnson TC, Diamond B, Memeo L, et al. Relationship of HLA-DQ8 and severity of celiac disease: comparison of New York and Parisian cohorts. Clin Gastroenterol Hepatol. 2004 Oct;2(10):888-94. PMID: 15476152.

37. Kapitany A, Toth L, Tumpek J, et al. Diagnostic significance of HLA-DQ typing in patients with previous coeliac disease diagnosis based on histology alone. Aliment Pharmacol Ther. 2006 Nov 1;24(9):1395-402. PMID: 17059521.

38. Kelly CP, Feighery CF, Gallagher RB, et al. Mucosal and Systemic Iga Antigliadin Antibody in Celiac-Disease - Contrasting Patterns of Response in Serum, Saliva, and Intestinal Secretions. Digestive Diseases and Sciences. 1991 Jun;36(6):743-51. PMID: WOS:A1991FP14300007.

39. Klapp G, Masip E, Bolonio M, et al. Celiac disease: the new proposed ESPGHAN diagnostic criteria do work well in a selected population. J Pediatr Gastroenterol Nutr. 2013 Mar;56(3):251-6. PMID: 23111763.

40. Kokkonen J, Holm K, Karttunen TJ, et al. Children with untreated food allergy express a relative increment in the density of duodenal gammadelta+ T cells. Scand J Gastroenterol. 2000 Nov;35(11):1137-42. PMID: 11145283.

41. Kotze LM. Celiac disease in Brazilian patients: associations, complications and causes of death. Forty years of clinical experience. Arq Gastroenterol. 2009 Oct-Dec;46(4):261-9. PMID: 20232004.

42. Kurppa K, Ashorn M, Iltanen S, et al. Celiac disease without villous atrophy in children: a prospective study. J Pediatr. 2010 Sep;157(3):373-80, 80 e1. PMID: 20400102.

43. Langenberg MC, Wismans PJ, van Genderen PJ. Distinguishing tropical sprue from celiac disease in returning travellers with chronic diarrhoea: a diagnostic challenge? Travel Med Infect Dis. 2014 Jul-Aug;12(4):401-5. PMID: 24889052.

44. Licata A, Cappello M, Arini A, et al. Serology in adults with celiac disease: limited accuracy in patients with mild histological lesions. Intern Emerg Med. 2012 Aug;7(4):337-42. PMID: 21468695.

45. Lichtwark IT, Newnham ED, Robinson SR, et al. Cognitive impairment in coeliac disease improves on a gluten-free diet and correlates with histological and serological indices of disease severity. Aliment Pharmacol Ther. 2014 Jul;40(2):160-70. PMID: 24889390.

46. Lurz E, Scheidegger U, Spalinger J, et al. Clinical presentation of celiac disease and the diagnostic accuracy of serologic markers in children. Eur J Pediatr. 2009 Jul;168(7):839-45. PMID: 18923841.

47. Macchini F, Selicorni A, Luzzani S, et al. Coeliac disease and Cornelia de Lange syndrome: lack of association. Acta Paediatr. 2007 Oct;96(10):1518-20. PMID: 17850398.

48. Makharia GK, Baba CS, Khadgawat R, et al. Celiac disease: variations of presentations in adults. Indian J Gastroenterol. 2007 Jul-Aug;26(4):162-6. PMID: 17986741.

49. Misak Z, Hojsak I, Jadresin O, et al. Diagnosis of coeliac disease in children younger than 2 years. J Pediatr Gastroenterol Nutr. 2013 Feb;56(2):201-5. PMID: 23325441.

50. Mohindra S, Yachha SK, Srivastava A, et al. Coeliac disease in Indian children: assessment of clinical, nutritional and pathologic characteristics. J Health Popul Nutr. 2001 Sep;19(3):204-8. PMID: 11761775.

51. Mubarak A, Gmelig-Meyling FH, Wolters VM, et al. Immunoglobulin G antibodies against deamidated-gliadin-peptides outperform anti-endomysium and tissue transglutaminase antibodies in children <2 years age. APMIS. 2011 Dec;119(12):894-900. PMID: 22085366.

52. Mubarak A, Spierings E, Wolters V, et al. Human leukocyte antigen DQ2.2 and celiac disease. J Pediatr Gastroenterol Nutr. 2013 Apr;56(4):428-30. PMID: 23085892.

53. Mubarak A, Wolters VM, Gerritsen SA, et al. A biopsy is not always necessary to diagnose celiac disease. J Pediatr Gastroenterol Nutr. 2011 May;52(5):554-7. PMID: 21240025.

54. Pacht A, Sinai N, Hornstein L, et al. The diagnostic reliability of anti-endomysial antibody in celiac disease: the north Israel experience. Isr J Med Sci. 1995 Apr;31(4):218-20. PMID: 7721558.

55. Parizade M, Bujanover Y, Weiss B, et al. Performance of serology assays for diagnosing celiac disease in a clinical setting. Clin Vaccine Immunol. 2009 Nov;16(11):1576-82. PMID: 19776198.

56. Rabbani MW, Aziz MT, Ali I, et al. Diagnostic usefulness of Anti-Tissue Transglutaminase in Celiac Disease: Correlation with Intestinal Mucosal Biopsy. Pakistan Journal of Medical Sciences. 2011 Apr-Jun;27(3):599-602. PMID: WOS:000293485000028.

57. Rawal P, Thapa BR, Nain CK, et al. Changing spectrum of celiac disease in India. Iran J Pediatr. 2010 Dec;20(4):459-65. PMID: 23056746.

58. Ribes-Koninckx C, Alfonso P, Ortigosa L, et al. A beta-turn rich oats peptide as an antigen in an ELISA method for the screening of coeliac disease in a paediatric population. Eur J Clin Invest. 2000 Aug;30(8):702-8. PMID: 10964162.

59. Richter T, Bossuyt X, Vermeersch P, et al. Determination of IgG and IgA antibodies against native gliadin is not helpful for the diagnosis of coeliac disease in children up to 2 years old. J Pediatr Gastroenterol Nutr. 2012 Jul;55(1):21-5. PMID: 22249805.

60. Rodrigo-Saez L, Fuentes-Alvarez D, Perez-Martinez I, et al. Differences between pediatric and adult celiac disease. Rev Esp Enferm Dig. 2011 May;103(5):238-44. PMID: 21619387.

61. Rose C, Brocker EB, Zillikens D. Clinical, histological and immunpathological findings in 32 patients with dermatitis herpetiformis Duhring. J Dtsch Dermatol Ges. 2010 Apr;8(4):265-70, -71. PMID: 19878401.

62. Rostami K, Kerckhaert J, Tiemessen R, et al. Sensitivity of antiendomysium and antigliadin antibodies in untreated celiac disease: disappointing in clinical practice. Am J Gastroenterol. 1999 Apr;94(4):888-94. PMID: 10201452.

63. Rozenberg O, Lerner A, Pacht A, et al. A new algorithm for the diagnosis of celiac disease. Cell Mol Immunol. 2011 Mar;8(2):146-9. PMID: 21317919.

64. Rozenberg O, Lerner A, Pacht A, et al. A novel algorithm for the diagnosis of celiac disease and a comprehensive review of celiac disease diagnostics. Clin Rev Allergy Immunol. 2012 Jun;42(3):331-41. PMID: 21279475.

65. Russo PA, Chartrand LJ, Seidman E. Comparative analysis of serologic screening tests for the initial diagnosis of celiac disease. Pediatrics. 1999 Jul;104(1 Pt 1):75-8. PMID: 10390263.

66. Saginur M, AlRefaee FAM, Spady DW, et al. Antitissue transglutaminase antibody determination versus upper endoscopic biopsy diagnosis of paediatric celiac disease. Paediatrics & Child Health. 2013 May;18(5):246-50. PMID: WOS:000318787700007.

67. Sayed SK, Imam HM, Mahran AM, et al. Diagnostic utility of deamidated gliadin peptide antibody in celiac disease compared to anti-tissue transglutaminase and IgA- endomysium antibodies. Egypt J Immunol. 2012;19(2):41-52. PMID: 23885406.

68. Schirru E, Danjou F, Cicotto L, et al. Anti-actin IgA antibodies identify celiac disease patients with a Marsh 3 Intestinal damage among subjects with moderate anti-TG2 levels. Biomed Res Int. 2013;2013:630463. PMID: 24083232.

69. Scoglio R, Di Pasquale G, Pagano G, et al. Is intestinal biopsy always needed for diagnosis of celiac disease? Am J Gastroenterol. 2003 Jun;98(6):1325-31. PMID: 12818277.

70. Sorell L, Garrote JA, Galvan JA, et al. Celiac disease diagnosis in patients with giardiasis: high value of antitransglutaminase antibodies. Am J Gastroenterol. 2004 Jul;99(7):1330-2. PMID: 15233673.

71. Tighe MR, Hall MA, Ashkenazi A, et al. Celiac disease among Ashkenazi Jews from Israel. A study of the HLA class II alleles and their associations with disease susceptibility. Hum Immunol. 1993 Dec;38(4):270-6. PMID: 8138422.

72. Tosic Z, Salkic N, Krizic N, et al. Celiac disease in adult population in Tuzla region of Bosnia and Herzegovina: a 3-year surveillance (2007-2009). Med Arh. 2013;67(5):333-5. PMID: 24601164.

73. Tursi A, Brandimarte G, Giorgetti GM. Prevalence of antitissue transglutaminase antibodies in different degrees of intestinal damage in celiac disease. J Clin Gastroenterol. 2003 Mar;36(3):219-21. PMID: 12590232.

74. Tursi A, Brandimarte G, Giorgetti GM, et al. Endoscopic features of celiac disease in adults and their correlation with age, histological damage, and clinical form of the disease. Endoscopy. 2002 Oct;34(10):787-92. PMID: 12244499.

75. Tursi A, Giorgetti G, Brandimarte G, et al. Prevalence and clinical presentation of subclinical/silent celiac disease in adults: an analysis on a 12-year observation. Hepatogastroenterology. 2001 Mar-Apr;48(38):462-4. PMID: 11379333.

76. Vahedi K, Mascart F, Mary JY, et al. Reliability of antitransglutaminase antibodies as predictors of gluten-free diet compliance in adult celiac disease. Am J Gastroenterol. 2003 May;98(5):1079-87. PMID: 12809831.

77. Vaquero L, Caminero A, Nunez A, et al. Coeliac disease screening in first-degree relatives on the basis of biopsy and genetic risk. Eur J Gastroenterol Hepatol. 2014 Mar;26(3):263-7. PMID: 24300305.

78. Villanacci V, Bassotti G, Liserre B, et al. Helicobacter pylori infection in patients with celiac disease. Am J Gastroenterol. 2006 Aug;101(8):1880-5. PMID: 16780559.

79. Vivas S, de Morales JMR, Fernandez M, et al. Age-Related Clinical, Serological, and Histopathological Features of Celiac Disease. American Journal of Gastroenterology. 2008 Sep;103(9):2360-5. PMID: WOS:000260170000028.

80. Volta U, De Giorgio R, Petrolini N, et al. Clinical findings and anti-neuronal antibodies in coeliac disease with neurological disorders. Scand J Gastroenterol. 2002 Nov;37(11):1276-81. PMID: 12465725.

81. Volta U, Granito A, Parisi C, et al. Deamidated gliadin peptide antibodies as a routine test for celiac disease: a prospective analysis. J Clin Gastroenterol. 2010 Mar;44(3):186-90. PMID: 20042872.

82. Volta U, Tovoli F, Cicola R, et al. Serological tests in gluten sensitivity (nonceliac gluten intolerance). J Clin Gastroenterol. 2012 Sep;46(8):680-5. PMID: 22138844.

83. Westerbeek E, Mouat S, Wesley A, et al. Coeliac disease diagnosed at Starship Children's Hospital: 1999-2002. N Z Med J. 2005 Aug 12;118(1220):U1613. PMID: 16132074.

Appendix C. Evidence Table

Table C-1. Studies of serology accuracy

			Outcomes	
			Sensitivity,	
	Number of Porticipants	Type of Diagnostic Test	Specificity, Resitive Predictive Value	
Author Year	Populations	Cut-Off Value	Negative Predictive Value,	
Barada et al.,	Number of Participants: 999 Adults	Type of Diagnostic Test: EMA IgA	Sensitivity: 72.2%	QUADAS Domain 1
2014 ³¹		Cut-off value: NR	Specificity: 99.7%	Consecutive or random sample: Yes
			Positive predictive value: 90	Case control design avoided: Yes
			Negative predictive value: 99.2	Inappropriate exclusions: Yes
				Biased patient Selection: Low
			Sensitivity: 72.2%	
		Type of Diagnostic Test: tTG IgA	Specificity: 98.4%	QUADAS Domain 2
		Cut-off value: NR	Positive predictive value: 44.8 Negative predictive value: 99.5	Blinded interpretation of index test results: Yes
			-3	Prespecified test threshold: Yes
			Sensitivity: 72.2% Specificity: 97.4%	Bias due to testing: Low
		Type of Diagnostic Test: Combined	Positive predictive value: 34.2	QUADAS Domain 3
		screen tTG IgA, DGP IgA	Negative predictive value: 99.5	Valid reference standard: Yes
	Comments: Marsh 2 & 3 were considered	Cut-off value: NR	5	Blinded analysis of reference test: Yes
	celiac. In addition, authors classified 1 person			Bias due to reference test: Low
	with Marsh T and positive EMA as cellac.			QUADAS Domain 4
				Appropriate interval between reference and
				index test: Yes
				All patients received reference test: Yes
				All patients received same test: Yes
				All patients included analysis: Yes
				Could patient flow have introduced bias:
				Not Applicable

			Outcomes Sensitivity, Specificity,	
Author Year	Number of Participants, Populations	Type of Diagnostic Test, Cut-Off Value	Positive Predictive Value,	OUADAS
Basso et al., 2011 ³²	Number of Participants: 703 Adults	Type of Diagnostic Test: tTG IgA	Sensitivity: 75.7%	QUADAS Domain 1
,		Cut-off value: 100 U/mL	Specificity: 100%	Consecutive or random sample: Yes
			Positive predictive value: 100	Case control design avoided: No
			Negative predictive value: 82.4	Inappropriate exclusions: Yes
				Biased patient Selection: Low
		Type of Diagnostic Test: tTG IgA	Sensitivity: 94.5%	
		Cut-off value: 17.5 U/mL	Specificity: 97.1%	QUADAS Domain 2
			Positive predictive value: 96.6	Blinded interpretation of index test results:
			Negative predictive value: 95.3	Unclear
				Prespecified test threshold: Yes
		Type of Diagnostic Test: tTG IgA	Sensitivity: 94.2%	Bias due to testing: Unclear
		Cut-off value: 20 U	Specificity: 97.3%	
			Positive predictive value: 96.9	QUADAS Domain 3
			Negative predictive value: 95	Valid reference standard: Yes
		Type of Diagnostic Test: TIG IgA		Blinded analysis of reference test: Unclear
		Cut-on value: 24 U/mL	Sensitivity: 90.3%	Bias due to reference test: Unclear
			Bositivo prodictivo valuo: 81.0	OLIADAS Domain 4
			Negative predictive value: 96.2	Appropriate interval between reference and
		Type of Diagnostic Test: tTG IgA	Negative predictive value. 90.2	index test. Yes
		Cut-off value: 75.6 Ll/ml	Sensitivity: 90.9%	All natients received reference test. Yes
			Specificity: 96.5%	All patients received same test. Yes
			Positive predictive value: 95.8	All patients included analysis: No
			Negative predictive value: 92.3	Could patient flow have introduced bias:
				Not Applicable
		Type of Diagnostic Test: tTG IgA	Sensitivity: 62.6%	
		Cut-off value: 909.3 U/mL	Specificity: 100%	
			Positive predictive value: 100	
			Negative predictive value: 75.2	

			Outcomes Sensitivity, Specificity,	
	Number of Participants,	Type of Diagnostic Test,	Positive Predictive Value,	
Author, Year	Populations	Cut-Off Value	Negative Predictive Value	QUADAS
Basso et al., 2011 ³²	Number of Participants: 703 Adults	Type of Diagnostic Test: tTG IgA,	Sensitivity: 65.3%	QUADAS Domain 1
		DGP IgA	Specificity: 100%	Consecutive or random sample: Yes
		Cut-off value: 145 U	Positive predictive value: 100	Case control design avoided: No
			Negative predictive value: 76.6	Inappropriate exclusions: Yes
				Biased patient Selection: Low
		Type of Diagnostic Test: tTG IgA,	Sensitivity: 96.7%	
		DGP IgA	Specificity: 89.8%	QUADAS Domain 2
		Cut-off value: 20 U	Positive predictive value: 89.3	Blinded interpretation of index test results:
			Negative predictive value: 96.8	Unclear
				Prespecified test threshold: Yes
			Sensitivity: 95.4%	Bias due to testing: Unclear
		Type of Diagnostic Test: tTG IgA,	Specificity: 95.7%	
		DGP IgA	Positive predictive value: 95.2	QUADAS Domain 3
		Cut-off value: 32 U	Negative predictive value: 96	Valid reference standard: Yes
				Blinded analysis of reference test: Unclear
			Sensitivity: 96.7%	Bias due to reference test: Unclear
		Type of Diagnostic Test: tTG IgG	Specificity: 83.4%	
		Cut-off value: 20 U/mL	Positive predictive value: 83.7	QUADAS Domain 4
			Negative predictive value: 96.6	Appropriate interval between reference and index test: Yes
			Sensitivity: 93.3%	All patients received reference test: Yes
		Type of Diagnostic Test: tTG IgG	Specificity: 94.1%	All patients received same test: Yes
		Cut-off value: 47.6 U/mL	Positive predictive value: 93.3	All patients included analysis: No
			Negative predictive value: 94.1	Could patient flow have introduced bias:
				Not Applicable
			Sensitivity: 59.6%	
		Type of Diagnostic Test: tTG IgG	Specificity: 100%	
		Cut-off value: 976.8 U/mL	Positive predictive value: 100	
			Negative predictive value: 73.8	

Author, Year	Number of Participants, Populations	Type of Diagnostic Test, Cut-Off Value	Outcomes Sensitivity, Specificity, Positive Predictive Value, Negative Predictive Value	QUADAS
Bienvenu et al., 2014 ³³	Number of Participants: 45 Population: Selective IgA deficient children	Type of Diagnostic Test: CD-LFIA (detects both human IgA and IgG anti-DGP) Cut-off value: NA	Sensitivity: 100.0% Specificity: 89.2% Negative predictive value: 100.0%	QUADAS Domain 1 Consecutive or random sample: Yes Case control design: Yes Inappropriate exclusions: Yes Biased patient Selection: Low QUADAS Domain 2 Blinded interpretation of index test results: Yes Prespecified test threshold: Yes Bias due to testing: Low QUADAS Domain 3 Valid reference standard: Yes Blinded analysis of reference test: Yes Bias due to reference test: Low QUADAS Domain 4 Appropriate interval between reference and index test: Unclear All patients received same test: Unclear All patients included analysis: Yes Could patient flow have introduced bias: Hiph

Author, Year	Number of Participants, Populations	Type of Diagnostic Test, Cut-Off Value	Outcomes Sensitivity, Specificity, Positive Predictive Value, Negative Predictive Value	QUADAS
Cekin et al., 2012 ³⁴	Number of Participants: 84 Adults with Iron Deficiency	Type of Diagnostic Test: EMA IgA	Sensitivity: 100% Specificity: 98.72% Positive predictive value: 85.71 Negative predictive value: 100	QUADAS Domain 1 Consecutive or random sample: Yes Case control design avoided: Yes Inappropriate exclusions: Yes Biased patient Selection: Low
		Type of Diagnostic Test: EMA IgG	Sensitivity: 33.33% Specificity: 96.15% Positive predictive value: 40 Negative predictive value: 94.94	QUADAS Domain 2Blinded interpretation of index test results: YesPrespecified test threshold: YesBias due to testing: LowQUADAS Domain 3Valid reference standard: YesBlinded analysis of reference test: UnclearBias due to reference test: UnclearQUADAS Domain 4Appropriate interval between reference and index test: UnclearAll patients received reference test: YesAll patients included analysis: Yes Could patient flow have introduced bias:

Author, Year	Number of Participants, Populations	Type of Diagnostic Test, Cut-Off Value	Outcomes Sensitivity, Specificity, Positive Predictive Value, Negative Predictive Value	QUADAS
Dahlbom et al., 2010 ³⁵	Number of Participants: 301 Children and Adults	Type of Diagnostic Test: tTG IgA Cut-off value: >3 U m/L	Sensitivity: 100% Specificity: 99.24% Positive predictive value: 99.42	QUADAS Domain 1 Consecutive or random sample: Yes Case control design avoided: No Inappropriate exclusions: Not Applicable
		Type of Diagnostic Test: tTC IgC	Negative predictive value: 100	Blased patient Selection: High
		Cut-off value: >3 U m/L	Sensitivity: 84.12% Specificity: 98.47% Positive predictive value: 98.62 Negative predictive value: 82.69	QUADAS Domain 2 Blinded interpretation of index test results: Unclear Prespecified test threshold: Yes Bias due to testing: Unclear
				QUADAS Domain 3 Valid reference standard: Yes Blinded analysis of reference test: Unclear Bias due to reference test: Unclear
				QUADAS Domain 4 Appropriate interval between reference and index test: Yes All patients received reference test: Yes All patients received same test: Yes All patients included analysis: Yes Could patient flow have introduced bias: Not Applicable

Author, Year	Number of Participants, Populations	Type of Diagnostic Test, Cut-Off Value	Outcomes Sensitivity, Specificity, Positive Predictive Value, Negative Predictive Value	QUADAS
Dahle et al., 2010 ³⁶	Number of Participants: 176 Adults	Type of Diagnostic Test: EMA IgA Cut-off value: Serum dilution 1/5	Sensitivity: 61% Specificity: 100%	QUADAS Domain 1 Consecutive or random sample: Yes Case control design avoided: Yes
		Type of Diagnostic Test: tTG IgA Cut-off value: 5 U/mL	Sensitivity: 76% Specificity: 95%	Inappropriate exclusions: Yes Biased patient Selection: Low
		Type of Diagnostic Test:, DGP IgA or DGP IgG Cut-off value: 20 Au/mL	Sensitivity: 87% Specificity: 96%	QUADAS Domain 2 Blinded interpretation of index test results: Unclear Prespecified test threshold: Yes
		Type of Diagnostic Test: tTG IgG or IgA combined with DGP IgG or IgA Cut-off value: 20 Au/mL	Sensitivity: 91% Specificity: 80%	Bias due to testing: Unclear QUADAS Domain 3 Valid reference standard: Yes
		Type of Diagnostic Test: tTG IgG or IgA combined with DGP IgG or IgA Cut-off value: 35 AU/mL	Sensitivity: 85% Specificity: 98%	Blinded analysis of reference test: Yes Bias due to reference test: Low
				QUADAS Domain 4 Appropriate interval between reference and index test: Unclear All patients received reference test: Yes All patients received same test: No All patients included analysis: Yes Could patient flow have introduced bias: Unclear

Author, Year	Number of Participants, Populations	Type of Diagnostic Test, Cut-Off Value	Outcomes Sensitivity, Specificity, Positive Predictive Value, Negative Predictive Value	QUADAS
DeGaetani et al., 2013 ³⁷	Number of Participants: 59 Adults with prior negative serology but villious atrophy. HLA test was used to rule out celiac disease.	Type of Diagnostic Test: HLA DQ2, HLA DQ2	Sensitivity: 100% Specificity: 18.18% Positive predictive value: 29.41 Negative predictive value: 100	QUADAS Domain 1 Consecutive or random sample: Yes Case control design avoided: Yes Inappropriate exclusions: No Biased patient Selection: High QUADAS Domain 2 Blinded interpretation of index test results: Unclear Prespecified test threshold: Unclear Bias due to testing: Unclear
				QUADAS Domain 3 Valid reference standard: Yes Blinded analysis of reference test: Unclear Bias due to reference test: Unclear QUADAS Domain 4 Appropriate interval between reference and index test: Unclear All patients received reference test: Yes All patients included analysis: Yes Could patient flow have introduced bias: Unclear
Author, Year	Number of Participants, Populations	Type of Diagnostic Test, Cut-Off Value	Outcomes Sensitivity, Specificity, Positive Predictive Value, Negative Predictive Value	QUADAS
--	---	--	---	---
Author, Year Dutta et al., 2010 ³⁸	Populations Number of Participants: 92 symptomatic adults in India Comment: Unclear why tTG IgG test was used	Cut-Off Value Type of Diagnostic Test: tTG IgG Cut-off value: >15 U/mL	Negative Predictive Value Sensitivity: 77.8% Specificity: 89.1% Positive predictive value: 63.6 Negative predictive value: 94.2	QUADASQUADAS Domain 1Consecutive or random sample: YesCase control design avoided: YesInappropriate exclusions: YesBiased patient Selection: LowQUADAS Domain 2Blinded interpretation of index test results:YesPrespecified test threshold: YesBias due to testing: LowQUADAS Domain 3Valid reference standard: YesBlinded analysis of reference test: YesBias due to reference test: LowQUADAS Domain 4Appropriate interval between reference andindex test: UnclearAll patients received same test: Yes
				All patients included analysis: Yes Could patient flow have introduced bias: Not Applicable

Author, Year	Number of Participants, Populations	Type of Diagnostic Test, Cut-Off Value	Outcomes Sensitivity, Specificity, Positive Predictive Value, Negative Predictive Value	QUADAS
Emami et al., 2012 ³⁹	Number of Participants: 130 Population: IgA Deficient adults in Iran	Type of Diagnostic Test: tTG IgA Cut-off value: >10 AU/ml	Sensitivity: 38.46% Specificity: 96.58% Positive predictive value: 55.56 Negative predictive value: 93.39	QUADAS Domain 1 Consecutive or random sample: Yes Case control design avoided: Yes Inappropriate exclusions: Yes Biased patient Selection: Low QUADAS Domain 2 Blinded interpretation of index test results: Unclear Prespecified test threshold: Yes Bias due to testing: Unclear QUADAS Domain 3 Valid reference standard: Yes Blinded analysis of reference test: Unclear Bias due to reference test: Unclear QUADAS Domain 4 Appropriate interval between reference and index test: Unclear All patients received reference test: Yes All patients received same test: Yes All patients included analysis: Yes Could patient flow have introduced bias: Unclear

Author, Year	Number of Participants, Populations	Type of Diagnostic Test, Cut-Off Value	Outcomes Sensitivity, Specificity, Positive Predictive Value, Negative Predictive Value	QUADAS
Harrison et al., 2013 ⁴⁰	Number of Participants: 12,289, age unclear. Some IgA deficient, but number not reported	Type of Diagnostic Test: tTG IgA, Cut-off value: 5 U/mL	Sensitivity: 86.8% Specificity: 99.9%	QUADAS Domain 1 Consecutive or random sample: Yes Case control design avoided: Yes Inappropriate exclusions: Yes
		Type of Diagnostic Test: tTG IgA, tTG IgG Cut-off value: 5 U/mL	Sensitivity: 92.1% Specificity: 99.9%	 Biased patient Selection: Low QUADAS Domain 2 Blinded interpretation of index test results: Unclear Prespecified test threshold: Yes Bias due to testing: Low QUADAS Domain 3 Valid reference standard: Yes Blinded analysis of reference test: Unclear Bias due to reference test: Low QUADAS Domain 4 Appropriate interval between reference and index test: Unclear All patients received reference test: Yes All patients included analysis: Yes Could patient flow have introduced bias: Not Applicable

Author, Year	Number of Participants, Populations	Type of Diagnostic Test, Cut-Off Value	Outcomes Sensitivity, Specificity, Positive Predictive Value, Negative Predictive Value	QUADAS
Kaukinen et al., 1999 ⁴¹	Number of Participants: 26 Population: Patients with endocrinologic disorders in Finland	Type of Diagnostic Test: HLA DQ2, HLA DQ2	Sensitivity: 100% Specificity: 33.33% Positive predictive value: 5.26 Negative predictive value: 100	QUADAS Domain 1 Consecutive or random sample: Yes Case control design avoided: No Inappropriate exclusions: No Biased patient Selection: High QUADAS Domain 2 Blinded interpretation of index test results: Yes Prespecified test threshold: Yes Bias due to testing: Low QUADAS Domain 3 Valid reference standard: Yes Blinded analysis of reference test: Yes Bias due to reference test: Low QUADAS Domain 4 Appropriate interval between reference and index test: Unclear All patients received reference test: No All patients included analysis: No Could patient flow have introduced bias:

Author, Year	Number of Participants, Populations	Type of Diagnostic Test, Cut-Off Value	Outcomes Sensitivity, Specificity, Positive Predictive Value, Negative Predictive Value	QUADAS
Mansour et al., 2011 ⁴²	Number of Participants: 62 Population: Type 1 diabetes, Iraq	Type of Diagnostic Test: EMA IgA Cut-off value: 20 U/mL	Sensitivity: 71.43% Specificity: 96.36% Positive predictive value: 71.43 Negative predictive value: 96.36	QUADAS Domain 1 Consecutive or random sample: Yes Case control design avoided: Yes Inappropriate exclusions: Yes Biased patient Selection: Low
		Type of Diagnostic Test: tTG IgA Cut-off value: 15 U/mL	Sensitivity: 71.43% Specificity: 92.73% Positive predictive value: 55.56 Negative predictive value:	QUADAS Domain 2 Blinded interpretation of index test results: Yes Prespecified test threshold: Yes Bias due to testing: Low
		Type of Diagnostic Test: tTG IgG Cut-off value: 15 U/mL	96.23 Sensitivity: 57.14% Specificity: 92.73% Positive predictive value: 50 Negative predictive value: 94.44	QUADAS Domain 3 Valid reference standard: Yes Blinded analysis of reference test: Yes Bias due to reference test: Low QUADAS Domain 4 Appropriate interval between reference and index test: Unclear All patients received reference test: Yes All patients received same test: Yes All patients included analysis: Yes Could patient flow have introduced bias: Unclear

Author, Year	Number of Participants, Populations	Type of Diagnostic Test, Cut-Off Value	Outcomes Sensitivity, Specificity, Positive Predictive Value, Negative Predictive Value	QUADAS
Mozo et al., 2012 ⁴³	Number of Participants: 200	Type of Diagnostic Test: DGP IgA Cut-off value: >7 U/mL	Sensitivity: 96% Specificity: 96% Positive predictive value: 96 Negative predictive value: 96	QUADAS Domain 1 Consecutive or random sample: Yes Case control design avoided: No Inappropriate exclusions: No Biased patient Selection: High
		Type of Diagnostic Test: DGP IgG Cut-off value: >7 U/mL	Sensitivity: 95% Specificity: 99% Positive predictive value: 98.9 Negative predictive value: 95.2	QUADAS Domain 2 Blinded interpretation of index test results: Yes Prespecified test threshold: Yes
		Type of Diagnostic Test: tTG IgA Cut-off value: >7 U/mL	Sensitivity: 89% Specificity: 94% Positive predictive value: 93.7 Negative predictive value: 89.5	Bias due to testing: Low QUADAS Domain 3 Valid reference standard: Yes Blinded analysis of reference test: Unclear Bias due to reference test: Unclear
				QUADAS Domain 4 Appropriate interval between reference and index test: Yes All patients received reference test: No All patients received same test: Not Applicable All patients included analysis: Yes Could patient flow have introduced bias: Unclear

Author, Year	Number of Participants, Populations	Type of Diagnostic Test, Cut-Off Value	Outcomes Sensitivity, Specificity, Positive Predictive Value, Negative Predictive Value	QUADAS
Nevoral et al., 2013 ⁴⁴	Number of Participants: 345 children and adolescents	Type of Diagnostic Test: tTG IgA, EMA IgG Cut-off value: 12 U/mL	Sensitivity: 76% Specificity: 85% Positive predictive value: 94 Negative predictive value: 53	QUADAS Domain 1 Consecutive or random sample: Yes Case control design avoided: Yes Inappropriate exclusions: Yes Biased patient Selection: Low
	Number of Participants: 32 first degree relatives	Type of Diagnostic Test: tTG IgA, EMA IgA Cut-off value: 12 U/mL	Sensitivity: 81% Specificity: 70%	QUADAS Domain 2 Blinded interpretation of index test results: Yes
	Number of Participants: 263 with Marsh 2 or 3 classification	Type of Diagnostic Test: tTG IgA, EMA IgA Cut-off value: 12 U/mL	Sensitivity: 83% Specificity: 67%	Prespecified test threshold: Yes Bias due to testing: Low
	Number of Participants: 40 Type 1 diabetes	Type of Diagnostic Test: tTG IgA, EMA IgA Cut-off value: 12 U/mL	Sensitivity: 93% Specificity: 64%	Valid reference standard: Yes Blinded analysis of reference test: Yes Bias due to reference test: Low
		Comment: New ESPGHAN algorithm used		QUADAS Domain 4 Appropriate interval between reference and index test: Yes All patients received reference test: Yes All patients received same test: Yes All patients included analysis: Yes Could patient flow have introduced bias: Not Applicable

Author, Year	Number of Participants, Populations	Type of Diagnostic Test, Cut-Off Value	Outcomes Sensitivity, Specificity, Positive Predictive Value, Negative Predictive Value	QUADAS
Olen et al., 2012 ⁴⁵	Number of Participants: 69 Population: <2 years old	Type of Diagnostic Test: DGP IgA Cut-off value: NR	Sensitivity: 100% Specificity: 31% Positive predictive value: 44	QUADAS Domain 1 Consecutive or random sample: Yes Case control design avoided: Yes Inappropriate exclusions: No
	Number of Participants: 408 Population: all patients	Type of Diagnostic Test: DGP IgA Cut-off value: NR	Sensitivity: 91% Specificity: 26% Positive predictive value: 51	Biased patient Selection: High QUADAS Domain 2 Blinded interpretation of index test results:
	Number of Participants: 67 Population: <2 years old	Type of Diagnostic Test: tTG IgA Cut-off value: NR	Sensitivity: 96% Specificity: 98% Positive predictive value: 96	Prespecified test threshold: Yes Bias due to testing: Low
	Number of Participants: 530 Population: all patients	Type of Diagnostic Test: tTG IgA Cut-off value:NR	Sensitivity: 94% Specificity: 86% Positive predictive value: 88	QUADAS Domain 3 Valid reference standard: Yes Blinded analysis of reference test: Yes Bias due to reference test: Low
	Comments: 93 individuals were excluded from study because the serology analyses had not been carried out at the participating immunology departments. Also, it isn't clear why some patients did not undergo DGP tests.			QUADAS Domain 4 Appropriate interval between reference and index test: Yes All patients received reference test: Yes All patients received same test: Yes All patients included analysis: Yes Could patient flow have introduced bias: Not Applicable

Author, Year	Number of Participants, Populations	Type of Diagnostic Test, Cut-Off Value	Outcomes Sensitivity, Specificity, Positive Predictive Value, Negative Predictive Value	QUADAS
Sakly et al., 2012 ⁴⁶	Number of Participants: 297 adults and children	Type of Diagnostic Test: DGP IgA Cut-off value: 25 IU/mL Type of Diagnostic Test: DGP IgG Cut-off value: 25 IU/mL	Sensitivity: 97% Specificity: 90.7% Sensitivity: 94.2% Specificity: 95.4%	QUADAS Domain 1 Consecutive or random sample: Yes Case control design avoided: No Inappropriate exclusions: Yes Biased patient Selection: High QUADAS Domain 2 Blinded interpretation of index test results:
				Unclear Prespecified test threshold: Yes Bias due to testing: Unclear QUADAS Domain 3 Valid reference standard: Yes Blinded analysis of reference test: Unclear Bias due to reference test: Unclear QUADAS Domain 4 Appropriate interval between reference and index test: Unclear All patients received reference test: Yes All patients received same test: Yes All patients included analysis: Yes Could patient flow have introduced bias: Unclear

Author, Year	Number of Participants, Populations	Type of Diagnostic Test, Cut-Off Value	Outcomes Sensitivity, Specificity, Positive Predictive Value, Negative Predictive Value	QUADAS
Srinivas et al., 2014 ⁴⁷	Number of Participants: 752 Population: Clinical features of celiac disease	Type of Diagnostic Test: tTG IgA Cut-off value: : <10 IU/mL	Sensitivity: 0.83 Specificity: 0.96 Sensitivity: 0.80 Specificity: 0.99	QUADAS Domain 1 Consecutive or random sample: Yes Case control design: Yes Inappropriate exclusions: Yes Biased patient Selection: Low
		Type of Diagnostic Test: IgA EMA	Negative predictive value:	Blinded interpretation of index test results: Unclear Prespecified test threshold: Yes Bias due to testing: Unclear QUADAS Domain 3 Valid reference standard: Yes Blinded analysis of reference test: Unclear
				Bias due to reference test: Unclear QUADAS Domain 4 Appropriate interval between reference and index test: No All patients received reference test: Yes All patients received same test: Yes All patients included analysis: Yes Could patient flow have introduced bias: High

Author, Year	Number of Participants, Populations	Type of Diagnostic Test, Cut-Off Value	Outcomes Sensitivity, Specificity, Positive Predictive Value, Negative Predictive Value	QUADAS
Srinivas et al., 2013 ⁴⁸	Number of Participants: 75	Type of Diagnostic Test: EMA IgG	Sensitivity: 83% Specificity: 99% Positive predictive value: 93 Negative predictive value: 98	QUADAS Domain 1 Consecutive or random sample: Yes Case control design avoided: Yes Inappropriate exclusions: Yes
	Number of Participants: 102	Type of Diagnostic Test: tTG IgA Cut-off value: 10 IU/mL	Sensitivity: 84% Specificity: 96% Positive predictive value: 72 Negative predictive value: 98	Biased patient Selection: Low QUADAS Domain 2 Blinded interpretation of index test results: Yes
	Number of Participants: 71	Type of Diagnostic Test: tTG IgA, EMA IgA	Sensitivity: 83% Specificity: 99% Positive predictive value: 97 Negative predictive value: 98	Prespecified test threshold: Yes Bias due to testing: Low QUADAS Domain 3 Valid reference standard: Yes Blinded analysis of reference test: Yes Bias due to reference test: Low QUADAS Domain 4 Appropriate interval between reference and index test: Unclear All patients received reference test: Yes All patients received same test: Yes All patients included analysis: Yes Could patient flow have introduced bias: Not Applicable

Author, Year	Number of Participants, Populations	Type of Diagnostic Test, Cut-Off Value	Outcomes Sensitivity, Specificity, Positive Predictive Value, Negative Predictive Value	QUADAS
Sugai et al., 2010 ⁴⁹	Number of Participants: 17 IgA tTG negative adults with villous atrophy	Type of Diagnostic Test: DGP	Sensitivity: 35.71% Specificity: 100%	QUADAS Domain 1 Consecutive or random sample: Yes Case control design avoided: Yes
		Type of Diagnostic Test: tTG IgA, DGP IgA	Sensitivity: 42.86% Specificity: 100%	Inappropriate exclusions: Yes Biased patient Selection: Low
				QUADAS Domain 2 Blinded interpretation of index test results: Yes Prespecified test threshold: Yes Bias due to testing: Low QUADAS Domain 3
	Comments: Original N = 22, five patients refused biopsy.			Valid reference standard: Yes Blinded analysis of reference test: Yes Bias due to reference test: Low QUADAS Domain 4
				Appropriate interval between reference and index test: Yes All patients received reference test: No All patients received same test: Yes All patients included analysis: No Could patient flow have introduced bias: High

Author, Year	Number of Participants, Populations	Type of Diagnostic Test, Cut-Off Value	Outcomes Sensitivity, Specificity, Positive Predictive Value, Negative Predictive Value	QUADAS
Swallow et al., 2013 ⁵⁰	Number of Participants: 733 Adults Results when Marsh 1-2 considered celiac	Type of Diagnostic Test: EMA IgA	Sensitivity: 42.9% Specificity: 99.5% Positive predictive value: 42.9 Negative predictive value: 99.5	QUADAS Domain 1 Consecutive or random sample: Yes Case control design avoided: Yes Inappropriate exclusions: No Biased patient Selection: High
	Number of Participants: 756 Adults Results when Marsh 1-3 considered celiac	Type of Diagnostic Test: EMA IgA	Sensitivity: 73.3% Specificity: 99.5% Positive predictive value: 84.6 Negative predictive value: 98.9	QUADAS Domain 2 Blinded interpretation of index test results: Yes Prespecified test threshold: Yes
	Number of Participants: 756 Adults Results when Marsh 3 considered celiac	Type of Diagnostic Test: EMA IgA	Sensitivity: 82.6% Specificity: 99.1% Positive predictive value: 73.1 Negative predictive value: 99.5	Bias due to testing: Low QUADAS Domain 3 Valid reference standard: Yes
	Number of Participants: 733Adults Results when Marsh 1-2 considered celiac	Type of Diagnostic Test: , tTG IgA followed by EMA IgA, (NICE two step strategy)	Sensitivity: 57.1% Specificity: 97.3% Positive predictive value: 16.7 Negative predictive value: 99.6	Blinded analysis of reference test: Yes Bias due to reference test: Low QUADAS Domain 4 Appropriate interval between reference and index test: Yes
	Number of Participants: 756 Adults Results when Marsh 1-3 considered celiac	Type of Diagnostic Test: , tTG IgA followed by EMA IgA, (NICE two step strategy)	Sensitivity: 80% Specificity: 97.3% Positive predictive value: 54.6 Negative predictive value: 99.2	All patients received reference test: Yes All patients received same test: Yes All patients included analysis: Yes Could patient flow have introduced bias: Not Applicable
	Number of Participants: 756 Population: Marsh 3 Comments: 473 patients were excluded because only one of the two serology tests was performed. 14 of these were diagnosed as CD via biopsy.	Type of Diagnostic Test: , tTG IgA followed by EMA IgA, (NICE two step strategy)	Sensitivity: 87% Specificity: 96.9% Positive predictive value: 46.5 Negative predictive value: 99.6	

Author, Year	Number of Participants, Populations	Type of Diagnostic Test, Cut-Off Value	Outcomes Sensitivity, Specificity, Positive Predictive Value, Negative Predictive Value	QUADAS
Swallow et al., 2013 ⁵⁰	Number of Participants: 733Adults Results when Marsh 1-2 considered celiac		Sensitivity: 42.9% Specificity: 99.5% Positive predictive value: 42.9 Negative predictive value: 99.5	QUADAS Domain 1 Consecutive or random sample: Yes Case control design avoided: Yes Inappropriate exclusions: No Biased patient Selection: High
	Number of Participants: 756 Adults	Type of Diagnostic Test: TTG IgA	Sensitivity: 73.3% Specificity: 99.5% Positive predictive value: 84.6 Negative predictive value: 98.9	QUADAS Domain 2 Blinded interpretation of index test results: Yes
	Results when Marsh 1-3 considered celiac	Type of Diagnostic Test: tTG IgA	Sensitivity: 82.6% Specificity: 99.1%	Prespecified test threshold: Yes Bias due to testing: Low
	Results when Marsh 3 considered celiac as CD via biopsy.	Type of Diagnostic Test: tTG IgA	Negative predictive value: 99.5	Valid reference standard: Yes Blinded analysis of reference test: Yes Bias due to reference test: Low
				Appropriate interval between reference and index test: Yes All patients received reference test: Yes All patients received same test: Yes All patients included analysis: Yes Could patient flow have introduced bias: Not Applicable

Author, Year	Number of Participants, Populations	Type of Diagnostic Test, Cut-Off Value	Outcomes Sensitivity, Specificity, Positive Predictive Value, Negative Predictive Value	QUADAS
Van Meensel et al., 2004 ⁵¹	Number of Participants: 175 Adults	Type of Diagnostic Test: tTG IgA Cut-off value: 10 kilounits/L	Sensitivity: 94% Specificity: 100%	QUADAS Domain 1 Consecutive or random sample: Yes
	Comment: 5 patients were IgA deficient	Type of Diagnostic Test: tTG IgA Cut-off value: 15 kilounits	Sensitivity: 94% Specificity: 100%	Case control design avoided: No Inappropriate exclusions: Yes Biased patient Selection: High
		Type of Diagnostic Test: tTG IgA Cut-off value: 19.05 kilounits/L	Sensitivity: 93% Specificity: 100%	QUADAS Domain 2 Blinded interpretation of index test results:
		Type of Diagnostic Test: tTG IgA Cut-off value: 2.64 kilounits/L	Sensitivity: 96% Specificity: 99%	Yes Prespecified test threshold: Yes Bias due to testing: Unclear
		Type of Diagnostic Test: tTG IgA Cut-off value: 20 kilounits	Sensitivity: 97% Specificity: 96%	QUADAS Domain 3 Valid reference standard: Yes
		Type of Diagnostic Test: tTG IgA Cut-off value: 20 kilounits/L	Sensitivity: 93% Specificity: 100%	Blinded analysis of reference test: Yes Bias due to reference test: Low
				QUADAS Domain 4 Appropriate interval between reference and index test: Yes All patients received reference test: Yes All patients received same test: Yes All patients included analysis: Yes Could patient flow have introduced bias: Not Applicable

Author, Year	Number of Participants, Populations	Type of Diagnostic Test, Cut-Off Value	Outcomes Sensitivity, Specificity, Positive Predictive Value, Negative Predictive Value	QUADAS
Van Meensel et al., 2004 ⁵¹	Number of Participants: 175 Adults	Type of Diagnostic Test: tTG IgA Cut-off value: 20.47 kilounits	Sensitivity: 97% Specificity: 100%	QUADAS Domain 1 Consecutive or random sample: Yes
	Comment. 5 patients were igA dencient	Type of Diagnostic Test: tTG IgA Cut-off value: 3.13 kilounits/L	Sensitivity: 96% Specificity: 99%	Inappropriate exclusions: Yes Biased patient Selection: High
		Type of Diagnostic Test: tTG IgA Cut-off value: 3.69 kilounits/L	Sensitivity: 96% Specificity: 100%	QUADAS Domain 2 Blinded interpretation of index test results:
		Type of Diagnostic Test: tTG IgA Cut-off value: 4 kilounits/L	Sensitivity: 93% Specificity: 99%	Yes Prespecified test threshold: Yes Bias due to testing: Unclear
		Type of Diagnostic Test: tTG IgA Cut-off value: 4.43 kilounits/L	Sensitivity: 99% Specificity: 99%	QUADAS Domain 3 Valid reference standard: Yes Blinded applysis of reference test: Yes
		Type of Diagnostic Test: tTG IgA Cut-off value: 40 kilounits/L	Sensitivity: 96% Specificity: 96%	Bias due to reference test: Low
				QUADAS Domain 4 Appropriate interval between reference and index test: Yes All patients received reference test: Yes All patients received same test: Yes All patients included analysis: Yes Could patient flow have introduced bias: Not Applicable

Author, Year	Number of Participants, Populations	Type of Diagnostic Test, Cut-Off Value	Outcomes Sensitivity, Specificity, Positive Predictive Value, Negative Predictive Value	QUADAS
Van Meensel et al., 2004 ⁵¹	Number of Participants: 175 Adults	Type of Diagnostic Test: tTG IgA Cut-off value: 5 kilounits/L	Sensitivity: 93% Specificity: 99%	QUADAS Domain 1 Consecutive or random sample: Yes
	Comment: 5 patients were IgA deficient			Case control design avoided: No
		Type of Diagnostic Test: tTG IgA	Sensitivity: 93%	Inappropriate exclusions: Yes
		Cut-off value: 50 kilounits/L	Specificity: 93%	Biased patient Selection: High
		Type of Diagnostic Test: tTG IgA	Sensitivity: 91%	QUADAS Domain 2
		Cut-off value: 56.9 kilounits/L	Specificity: 99%	Blinded interpretation of index test results: Yes
		Type of Diagnostic Test: tTG IgA	Sensitivity: 91%	Prespecified test threshold: Yes
		Cut-off value: 7 kilounits/L	Specificity: 100%	Bias due to testing: Unclear
		Type of Diagnostic Test: tTG IgA	Sensitivity: 97%	QUADAS Domain 3
		Cut-off value: 7 kilounits/L	Specificity: 100%	Valid reference standard: Yes
				Blinded analysis of reference test: Yes
		Type of Diagnostic Test: tTG IgA Cut-off value: 7.16 kilounits/L	Sensitivity: 97% Specificity: 100%	Bias due to reference test: Low
				QUADAS Domain 4
				Appropriate interval between reference and
				index test: Yes
				All patients received reference test: Yes
				All patients received same test: Yes
				All patients included analysis: Yes
				Not Applicable

Author, Year	Number of Participants, Populations	Type of Diagnostic Test, Cut-Off Value	Outcomes Sensitivity, Specificity, Positive Predictive Value, Negative Predictive Value	QUADAS
Van Meensel et al., 2004 ⁵¹	Number of Participants: 175 Comment: 5 patients were IgA deficient	Type of Diagnostic Test: tTG IgA Cut-off value: 7.98 kilounits Type of Diagnostic Test: tTG IgA Cut-off value: 9.73 kilounits/L	Sensitivity: 96% Specificity: 100% Sensitivity: 94% Specificity: 100%	QUADAS Domain 1 Consecutive or random sample: Yes Case control design avoided: No Inappropriate exclusions: Yes Biased patient Selection: High QUADAS Domain 2 Blinded interpretation of index test results: Yes Prespecified test threshold: Yes Bias due to testing: Unclear QUADAS Domain 3 Valid reference standard: Yes Blinded analysis of reference test: Yes Bias due to reference test: Low QUADAS Domain 4 Appropriate interval between reference and index test: Yes All patients received reference test: Yes All patients included analysis: Yes Could patient flow have introduced bias: Not Applicable

			Outcomes Sensitivity, Specificity.	
	Number of Participants,	Type of Diagnostic Test,	Positive Predictive Value,	
Author, Year	Populations	Cut-Off Value	Negative Predictive Value	QUADAS
Vermeersch et al., 2010 ⁵²	Number of Participants: 827 (599 adults, 228 children)	Type of Diagnostic Test: DGP IgA Cut-off value: >7	Sensitivity: 65.1% Specificity: 99.1%	QUADAS Domain 1 Consecutive or random sample: Yes Case control design avoided: No
	Number of Participants: 827			Inappropriate exclusions: Yes
		Type of Diagnostic Test: DGP IgG Cut-off value: 10	Sensitivity: 79.1% Specificity: 97.6%	Biased patient Selection: High
	Number of Participants: 827			QUADAS Domain 2
				Blinded interpretation of index test results:
		Type of Diagnostic Test: DGP IgG	Sensitivity: 83.7%	Unclear
	Number of Participants: 827	Cut-off value: 20	Specificity: 99.3%	Prespecified test threshold: Yes
				Bias due to testing: Unclear
	Number of Participants: 827	Type of Diagnostic Test: DGP IgG	Sensitivity: 76.7%	QUADAS Domain 3
		Cut-off value: 25	Specificity: 99.2%	Valid reference standard: Yes
				Blinded analysis of reference test: Yes
				Bias due to reference test: Low
	Number of Participants: 827	Type of Diagnostic Test: DGP IgG	Sensitivity: 86%	
		Cut-off value: >7	Specificity: 97.3%	QUADAS Domain 4
				Appropriate interval between reference and
		Turne of Discrepatio Tasts tTO Is A	Constitute a 0.4 00/	Index test: Unclear
		Cut-off value: 7	Specificity: 02%	All patients received same test: Yes
			opeomony. 92 /0	All patients included analysis: Ves
				Could natient flow have introduced higs:
				Unclear

Author, Year	Number of Participants, Populations	Type of Diagnostic Test, Cut-Off Value	Outcomes Sensitivity, Specificity, Positive Predictive Value, Negative Predictive Value	QUADAS
Vermeersch et al., 2010 ⁵²	Number of Participants: 827	Type of Diagnostic Test: tTG IgA Cut-off value: >15	Sensitivity: 88.4% Specificity: 94.9%	QUADAS Domain 1 Consecutive or random sample: Yes Case control design avoided: No Inappropriate exclusions: Yes
	Number of Participants: 827	Type of Diagnostic Test: tTG IgA Cut-off value: >7	Sensitivity: 83.7% Specificity: 98.4%	Biased patient Selection: High QUADAS Domain 2 Blinded interpretation of index test results:
	Number of Participants: 827	Type of Diagnostic Test: tTG IgG Cut-off value: >15	Sensitivity: 60.5% Specificity: 98.1%	Unclear Prespecified test threshold: Yes Bias due to testing: Unclear
	Number of Participants: 827	Type of Diagnostic Test: tTG IgG Cut-off value: >7	Sensitivity: 38.4% Specificity: 98.5%	QUADAS Domain 3 Valid reference standard: Yes Blinded analysis of reference test: Yes Bias due to reference test: Low QUADAS Domain 4 Appropriate interval between reference and index test: Unclear All patients received reference test: Yes All patients received same test: Yes All patients included analysis: Yes Could patient flow have introduced bias: Unclear

Author, Year	Number of Participants, Populations	Type of Diagnostic Test, Cut-Off Value	Outcomes Sensitivity, Specificity, Positive Predictive Value, Negative Predictive Value	QUADAS
Vermeersch et al., 2010 ⁵³	Number of Participants: 588 Adults	Type of Diagnostic Test: tTG IgA Cut-off value: >15 U/mL	Sensitivity: 86% Specificity: 95%	QUADAS Domain 1 Consecutive or random sample: Yes Case control design avoided: No Inappropriate exclusions: No
	Number of Participants: 588 Adults	Type of Diagnostic Test: tTG IgA Cut-off value: >=7 U/mL	Specificity: 92.7% Positive predictive value: 50.6	Biased patient Selection: High QUADAS Domain 2 Blinded interpretation of index test results: Unclear Prespecified test threshold: Yes Bias due to testing: Unclear QUADAS Domain 3 Valid reference standard: Yes Blinded analysis of reference test: Yes Bias due to reference test: Low QUADAS Domain 4 Appropriate interval between reference and index test: Unclear All patients received reference test: Yes All patients received same test: Yes All patients included analysis: Yes Could patient flow have introduced bias: Unclear

			Outcomes Sensitivity	
			Specificity,	
	Number of Participants,	Type of Diagnostic Test,	Positive Predictive Value,	
Author, Year	Populations	Cut-Off Value	Negative Predictive Value	QUADAS
Vermeersch et al.,	Number of Participants: 649 Adults and	Type of Diagnostic Test: DGP IgA +	Sensitivity: 89.7%	QUADAS Domain 1
2012 ⁵⁴	Children		Specificity: 93.3%	Consecutive or random sample: Yes
	Commenter Detrognestive study the controls	Cut-off value: 20 U/mL		Case control design avoided: No
	comments. Retrospective study, the controls		Soncitivity: 88 8%	Riasod patient Selection: High
	spanned years 2004 to 2000, while cases		Specificity: 95.6%	blased patient Selection. Flight
		tTG laG*	opeomory: 50.070	QUADAS Domain 2
		Cut-off value: 7 U/mL		Blinded interpretation of index test results:
			Sensitivity: 85%	Unclear
		Type of Diagnostic Test: DGP IgG	Specificity: 99.3%	Prespecified test threshold: Yes
		Cut-off value: 20 U/mL		Bias due to testing: Unclear
		Turne of Discrepation Tests DOD InC	Sensitivity: 86.9%	QUADAS Domain 3
		Cut off value: 7 1/ml	Specificity. 96.7%	Rlinded applysis of reference test: Vec
		Cut-on value. 7 O/IIIL		Binded analysis of reference test. Tes
			Sensitivity: 84.1%	Dida due la reference leat. Low
		Type of Diagnostic Test: tTG IgA	Specificity: 95.9%	QUADAS Domain 4
		Cut-off value: 20 U/mL	1 5	Appropriate interval between reference and
				index test: Unclear
			Sensitivity: 81.3%	All patients received reference test: Yes
		Type of Diagnostic Test: tTG IgA	Specificity: 98.5%	All patients received same test: Yes
		Cut-off value: 7 U/mL		All patients included analysis: Yes
		Combined test determines whether		Could patient flow have introduced blas:
		Inc tests instead of Ind tests		Unclear

Author, Year	Number of Participants, Populations	Type of Diagnostic Test, Cut-Off Value	Outcomes Sensitivity, Specificity, Positive Predictive Value, Negative Predictive Value	QUADAS
Wakim-Fleming et al., 2014 ⁵⁵	Number of Participants: 204 Population: Consecutive patients with biopsy proven cirrhosis	Type of Diagnostic Test: EMA Serum dilution >= 1/10 Type of Diagnostic Test: TTG Cut-off value: above 20 U	Sensitivity: 1.00 Specificity: 1.00 Positive predictive value: Negative predictive value: Sensitivity: 1.00 Specificity: 0.96 Positive predictive value: Negative predictive value:	QUADAS Domain 1 Consecutive or random sample: Yes Case control design: Yes Inappropriate exclusions: Yes Biased patient Selection: Low QUADAS Domain 2 Blinded interpretation of index test results: Yes Prespecified test threshold: Yes Bias due to testing: Low QUADAS Domain 3 Valid reference standard: Yes Blinded analysis of reference test: Yes Bias due to reference test: Low QUADAS Domain 4 Appropriate interval between reference and index test: Yes All patients received same test: Yes All patients received same test: Yes All patients included analysis: Yes Could patient flow have introduced bias: Low

Author, Year	Number of Participants, Populations	Type of Diagnostic Test, Cut-Off Value	Outcomes Sensitivity, Specificity, Positive Predictive Value, Negative Predictive Value	QUADAS
Wolf et al., 2014 ⁵⁷	Number of Participants: 1071 children Population: Selective IgA deficiency (sIgAD) was found in 27 patients	Type of Diagnostic Test: tTG IgA Cut-off value: >10 U/mL Type of Diagnostic Test: DGP IgG Cut-off value: >10 U/mL	Sensitivity: 0.88 Specificity: 0.97 Sensitivity: 0.89 Specificity: 0.95 when added to tTG in children without IgA deficiency Sensitivity: 0.29 Specificity: 1.00 when added to tTG in children WITH IgA deficiency	QUADAS Domain 1 Consecutive or random sample: Yes Case control design: No Inappropriate exclusions: Yes Biased patient Selection: High QUADAS Domain 2 Blinded interpretation of index test results: Yes Prespecified test threshold: Yes Bias due to testing: Low QUADAS Domain 3 Valid reference standard: Yes Blinded analysis of reference test: Yes Bias due to reference test: Low QUADAS Domain 4 Appropriate interval between reference and index test: Yes All patients received reference test: Yes All patients included analysis: Yes Could patient flow have introduced bias:

			Outcomes Sensitivity	
			Specificity.	
	Number of Participants,	Type of Diagnostic Test,	Positive Predictive Value,	
Author, Year	Populations	Cut-Off Value	Negative Predictive Value	QUADAS
Zanini et al., 201258	Number of Participants: 263 Adults, (Brand B	Type of Diagnostic Test: tTG IgA	Sensitivity: 89.4%	QUADAS Domain 1
	used)	Cut-off value: 16 U/mL	Specificity: 88.1%	Consecutive or random sample: Yes
			Positive predictive value: 90.3	Case control design avoided: Yes
			Negative predictive value: 77.4	Inappropriate exclusions: No
				Biased patient Selection: High
		Type of Diagnostic Test: tTG IgA	Sensitivity: 38.2%	
	Number of Participants: 393 Adults, (Brand A	Cut-off value: 21 U/mL	Specificity: 97.4%	QUADAS Domain 2
	used)		Positive predictive value: 95.8	Blinded interpretation of index test results:
	Number of Participants: 289Adults, (Brand C		Negative predictive value: 50.7	Unclear
	used)			Prespecified test threshold: Yes
		Type of Diagnostic Test: tTG IgA	Sensitivity: 58.8%	Bias due to testing: Unclear
		Cut-off value: 24 U/mL	Specificity: 99%	
			Positive predictive value: 99	QUADAS Domain 3
	Number of Participants: 393		Negative predictive value: 60.7	Valid reference standard: Yes
				Blinded analysis of reference test: Unclear
		Type of Diagnostic Test: tTG IgA	Sensitivity: 10.1%	Bias due to reference test: Unclear
		Cut-off value: 35 U/mL	Specificity: 100%	
			Positive predictive value: 100	QUADAS Domain 4
	Number of Participants: 289		Negative predictive value: 42	Appropriate interval between reference and index test: Yes
		Type of Diagnostic Test: tTG IgA	Sensitivity: 43.1%	All patients received reference test: Yes
		Cut-off value: 40 U/mL	Specificity: 100%	All patients received same test: Yes
			Positive predictive value: 100	All patients included analysis: Yes
	Number of Participants: 263		Negative predictive value: 53.1	Could patient flow have introduced bias:
				Not Applicable
		Type of Diagnostic Test: tTG IgA	Sensitivity: 69.7%	
		Cut-off value: 48 U/mL	Specificity: 58.8%	
			Positive predictive value: 100	
			Negative predictive value: 60.3	

Author Year	Number of Participants,	Type of Diagnostic Test,	Outcomes Sensitivity, Specificity, Positive Predictive Value,	OUADAS
Zanini et al., 2012 ⁵⁸	Number of Participants: 393	Type of Diagnostic Test: tTG IgA Cut-off value: 7 U/mL	Negative Predictive Value Sensitivity: 94.5% Specificity: 76.1% Positive predictive value: 85.9 Negative predictive value: 90.1 Sensitivity: 88.1%	QUADAS QUADAS Domain 1 Consecutive or random sample: Yes Case control design avoided: Yes Inappropriate exclusions: No Biased patient Selection: High
	Number of Participants: 289	Type of Diagnostic Test: tTG IgA Cut-off value: 8 U/mL	Specificity: 92.2% Positive predictive value: 94.6 Negative predictive value: 83.3 Sensitivity: 59.1%	QUADAS Domain 2 Blinded interpretation of index test results: Unclear Prespecified test threshold: Yes Bias due to testing: Unclear
	Number of Participants: 263	Type of Diagnostic Test: tTG IgA Cut-off value: 80 U/mL	Specificity: 43.1% Positive predictive value: 100 Negative predictive value: 52.9	QUADAS Domain 3 Valid reference standard: Yes Blinded analysis of reference test: Unclear Bias due to reference test: Unclear QUADAS Domain 4 Appropriate interval between reference and index test: Yes All patients received reference test: Yes All patients included analysis: Yes Could patient flow have introduced bias: Not Applicable

Table Notes: Au/ml – Absorbance Units per Milliliter; DGP – Deamidated Gliadin Peptide (DGP); DM = Diabetes; EMA – Endomysial Antibodies; HLA Human Leukocyte Antigen; IgA - Immunoglobulin A; IgG - Immunoglobulin G; L – Liter; NR – Not Reported; QUADAS – Quality Assessment of Diagnostic Studies; tTG - Anti-tissue Transglutaminase; U – Units; U/mL – Units per milliliter

Appendix D. Data Abstraction Tools

- 1. Celiac Disease Abstract Screening Form
- 2. Celiac Disease Full Text Screening Tool
- **3.** Celiac Disease Data Abstraction Tool

Celiac Disease Abstract Screening Form

If the title is to be excluded, please indicate by clicking the "exclude" button below. If included, please go to the abstract screening question below.

Exclude **Clear Response** Based on this abstract, is this an include or exclude? Include Exclude Needs discussion No abstract (exclude) Background Duplicate Data [STOP] {specify ID number of which it's a duplicate} **Clear Response** Exclude reason Not English language Not human Not about celiac disease (CD) Not about diagnosis of CD or under-diagnosis of CD No original data - letter, commentary, editorial, etc. Individual case report (Less than 10)

.

Prevalence, outside U.S.

Diagnostic method outside the scope of study.

(Diagnostic methods included in the KQs are Endomysial antibodies (EmA) test, Anti-tissue Transglutaminase (tTG) test, Deamidated Gliadin Peptide (DGP) antibody, HLA typing, video capsule endoscopy, and endoscopy with biopsy)



Test processing issue (E.g., PCR vs. other method)

-

Serology Only - No comparison with biopsy

Pre dates systematic review (SR) on topic Clear Response Comment

	A
	_

Celiac Disease Full Text Screening Tool

1. Should the study have been rejected at abstract screening? If yes, please state the reason.

Yes (STOP), submit form Specify: Not English language Not human Not about celiac disease (CD) Not about diagnosis of CD or under-diagnosis of CD No original data - letter, commentary, editorial, etc. Individual case report Prevalence, outside U.S. Diagnostic method outside the scope of study. (Diagnostic methods included in the KQs are Endomysial antibodies (EmA) test, Anti-tissue Transglutaminase (tTG) test, Deamidated Gliadin Peptide (DGP) antibody, HLA typing, video capsule endoscopy, and endoscopy with biopsy) Test processing issue (E.g., PCR vs. other method) Serology Only Pre dates systematic review (SR) on topic No 2. Does the article address Key Question 4 - adverse effects of invasive methods (endoscopy with biopsy. video capsule endoscopy)? Yes (STOP), submit form - study will be included No

3. Does the article address Key Question 2 - how the accuracy of duodenal biopsy varies by MD training, method, or length of gluten ingestion?

If yes	Yes (STOP), submit form - study will be included s, indicate which one:
	Pathologist characteristics, i.e. level of experience or specific training
	Method, i.e. type or number of specimens
	Length of time ingesting gluten before diagnostic testing

	-		
ļ			

Exclude study (indicate reason)

lf Г

4. Does the article address Key Question 1 or Key Question 3 - assess the accuracy or effectiveness (e.g. outcomes) of any of the following? (check all that apply)

Endomysial antibodies (EmA) IgA test

- Anti-tissue transglutaminase (tTG) IgA test
- EmA IgG, tTG IgG, and DGP IgG tests for IgA deficient individuals
- Deamidated Gliadin Peptide (DGP) IgA Antibodies
- HLA-DQ2 or HLA-DQ8
- Video capsule endoscopy
- None of the above (STOP), submit form study will be excluded

Does not assess accuracy or effectiveness e.g. only prevalence, test processing, etc. (STOP), submit form - study will be excluded

5. If this is an accuracy study:

Are the tests compared to biopsy?



No (STOP), submit form - study will be excluded

Did **ALL** subjects undergo both the "intervention" diagnostic method and the "reference" standard (usually biospy)? (i.e. everyone who received a serological test should get biopsy too, regardless of whether the test was negative or positive)

•	Yes
-	

No (STOP), submit form - study will be excluded

6. What is the sample size?



7. Was a consecutive or random sample of patients enrolled?

•	Yes
•	No
٠	Unclear

8. Does the article present results specific to the following populations? (check all that apply)

Children, under age 24 months vs. older children & adolescents
Adults (aged 18+)
Ethnic or geographic populations (Specify)
Low socioeconomic status (SES)
InA deficient

IgA deficient

Previously negative for CD

With signs and symptoms of CD (i.e., diarrhea, nutrional deficits, etc.)

	With type 1 diabetes				
	With auto-immune disease				
	With Turner's syndrome				
	With trisomy 21/Down Syndrome				
	With family history				
	Other special population (Specify)				
	General population, no special populations				
9. D	9. Does the article need further discussion?				
•	Yes				
● Note	No es:				
•					

Celiac Disease Data Abstraction Tool

Study Design Specify the study design



Please indicate the % of female? If not reported, please write "Not reported".

Please indicate % of race/ethnicity
Asian (specify %)
Black (specify %)
Latino (specify %)
Middle Eastern (specify %)
White (specify %)
Other 1 (specify ethnicity and %)
Other 2 (specify ethnicity and %)
Other 3 (specify ethnicity and %)
Other 4 (specify ethnicity and %)
Not reported
Arms
How many arms are there?
· · · · · · · · · · · · · · · · · · ·

Arm 1

Please describe the population as best as possible.



Please indicate the sample size.



For tTG tests: threshold level in terms of X times upper limit of normal. If not reported, please indicate "Not reported"



Arm 2

Please describe the population as best as possible.



Please indicate the sample size.



For tTG tests: threshold level in terms of X times upper limit of normal. If not reported, please indicate "Not reported"



Please describe the population as best as possible.



Please indicate the sample size.



For tTG tests: threshold level in terms of X times upper limit of normal. If not reported, please indicate "Not reported"



Arm 4

Please describe the population as best as possible.



Please indicate the sample size.



For tTG tests: threshold level in terms of X times upper limit of normal. If not reported, please indicate "Not reported"



Arm 5

Please describe the population as best as possible.



Please indicate the sample size.



For tTG tests: threshold level in terms of X times upper limit of normal. If not reported, please indicate "Not reported"



Arm 6

Please describe the population as best as possible.



Please indicate the sample size.



For tTG tests: threshold level in terms of X times upper limit of normal. If not reported, please indicate "Not reported"



Arm 7

Please describe the population as best as possible.



Please indicate the sample size.



For tTG tests: threshold level in terms of X times upper limit of normal. If not reported, please indicate "Not reported"



Arm 8

Please describe the population as best as possible.



Please indicate the sample size.



For tTG tests: threshold level in terms of X times upper limit of normal. If not reported, please indicate "Not reported"



Arm 9

Please describe the population as best as possible.



Please indicate the sample size.



For tTG tests: threshold level in terms of X times upper limit of normal. If not reported, please indicate "Not reported"



Arm 10

Please describe the population as best as possible.



Please indicate the sample size.



For tTG tests: threshold level in terms of X times upper limit of normal. If not reported, please indicate "Not reported"



Arm 11

Please describe the population as best as possible.



Please indicate the sample size.



For tTG tests: threshold level in terms of X times upper limit of normal. If not reported, please indicate "Not reported"



Arm 12

Please describe the population as best as possible.



Please indicate the sample size.



For tTG tests: threshold level in terms of X times upper limit of normal. If not reported, please indicate "Not reported"



Arm 13

Please describe the population as best as possible.



Please indicate the sample size.



For tTG tests: threshold level in terms of X times upper limit of normal. If not reported, please indicate "Not reported"



Please describe the population as best as possible.





For tTG tests: threshold level in terms of X times upper limit of normal. If not reported, please indicate "Not reported"



Arm 15

Please describe the population as best as possible.



Please indicate the sample size.



For tTG tests: threshold level in terms of X times upper limit of normal. If not reported, please indicate "Not reported"



Arm 16

Please describe the population as best as possible.



Please indicate the sample size.



For tTG tests: threshold level in terms of X times upper limit of normal. If not reported, please indicate "Not reported"



Arm 17

Please describe the population as best as possible.



Please indicate the sample size.



For tTG tests: threshold level in terms of X times upper limit of normal. If not reported, please indicate "Not reported"



Arm 18

Please describe the population as best as possible.



Please indicate the sample size.


For tTG tests: threshold level in terms of X times upper limit of normal. If not reported, please indicate "Not reported"



Arm 19

Please describe the population as best as possible.



Please indicate the sample size.



For tTG tests: threshold level in terms of X times upper limit of normal. If not reported, please indicate "Not reported"



Arm 20

Please describe the population as best as possible.



Please indicate the sample size.



For tTG tests: threshold level in terms of X times upper limit of normal. If not reported, please indicate "Not reported"



Appendix E. AMSTAR Criteria

Table E-1. AMSTAR criteria

Author, Year	Q1: Design	Q2: Selection	Q3: Search	Q4: Publication Status Criterion	Q5: List Studies	Q6: Characteristics	Q7: Quality Assessed	Q8: Quality Conclusions	Q9: Methods Appropriate	Q10: Bias	Q11: COI	Score
Video Capsule Endoscopy												
Rokkas et al., 2012 ⁶³	Yes	Yes	Yes	Yes	No	Yes	No	No	Yes	No	No	6
El-Matary et al., 2009 ⁶⁴	No	Yes	Yes	Yes	No	Yes	Yes	No	Yes	No	No	6
Serologicia	l Tests				•	•				•	•	•
Giersiepe n et al., 2012 ⁶⁰	Yes	NR	No	No	No	Yes	Yes	Yes	Yes	No	No	5
Lewis et al., 2010 ⁵⁹	No	Yes	Yes	No	No	Yes	Yes	Yes	Yes	No	No	6
Rostom et al., 2005	No	Yes	No	No	No	Yes	Yes	Yes	Yes	No	No	5
Rostom et al., 2004 ²⁰	No	Yes	Yes	No	Yes	Yes	Yes	Yes	Yes	No	No	7
NICE Guideline s, 2009 ⁸	Yes	Yes	Yes	Yes	Yes	No	No	No	Yes	No	No	6
Schyum, 2013 ⁶¹	Yes	NR	Yes	NR	No	Yes	No	NA	No	No	Yes	4
Special Po	pulations											
Ford et al., 2009 ⁸⁶	No	Yes	Yes	Yes	No	Yes	No	No	Yes	Yes	No	6

Author,	Q1: Design	Q2:	Q3:	Q4: Publication	Q5: List	Q6:	Q7:	Q8: Quality	Q9: Methods	Q10:	Q11: COI	Score
Year		Selection	Search	Status Criterion	Studies	Characteristics	Quality	Conclusions	Appropriate	Bias		
							Assessed					
van der	No	Yes	Yes	No	No	Yes	Yes	Yes	Yes	No	No	6
Windt et												
al.,												
2010°′												
Other Key	Questions											
Bruins,	No	NR	No	No	No	Yes	No	No	Yes	No	No	2
2013 ⁸⁴ on												
gluten												
challenge												
Hall et	No	NR	Yes	NR	No	Yes	Yes	No	Yes	No	No	4
al., ⁶⁵ on												
adherenc												
e to GFD												
Liao et	No	Yes	No	No	No	No	No	No	Yes	No	No	2
al.,												
2010 ⁵³ on												
adverse												
events	1		1					1	1			

Table notes: AMSTAR= Assessment of Multiple Systematic Reviews; NR=Not reported.

AMSTAR - Assessment of Reporting Quality for Systematic Reviews

1. Was an 'a priori' design provided?

The research question and inclusion criteria should be established before the conduct of the review.

Note: Need to refer to a protocol, ethics approval, or pre-determined/a priori published research objectives to score a "yes."

 \Box Yes

 $\square \ No$

□ Can't answer

□ Not applicable

2. Was there duplicate study selection and data extraction?

There should be at least two independent data extractors and a consensus procedure for disagreements should be in place.

Note: 2 people do study selection, 2 people do data extraction, consensus process or one person checks the other's work.

 $\Box \ Yes$

 \square No

□ Can't answer

□ Not applicable

3. Was a comprehensive literature search performed?

At least two electronic sources should be searched. The report must include years and databases used (e.g., Central, EMBASE, and MEDLINE). Key words and/or MESH terms must be stated and where feasible the search strategy should be provided. All searches should be supplemented by consulting current contents, reviews, textbooks, specialized registers, or experts in the particular field of study, and by reviewing the references in the studies found.

Note: If at least 2 sources + one supplementary strategy used, select "yes" (Cochrane register/Central counts as 2 sources; a grey literature search counts as supplementary). □ Yes

- □ No
- □ Can't answer
- □ Not applicable

4. Was the status of publication (i.e. grey literature) used as an inclusion criterion?

The authors should state that they searched for reports regardless of their publication type. The authors should state whether or not they excluded any reports (from the systematic review), based on their publication status, language etc.

Note: If review indicates that there was a search for "grey literature" or "unpublished

literature," indicate "yes." SIGLE database, dissertations, conference proceedings, and trial registries are all considered grey for this purpose. If searching a source that contains both grey and non-grey, must specify that they were searching for grey/unpublished lit. \Box Yes

- \square Yes
- ⊔ INO = Comit on a
- \Box Can't answer \Box Not applicable
- □ Not applicable

5. Was a list of studies (included and excluded) provided?

A list of included and excluded studies should be provided.

Note: Acceptable if the excluded studies are referenced. If there is an electronic link to the list but the link is dead, select "no."

 \square Yes

 $\square \ No$

 \Box Can't answer

□ Not applicable

6. Were the characteristics of the included studies provided?

In an aggregated form such as a table, data from the original studies should be provided on the participants, interventions and outcomes. The ranges of characteristics in all the studies analyzed e.g., age, race, sex, relevant socioeconomic data, disease status, duration, severity, or other diseases should be reported.

Note: Acceptable if not in table format as long as they are described as above.

□ Yes

 $\square \ No$

- \Box Can't answer
- \square Not applicable

7. Was the scientific quality of the included studies assessed and documented? 'A priori' methods of assessment should be provided (e.g., for effectiveness studies if the

author(s) chose to include only randomized, double-blind, placebo controlled studies, or allocation concealment as inclusion criteria); for other types of studies alternative items will be relevant.

Note: Can include use of a quality scoring tool or checklist, e.g., Jadad scale, risk of bias, sensitivity analysis, etc., or a description of quality items, with some kind of result for EACH study ("low" or "high" is fine, as long as it is clear which studies scored "low" and which scored "high"; a summary score/range for all studies is not acceptable).

□ Yes
□ No
□ Can't answer
□ Not applicable

8. Was the scientific quality of the included studies used appropriately in formulating conclusions?

The results of the methodological rigor and scientific quality should be considered in the analysis and the conclusions of the review, and explicitly stated in formulating recommendations.

Note: Might say something such as "the results should be interpreted with caution due to poor quality of included studies." Cannot score "yes" for this question if scored "no" for question 7.

 $\square \ Yes$

 \square No

□ Can't answer

□ Not applicable

9. Were the methods used to combine the findings of studies appropriate? For the pooled results, a test should be done to ensure the studies were combinable, to assess their homogeneity (i.e., Chi-squared test for homogeneity, I2). If heterogeneity exists a random effects model should be used and/or the clinical appropriateness of combining should be taken into consideration (i.e., is it sensible to combine?).

Note: Indicate "yes" if they mention or describe heterogeneity, i.e., if they explain that they cannot pool because of heterogeneity/variability between interventions.

□ Yes

□ No

 \Box Can't answer

 \Box Not applicable

10. Was the likelihood of publication bias assessed?

An assessment of publication bias should include a combination of graphical aids (e.g., funnel plot, other available tests) and/or statistical tests (e.g., Egger regression test, Hedges-Olken).

Note: If no test values or funnel plot included, score "no". Score "yes" if mentions that publication bias could not be assessed because there were fewer than 10 included studies.

 \Box Yes

□ No

□ Can't answer

□ Not applicable

11. Was the conflict of interest included?

Potential sources of support should be clearly acknowledged in both the systematic review and the included studies.

Note: To get a "yes," must indicate source of funding or support for the systematic

review AND for each of the included studies.

Appendix F. Strength of Evidence for Accuracy of Serology Tests

Diagnostic Method	Population	Number of Studies	Risk of Bias	Consistency	Directness	Precision	SOE, Findings
tTG IgA	Overall (Studies mixed symptomatic and high risk)	1 prior meta- analysis of 8 studies; 1 prior meta of 12 studies, 16 new studies	Moderate	Consistent	Direct	Precise	High New meta- analysis of thresholds used in clinical practice: Sensitivity 92.5% (95% Cl: 89.7, 94.6) Specificity 97.9% (95% Cl: 96.5, 98.7)
	Abdominal symptoms	1 prior meta of 7 studies	Moderate	Consistent	Direct	Precise (Specificity)	High Sensitivity 89.0% (95% CI: 82.0, 94.0) Specificity 98.0% (95% CI: 95.0, 99.0)
	Type 1 diabetes	1 study (N = 62)	Low	NA	Direct	NA	Insufficient Sensitivity 71.0% Specificity 93.0%
	Iron deficient	1 study (N = 130)	High	NA	Direct	NA	Insufficient Sensitivity 38.0% Specificity 97.0%
	Asymptomatic – general population screening	1 study (N = 1,001)	Low	NA	Direct	NA	Low Sensitivity 100.0% Specificity 97.4%
	Children	1 prior meta of 5 point- of-care tests	Moderate	Consistent	Direct	Precise	High Sensitivity 96.4% (95% CI: 94.3, 97.9) Specificity 97.7% (95% CI: 95.8, 99.0)
EmA IgA	Overall (Studies mixed symptomatic and high risk)	3 prior SRs, 7 new studies	Moderate	Consistent	Direct	Precise (Specificity)	High New meta- analysis Sensitivity 76.6% (95% Cl: 68.7, 82.9) Specificity 99.0% (95% Cl: 98.4, 99.4)

Table F-1. Strength of evidence for the accuracy of serology tests

	Abdominal symptoms	1 prior meta of 8 studies	Moderate	Consistent	Direct	Precise (Specificity)	High Sensitivity 90.0% (95% Cl: 80.0, 95.0) Specificity 99.0% (95% Cl: 98.0, 100.0)
	Type 1 diabetes	1 study (N = 62)	Low	NA	Direct	NA	Insufficient Sensitivity 71.0% Specificity 96.0%
	Iron deficient	1 study (N = 84)	High	NA	Direct	NA	Insufficient Sensitivity 100.0% Specificity 99.0%
	Asymptomatic – general population screening	1 study (N = 1,001)	Low	NA	Direct	NA	Low Sensitivity 85.7% Specificity 99.0%
	Children	1 prior meta of 11 studies	Moderate	Consistent	Direct	Precise (Specificity)	High Sensitivity 82.6% to 100% (not pooled) Specificity 98.2% (95% CI: 96.7, 99.1)
DGP IgA	Overall (Studies mixed symptomatic and high risk)	1 prior meta of 11 studies, 2 new studies	Moderate	Consistent	Direct	Precise	High Sensitivity 87.8% (95% CI: 85.6, 89.9) Specificity 94.1% (95% CI: 92.5, 95.5) 2 new studies report increased sensitivity
	Children	1 prior meta of 3 studies	Moderate	Consistent	Direct	Precise (Specificity)	Moderate Sensitivity 80.7% to 95.1% (not pooled) Specificity 90.7% (95% CI: 87.8, 93.1)
DGP lgG	Overall (Studies mixed symptomatic and high risk)	1 prior SR of 7 studies, 1 new study	Moderate	Consistent	Direct	Precise (Specificity)	Moderate Sensitivity 75.4% to 97.0% Specificity 90.7% to 100%

	Children	1 prior SR of 3 studies, 2 new studies	High	Consistent	Direct	Imprecise	Moderate Sensitivity 80.1% to 100% Specificity 86.0% to 100% New study reports sensitivity and specificity of 100% in children under 7 years of age
DGP IgA + IgG combo	IgA deficient	1 study (N = 45)	Moderate	NA	Direct	NA	Insufficient Sensitivity 100% Specificity 89.2%
tTG IgA + DPG IgA	Symptomatic adults	1 study (N=998)	Low	NA	Direct	NA	Insufficient Sensitivity 72.2% Specificity 97.4%
	Children	1 study of 2 threshold levels (N= 703)	High	Consistent	Direct	NA	Insufficient Sensitivity 94.5% to 96.7% Specificity 89.8% to 95.7%
tTG IgA + DGP IgA/IgG combo	Symptomatic adults	1 study of 2 threshold levels (N = 176)	Moderate	Consistent	Direct	NA	Insufficient Sensitivity 85.0% to 91.0% Specificity 80.0% to 98.0%
tTG IgA + EmA IgA	Symptomatic	2 studies (N = 443)	Moderate	Consistent	Direct	Imprecise	Insufficient Sensitivity 76.0% to 87.0% Specificity 85.0% to 97.0%
	First degree relatives	1 study (N = 32)	Low	NA	Direct	NA	Insufficient Sensitivity 81.0% Specificity 70.%
	Type 1 diabetes	1 study (N = 60)	Low	NA	Direct	NA	Insufficient Sensitivity 93.0% Specificity 64.0%
	Mixed (symptomatic and / or Type 1 diabetes)	1 study (N = 752)	High	NA	Direct	NA	Insufficient Sensitivity 83.0% Specificity 99.0%